WORK SUMMARY AND DATA REPORT

COLLECTION OF BATS FROM THE HUDSON RIVER, NEW YORK 2001 AND 2002 SAMPLES

HUDSON RIVER NATURAL RESOURCE DAMAGE ASSESSMENT

HUDSON RIVER NATURAL RESOURCE TRUSTEES

STATE OF NEW YORK

U.S. DEPARTMENT OF COMMERCE

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1305 East-West Highway, Rm 10219
Silver Spring, MD 20910-3281

**Names of certain individuals and affiliations have been removed to maintain confidentiality*

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EXECUTIVE SUMMARY

Natural resources of the Hudson River have been contaminated through past and ongoing discharges of polychlorinated biphenyls (PCBs). The Hudson River Natural Resource Trustees – New York State, the U.S. Department of Commerce, and the U.S. Department of the Interior – are conducting a natural resource damage assessment (NRDA) to assess and restore those natural resources injured by PCBs. This Work Summary and Data Report documents a screening level survey of PCB contamination of select Hudson River bat species conducted pursuant to the NRDA.

In July 2001 bats were collected from along the Hudson River at the Saratoga National Historic Park and from reference sites in New York State. A second collection of bats from the Hudson River in July 2002 supplemented the 2001 collection, and entailed collection of bats from three other sites along the Upper Hudson River.

Thirty-one of the bats collected in 2001 (26 little brown bats and 5 big brown bats) were selected by the Trustees for sample preparation and analysis. The bats were dissected and the brains analyzed for PCB congeners, PCB homologue groups, and total PCBs. Percent lipids and percent moisture were not analyzed due to limited sample size.

The results show that Hudson River bats have been exposed to PCBs. Total PCBs in the brains of big brown bat females range from 31.7-642 ppb. Total PCBs in the brains of little brown bat males and females range from 1,760-2,390 ppb and 274-1,820 ppb, respectively. By comparison, total PCBs in brains of little brown bat females from reference areas range from non-detect (ND)-662 ppb.

1.0 INTRODUCTION

Natural resources of the Hudson River have been contaminated through past and ongoing discharges of polychlorinated biphenyls (PCBs). The Hudson River Natural Resource Trustees – New York State, the U.S. Department of Commerce, and the U.S. Department of the Interior – are conducting a natural resource damage assessment (NRDA) to assess and restore those natural resources injured by PCBs (Hudson River Natural Resource Trustees 2002a). This Work Summary and Data Report documents a screening level survey of PCB contamination of select Hudson River bat species conducted pursuant to the NRDA.

The Hudson River and surrounding area support nine species of bats, as follows: big brown bat, *Eptesicus fuscus*; silver-haired bat, *Lasionycteris noctivagans*; eastern red bat, *Lasiurus borealis*; hoary bat, *Lasiurus cinereus*; small-footed bat, *Myotis leibii* (a NYS species of special concern); little brown bat, *Myotis lucifugus*; northern long-eared bat, *Myotis septentrionalis*; Indiana bat, *Myotis sodalis* (a State and Federally listed endangered species); and, eastern pipistrelle, *Pipistrellus subflavus*.

Bats that reside in the Hudson River Valley may be exposed to PCBs through the food web and may be at risk from that exposure. Bats may be particularly at risk from mobilization of stored organochlorines, including PCBs, because of fat cycles associated with migration and hibernation (Clark and Shore 2001). The long life span of bats (some as long as 30 years) also allows more time for contact with, and accumulation of, contaminants. Similarly, relatively low reproductive rates (generally 1-2 young per year) can result in slow recovery for affected bat populations (Clark and Shore 2001, Harvey 1992).

The Trustees conducted this preliminary investigation to assess the extent and severity of PCB exposure in bats. The results of this preliminary investigation will allow the Trustees to determine whether bats have been exposed to PCBs and evaluate the possible effect these PCBs may have on bat health. The findings of this preliminary investigation will inform the Trustees regarding the need for future injury determination studies of bats.

The objective of this investigation was to collect samples of bats from the Hudson River for PCB analysis. In July 2001 the New York State Department of Environmental Conservation (NYSDEC) collected bats from along the Hudson River at the Saratoga National Historic Park (NHP) and from reference sites in conjunction with preliminary testing of the utility of the Anabat bat detectors in identifying free ranging bats in New York State (NYSDEC 2001). A second collection of bats from the Hudson River, gathered in July 2002, supplemented the 2001 collection, and entailed collection of bats from three other sites along the Hudson River. This preliminary investigation focused on analysis of samples from little brown bats and big brown bats collected in 2001.

2.0 BACKGROUND

Bats such as little brown bats and big brown bats are potential bioindicators of contamination due to their consumption of emergent aquatic insects. These insects spend the majority of their life cycle buried in or on river sediments, including those of the Hudson River. The Trustees have sampled emergent aquatic insects from some of the most highly contaminated reaches of the Hudson River and found those insect samples to contain in excess of 5.7 parts per million (ppm) total PCB wet weight (NYSDEC 1998).

Bats that utilize aquatic insects as prey items consume these at rates that vary between about 1.4 grams per hour (g/hr) to 3.5 g/hr when feeding (van Zyll de Jong 1985). Bats therefore have potentially high rates of PCB intake. Bats begin feeding soon after sunset and can consume insect prey each night at a rate which equals from between 30 to 50 percent of their body weight (Canadian Wildlife Service 2002).

Little brown bats *(Myotis lucifugus)*, which are prevalent along the Hudson River, feed upon Diptera, Ephemeroptera, Tricoptera, Lepidoptera, Isoptera and Coleoptera with midges, mayflies and caddisflies predominating in the diet. Big brown bats (*Eptesicus fuscus*), which are also prevalent along the Hudson River, feed upon Coleoptera, Hemiptera, Diptera, Plecoptera, and Lepidoptera (van Zyll de Jong 1985). Both little brown and big brown bats spend the winter in hibernacula. Adult bats live separately during the summer feeding season, the females living in maternity colonies, separate from adult males, where they rear their young by day and feed at night.

The State and Federally endangered Indiana bat hibernates in the Hudson River-New York Bight watershed. In addition, NYSDEC has confirmed several maternity colonies of Indiana bats located within the Hudson River Valley. Indiana bats feed on flying insects, and bats roosting in proximity to the river could potentially be exposed to PCBs through the consumption of insects with aquatic larval stages.

3.0 BAT SAMPLE COLLECTION, PROCESSING AND ANALYSIS

3.1 COLLECTION, PROCESSING AND ANALYSIS OF 2001 SAMPLES

The 2001 Hudson River bat collection effort took place July 23-24, 2001. Bat collection was conducted as part of a NYSDEC project testing the utility of the Anabat bat detector in identifying bat species, and was performed as specified in "Preliminary Testing of the Utility of the Anabat Bat Detectors in Identifying Free Ranging Bats in New York State" (NYSDEC 2001). Bats were collected from along the Hudson River at the Saratoga NHP and from five reference sites in New York State: Black River, White Birch Pond, Lower Cascade Lake, Chapel Pond, and Rome.

The teams of investigators assembled at the sites before dusk to assemble the acoustic detection equipment used to identify the selected bats and to acquaint the teams with duties and safety operations associated with the use of firearms and water safety. Upon dusk and the active feeding of the targeted species of bats, the bats were identified by their acoustic vocalizations, sighted in spotlights and shot using appropriate shot (lead shot was generally used). The bats were retrieved by boat by investigation team members after cessation of shooting. Bats that had fallen into the water were patted dry with paper towels before being placed individually into sample containers. The location of each site from which bats were collected was recorded with Global Positioning System (GPS) equipment.

Each bat collected was given a unique identification number. Collection activities in 2001 were documented through use of NY Anabat Test Study Site Data Sheets (NYSDEC 2001) providing the identification number, species, sex, and time and location of collection. Location was specified with both a written description and GPS coordinates.

The bats that were collected on the Hudson River were placed into individual chemically-clean glass jars. Bats collected at locations other than the Hudson River were wrapped in paper towels and then placed in plastic bags. The jars and bags containing bats were placed into coolers and stored on ice before the samples were frozen by NYSDEC personnel. The frozen samples were delivered to NYSDEC personnel on August 28, 2001, for storage at the NYSDEC Hale Creek Field Station. At the Hale Creek Field Station, the jars were placed into storage at -20 degrees C, in a secure walk-in freezer.

A total of 75 bats of 6 species were collected from the Hudson River and other areas in 2001, as follows:

Site	Little brown bat (*Myotis lucifugus*)	Big brown bat (*Eptesicus fuscus*)	Silver-haired bat (*Lasionycteris noctivagans*)	Red bat (*Lasiurus borealis*)	Hoary bat (*Lasiurus cinereus*)	Eastern pipistrelle (*Pipistrellus subflavus*)
Saratoga NHP	14	5	0	0	1	8
Black River	14	0	0	1	4	0
White Birch Pond	14	0	0	0	0	0
Lower Cascade Lake	5	0	2	0	0	0
Chapel Pond	1	0	0	0	0	0
Rome	0	3	0	1	1	1
Totals	48	8	2	2	6	9

Of the total of 75 bats collected by NYSDEC in 2001, 31 were made available to the Trustees for use in the Hudson River NRDA. The remaining bats were provided to the New York State Museum. Considerations as to the disposition of the specimens included the location from which the specimen was collected, the condition of the specimen, the availability of specimens of a given species, and the best use of the specimen.

The outcome of the 2001 field effort is further discussed in the "Field test of the ability of the Anabat system to quantitatively identify free-flying bats" (Britzke and Hicks 2001).

As noted above, 31 of the bats collected in 2001 were made available to the Trustees for use in the Hudson River NRDA. The Trustees selected 14 little brown bats (11 females and 3 males) and the 5 big brown bats (all females) from the Saratoga NHP for sample preparation and analysis. The Trustees selected these species because samples were available from both Saratoga NHP and multiple reference areas. For the reference areas, the Trustees selected female little brown bats from White Birch Pond and Black River (n = 12); these bats were selected for their comparability to the Hudson River bats in terms of species and sex. The locations of the three sample collection sites are shown in Hudson River Natural Resource Trustees 2004a (slide 25).

A total of 26 little brown bats and 5 big brown bats were thus selected by the Trustees for sample preparation and analysis. These 31 bats were shipped under chain-of-custody (COC) from the NYSDEC Hale Creek Field Station to the Trustees' processing laboratory. The bats were shipped by NYSDEC on November 12, 2002, and received by the processing laboratory on November 14, 2002.

At the processing laboratory the bats were dissected in accordance with the Trustees' Standard Operating Procedure (SOP) "Final Protocol for Handling and Dissection of Bat Samples Collected from the Vicinity of the Hudson River, New York and from Reference Areas in New York State," dated November 13, 2002 (see Appendix A).

The bats were dissected and separated into the following four components: brains; stomach contents; carcass; and, carcass remainders (consisting of head, feet, wings, skin with pelage and gastrointestinal tracts minus the stomach contents). Two wipe blanks also were collected during the dissection process.

The carcass samples (n=31), stomach content samples (n=31), brain samples (n=31), and wipe blanks (n=2) were shipped under COC from the processing laboratory to the Trustees' analytical laboratory on November 19, 2002. The analytical laboratory received the coolers containing these samples on November 20, 2002. The brain samples and wipe blanks were subsequently submitted for chemical analysis for PCBs in accordance with the Trustees Analytical Quality Assurance Plan (QAP) (Hudson River Natural Resource Trustees 2002b). The samples were analyzed for PCB congeners, PCB homologue groups, and total PCBs. Percent lipids and percent moisture were not analyzed due to limited sample size. The carcass samples and the stomach content samples were not analyzed and were archived.

3.2 Collection of 2002 Samples

The 2002 Hudson River bat collection was conducted on July 15-16, 2002. Bat collection in 2002 was conducted as further work by NYSDEC testing the Anabat bat detector in identifying bat species. Bat collection was conducted as specified in "Preliminary Testing of the Utility of the Anabat Bat Detectors in Identifying Free Ranging Bats in New York State" (NYSDEC 2001). Bat collection in 2002 was limited to sites on the Hudson River. One change from the 2001 sampling was that non-lead shot was used in the 2002 collection.

Collection activities in 2002 were documented through use of sequentially numbered Bat Collection Data Sheets (Appendix B). The data sheets were placed into a ring-binder. All information was recorded on these preformatted data sheets.

In 2002, bats were collected from three areas identified in advance, based on known contamination, habitat considerations, and the likelihood of finding bats located along the Hudson River between Fort Edward, New York and Schodack Island, New York.

The three areas that were the subject of the 2002 bat collection effort are as follows:

- Site 1: a New York State Department of Transportation dredge spoil disposal site along the west bank of the Hudson River, south of Roger's Island, in the Town of Moreau, known as Special Area 13;
- Site 2: near Site 8 from the "Floodplain Soil and Biota Screening Sampling Report" (SEA Consultants, Inc. 2002); and,
- Site 3: near Site 4 from the "Floodplain Soil and Biota Screening Sampling Report" (SEA Consultants, Inc. 2002).

A waterproof label was attached directly to each jar containing a bat. The label was marked in waterproof ink in the following format:

Bat number: ST-SPECIES-NUM, such as 01-BB-01, where:

ST = Site Number 1, 2, 3 or 4, designated respectively as 01, 02, 03 or 04;

SPECIES = BB for big brown bat or LB for little brown bat; and,

NUM = the specimen number from 01 through 05.

Date/time collection: _____

Sample collector name and initials: _____

A total of 15 bats of 1 species were collected from the Hudson River in 2002, as follows:

Site	Little brown bat (*Myotis lucifugus*)
Site 1 (Special Area 13)	5
Site 2 (Floodplain Site #8)	5
Site 3 (Floodplain Site #4)	5

The jars containing these bats were placed into a cooler and stored on ice for transport to the NYSDEC Hale Creek Field Station. At the Hale Creek Field Station, the jars were placed into storage at -20 degrees C, in a secure walk-in freezer. No preparation or analysis has been done on the 2002 bat samples. All 15 of the bats collected in 2002 have been archived by the Trustees for potential future use in the Hudson River NRDA and remain in secure storage at the NYSDEC Hale Creek Field Station.

3.3 CHAIN-OF-CUSTODY

Bat samples for 2001 and 2002 are maintained under Chain-of-Custody. Any transfer of custody has been, and will be, documented on a Chain-of-Custody record (see Appendix C).

3.4 QUALITY ASSURANCE/QUALITY CONTROL

In July 2002 the Trustees finalized an Analytical QAP (Hudson River Natural Resource Trustees 2002b). In September 2002, the Trustees issued a Quality Assurance Management Plan (as part of the NRDA Plan) (Hudson River Natural Resource Trustees 2002a).

The bat collections in July 2001 and July 2002 thus preceded the Trustees' Analytical QA Plan and the QA Management Plan. The processing and analysis of the 2001 samples, however, post-dates these two plans. Accordingly, the processing and analysis of the 2001 samples was conducted in compliance with the requirements of the QA Management Plan and the Analytical QA Plan.

The laboratories chosen for preparation (dissection, removal of stomach contents, etc.) of the 2001 samples, and for analysis of those samples followed the requirements of the Hudson River QA Management Plan and the Hudson River NRDA Analytical QA Plan.

The original data sheets have been retained at New York State Department of Environmental Conservation, 625 Broadway, Albany, NY 12233, and will be maintained and archived for a minimum of fifteen years. Any disposal of the data sheets will be coordinated with the DOI and NOAA after this timeframe unless a longer archive period is requested.

Data validation was conducted by the Trustees and was based on the quality assurance/quality control (QA/QC) criteria documented in the Trustees' Analytical Quality Assurance Plan for the Hudson River Natural Resource Damage Assessment (Hudson River Natural Resource Trustees 2002b), USEPA (1999), and the following SOPs from the laboratory:

- SOP # HR NRDA Project Tissue Prep: Tissue Preparation and Homogenization, Revision #1.0, 9/25/02;
- SOP # OP-016: Microscale Solvent Extraction (MSE), Revision #1.0, 1/29/04;
- SOP # O-010: Determination of PCB Homologues and Individual Congeners by GC/MS - SIM, Revision # 2.2, 10/24/02; and,
- Additional cleanup, sample handling, storage, custody SOPs as necessary.

The data packages submitted by the laboratory were reviewed to determine whether the analytical data quality objectives (DQOs) specified in the Analytical QAP were met.

Table 1.1 of the Trustees' Analytical QAP specifies the target Method Detection Limits (MDLs) for PCB congeners, homologues and total PCBs. For tissue, such as bat brains, the target MDLs are 0.1 ug/kg wet weight (equivalent to 0.1 ng/g or 0.1 ppb wet weight) for individual congeners, and 10 ug/kg (equivalent to 10 ng/g or 10 ppb) for PCB homologues and total PCBs. Because of the small size of the individual bat brains, achievable detection limits were higher than the QAP target limits. Actual MDLs are reported on the Bat Brain Tissue Data Sheets (Appendix D) in the "Detection Limit" column.

Appendix D contains the Data Quality Assessment Report (Hudson River Natural Resource Trustees 2004b) for the bat brain tissue. Table 1A to that appendix is a summary of standard reference material (SRM) analytical results for each sample delivery group (SDG) that was analyzed. A statistical evaluation of the SRM analytical results is found in Table 1B of that appendix.

Out of 1,798 individual analytical results reported by the laboratory (31 samples each with 47 congeners, ten homologue groups, and total PCBs), a total of 66 (3.7%) data points were qualified as estimated (J) because of laboratory accuracy outliers. No data were rejected. For all data, the overall quality of the data is acceptable and the results, as qualified, are considered usable. The completeness level attained for the analysis of the field samples is 100%.

A database containing the analytical results and related quality control results is available upon request (Hudson River Natural Resource Trustees 2004c).

4.0 METHODS

The Bat Brain Tissue Data Sheets (Appendix E) provide the results of the analyses. These Data Sheets contain information that has been extracted from the Trustees' Bat Brain Tissue Database (Hudson River Natural Resource Trustees 2004c). That complete database and the accompanying Database User Manual (Hudson River Natural Resource Trustees 2004d) are not included in this report due to the size of the database, but will be made available upon request.

The Bat Brain Tissue Data Sheets contain the following fields:

Sampling Date – Sampling Date (mm/dd/yy format).

Field ID – The field IDs were created using the following format:

SSSSS NN

where SSSSS represents the site code and NN is the numerical record of animals collected (e.g., "5" indicates the fifth animal collected. For example, Sarato 11 indicates the 11th specimen collected at the Saratoga NHP site.

Latitude – Decimal latitude readings (NAD83) as reported by the GPS system.

Longitude – Decimal longitude readings (NAD83) as reported by the GPS system.

Common Name – The common name of the animal (e.g. little brown bat).

Analyte – For the PCB congeners, the analyte names are reported using the following format:

Clx-BZ#NNN

where Clx refers to the chlorination level, BZ# refers to the Ballschmiter and Zell (1994) number, and NNN is the congener number. For example, PCB110 (a pentachlorinated biphenyl) is reported as Cl5-BZ#110.

The total concentration of all congeners within a chlorination level (including both target and non-target congeners) is represented by the chlorination level name. For example, the total of all pentachlorinated biphenyls is reported as Pentachlorobiphenyls.

Value, Interpretive Qualifier, and Units –

Value – Analytical result (3 significant figures).

Interpretive Qualifier – This field contains qualifiers applied to each data point after the data validation process. Data validation qualifiers were assigned to data points when associated QC sample results indicated the data did not meet the data quality objectives. The following definitions provide brief explanations of the qualifiers applied to the Hudson River NRDA data. Reasons for qualifications are explained further in the Data Quality Assessment Report (Appendix D).

U Not detected. The analyte was not detected. The associated value represents the detection limit.

J Estimated. The associated numerical value is an estimated quantity. The analyte was detected, but the reported value may not be accurate or precise. The "J" qualification indicates the data fell outside the QC limits, but the exceedance was not sufficient to cause rejection of the data, or that the reported result is within a range of elevated analytical uncertainty (greater than the Method Detection Limit (MDL) value, but less than the Practical Quantitation Limit (PQL) value).

Unit – The unit of measurement of the analytical result is provided (for example, µg/kg).

Detection Limit – 3 significant figures; this column includes units.

A brief description of some of the features of these data follows in sections 4.1 and 4.2 of this Data Report. Please note that the unit "µg/kg" used in the Data Sheets is equivalent to parts per billion (ppb) used in the discussion of these data in this Data Report.

Data fields, and data collected by the Trustees, that are not reported in this Data Report in the Bat Brain Tissue Data Sheets (found in Appendix E of this report), but that are contained within the Trustees' Bat Brain Tissue Database include the following: Site Code, Study Name, Analytical Batch Number, Laboratory Flag, Data Validation Qualifier, Data Validation Qualifier Reason Code, Analysis Group, Analytical Method, Matrix, Basis, Extraction Date, Analysis Date, Dilution Factor, Sample Size and Units, Specimen Number, Time Collected, Date Dissected, Species (scientific name), Biota Type, Sex, Age, Reproductive Status, Weight at Collection, Forearm Length, Site Name, Start Date (of collection), End Date (of collection), Location, Town, County, Universal Transverse Mercator (UTM) Northings Coordinates (Zone 18N), UTM Eastings Coordinates (Zone 18N), and Habitat.

All samples were included in calculating average PCB concentrations by location, species and sex. For "J" qualified results, the estimated value was used in the calculations.

For the purpose of reporting PCB results below and in the figures attached to this report, all values flagged with a U interpretive qualifier (that is, not detected; see Appendix D) were considered to be zero. Using zero, rather than the value reported by the laboratory for the analyte, which represents the detection limit for the analysis, potentially underreports the true value, but avoids overreporting the true value. This is thus a conservative result; the actual PCB concentration could be higher.

4.1 Total PCB Concentrations

Total PCB concentrations of bat brain samples, by location (Hudson River or Reference Areas), species and sex, are detailed in Table 1 and shown in Figures 1 and 2. Values in Table 1 are reported to three significant figures.

Table 1. Summary of Total PCBs (as sum of homologues) in Hudson River Bat Brain Tissue

Location	Species	Sex and Number (n) of Samples	Concentration Range (parts per billion (ppb))	Concentration Average ± 1 SD (ppb)
Hudson River (Saratoga NHP)	Big Brown Bats	Females (n=5)	31.7 – 642	271 ± 225
	Little Brown Bats	All (n=14)	274-2,390	1,130 ± 655
		Females (n=11)	274-1,820	889 ± 488
		Males (n=3)	1,760-2,390	2,010 ± 335
Reference Areas (White Birch Pond and Black River)	Little Brown Bats	Females (n=12)	ND-662	158 ± 173

4.2 PCB Homologues and Congeners

PCBs are synthetic (man-made) chemicals that form a group of 209 individual compounds that have similar chemical structures based on a biphenyl core with 1 to 10 chlorine atoms attached. PCBs have the generic formula $C_{12}H_{(10-x)}Cl_x$, where x is an integer from 1 to 10. Each individual PCB compound, called a congener, is identified by the unique number and location of chlorine atoms that attach to the compound's base structure. Congeners differ both in their physical properties and in their effects on fish and wildlife (Safe 1994; Van den Berg et al. 1998).

For this investigation, the bat brain tissue samples were analyzed for 48 specific target PCB congeners listed in Table 2. In addition, a total concentration for each homologue group was determined by summing all target and non-target congener concentrations within each homologue group. For any congener reported as non-detected, zero was used in the summation.

Table 2. PCB Congeners for which Bat Brain Tissues were Analyzed.

Current Ballschmiter and Zell (1994) and IUPAC Number	IUPAC Name
8	2,4'-Dichlorobiphenyl
18	2,2',5-Trichlorobiphenyl
28/31	2,4,4'-Trichlorobiphenyl/2,4',5-Trichlorobiphenyl
44	2,2',3,5'-Tetrachlorobiphenyl
45	2,2',3,6-Tetrachlorobiphenyl
47	2,2',4,4'-Tetrachlorobiphenyl
49	2,2',4,5'-Tetrachlorobiphenyl
52	2,2',5,5'-Tetrachlorobiphenyl
56	2,3,3',4'-Tetrachlorobiphenyl
66	2,3',4,4'-Tetrachlorobiphenyl
70	2,3',4',5-Tetrachlorobiphenyl
74	2,4,4',5-Tetrachlorobiphenyl
77	3,3',4,4'-Tetrachlorobiphenyl
81	3,4,4',5-Tetrachlorobiphenyl
87	2,2',3,4,5'-Pentachlorobiphenyl
95	2,2',3,5',6-Pentachlorobiphenyl
99	2,2',4,4',5-Pentachlorobiphenyl
101	2,2',4,5,5'-Pentachlorobiphenyl
105	2,3,3',4,4'-Pentachlorobiphenyl
110	2,3,3',4',6-Pentachlorobiphenyl
114	2,3,4,4',5-Pentachlorobiphenyl
118	2,3',4,4',5-Pentachlorobiphenyl
123	2,3',4,4',5'-Pentachlorobiphenyl
126	3,3',4,4',5-Pentachlorobiphenyl
128	2,2',3,3',4,4'-Hexachlorobiphenyl
138	2,2',3,4,4',5'-Hexachlorobiphenyl
146	2,2',3,4',5,5'-Hexachlorobiphenyl
149	2,2',3,4',5',6-Hexachlorobiphenyl
151	2,2',3,5,5',6-Hexachlorobiphenyl
153	2,2',4,4',5,5'-Hexachlorobiphenyl
156	2,3,3',4,4',5-Hexachlorobiphenyl
157	2,3,3',4,4',5'-Hexachlorobiphenyl
158	2,3,3',4,4',6-Hexachlorobiphenyl
167	2,3',4,4',5,5'-Hexachlorobiphenyl
169	3,3',4,4',5,5'-Hexachlorobiphenyl
170	2,2',3,3',4,4',5-Heptachlorobiphenyl
174	2,2',3,3',4,5,6'-Heptachlorobiphenyl

Current Ballschmiter and Zell (1994) and IUPAC Number	IUPAC Name
177	2,2',3,3',4,5',6'-Heptachlorobiphenyl
180	2,2',3,4,4',5,5'-Heptachlorobiphenyl
183	2,2',3,4,4',5',6-Heptachlorobiphenyl
187	2,2',3,4',5,5',6-Heptachlorobiphenyl
189	2,3,3',4,4',5,5'-Heptachlorobiphenyl
194	2,2',3,3',4,4',5,5'-Octachlorobiphenyl
195	2,2',3,3',4,4',5,6-Octachlorobiphenyl
201	2,2',3,3',4,5',6,6'-Octachlorobiphenyl
206	2,2',3,3',4,4',5,5',6-Nonachlorobiphenyl
209	Decachlorobiphenyl

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FIGURES

Figure 1. PCB Concentrations in Bat Brains by Species and Location

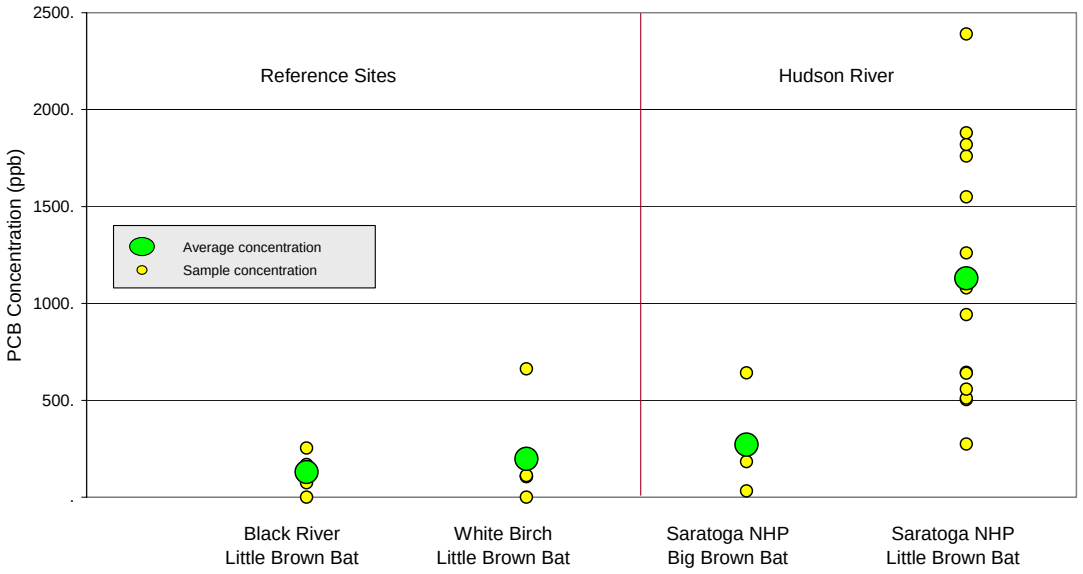
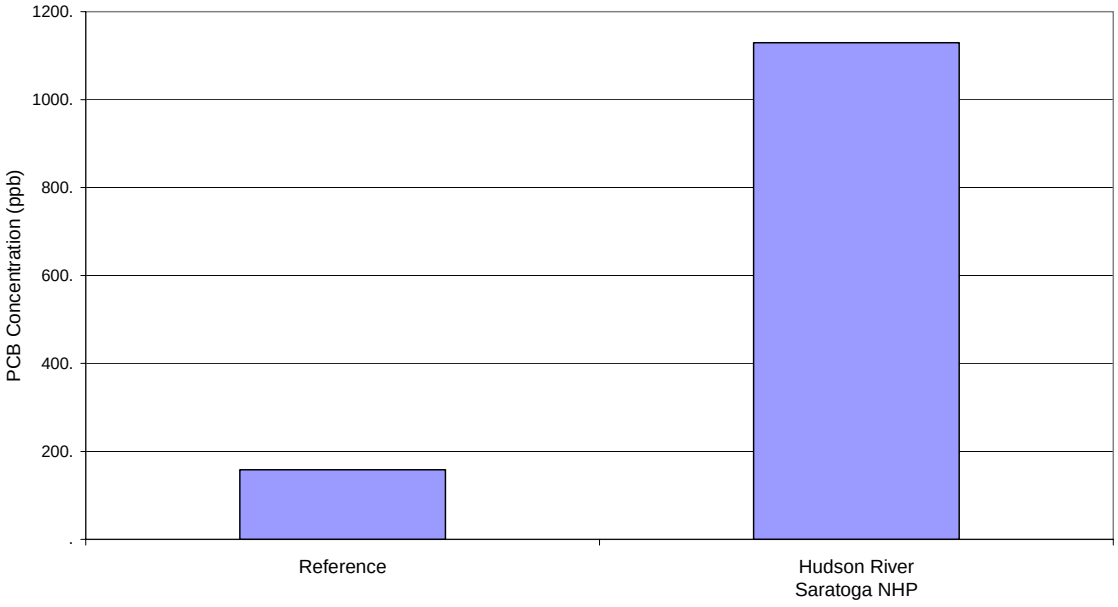


Figure 2. Average PCB Concentrations in Little Brown Bat Brains



Appendix A

Final Protocol for Handling and Dissection of Bat Samples Collected from the Vicinity of the Hudson River, New York and from Reference Areas in New York State

DISSECTION TEAM ACKNOWLEDGEMENT OF REVIEW OF FINAL PROTOCOL FOR BAT HANDLING AND DISSECTION

By my signature, I acknowledge that I have read and understand the "Final Protocol for Handling and Dissection of Bat Samples from the Vicinity of the Hudson River, New York, and from Reference Areas in New York State," including the Appendices. I will follow this protocol.

Name (printed): _____ Name (printed): _____

Signature: _____ Signature: _____

Date: _____ Date: _____

Name (printed): _____ Name (printed): _____

Signature: _____ Signature: _____

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PROTOCOL FOR BAT HANDLING AND DISSECTION, HUDSON RIVER STUDY

A. Background

A total of 31 bats collected in 2001 will be dissected in preparation for analysis for polychlorinated biphenyls (PCBs): 19 bats from the Hudson River study site and 12 bats from a reference area. The sample of bats from the Hudson River study site consists of 14 little brown bats (*Myotis lucifugus* – 2 adult males, 1 young-of-the-year male, and 11 adult females) and 5 big brown bats (*Eptesicus fuscus* – all adult females). The reference sample of bats consists of 12 adult female little brown bats from White Birch Pond and the Black River. Bats are currently frozen and stored at the New York State Department of Environmental Conservation Hale Creek laboratory. Details on collecting localities and conditions are available from this source.

There is a moderate amount of information in the literature on organochlorine residue concentrations in these two species from various locations in the United States, as well as results from a few experimental feeding studies (see Clark and Shore 2001, Clark 1981 for overviews). This information includes likely lethal residue concentrations of some contaminants in brains (e.g. Clark and Stafford 1981) as well as knowledge of residue relationships among some body components and potential impacts on reproduction based on dietary concentrations. However, the amount of information available on PCBs in bats (particularly specific congeners—Fernandez et al. 1993) is more limited than that available for insecticides such as dieldrin and DDT or their metabolites.

To obtain some basic preliminary information on PCBs from the Hudson River study area for comparison with the literature, carcass dissections will be carried out by technicians from Colorado State University (CSU) under the supervision of Thomas J. O'Shea of the U.S. Geological Survey (USGS). Technicians will be advanced veterinary students or recent graduates in biology who have had pre-exposure immunizations to rabies (freezing specimens does not kill the rabies virus), and who also have experience in dissecting bat carcasses for joint disease studies carried out by CSU and USGS in Fort Collins. The analytical laboratory will determine the risk and need for immunizations of analytical chemistry laboratory personnel. Extraction solvents will kill the rabies virus and subsequent handling will not require these health precautions (D.R. Clark Jr., in litt; Clark et al. 1975, Clark and Prouty 1976).

Tissues that will be submitted for chemical analysis are brains, carcasses, and stomach contents from each bat. Carcass remainders will be saved frozen and returned as directed by the Trustee representatives for possible future analyses for calculations of total body loads. It is important to note that the masses of brain samples will be small (brain tissue may not be available from all specimens, depending on degree of post-mortem decomposition), and not all stomachs will be full. This may require lower levels of analytical sensitivity for brain samples, and possibly for some stomach contents samples. It is also important to note that for maximum ability to make comparisons with the literature, reporting of analytical results by the chemistry laboratory must include percent lipid and percent moisture values for each sample. Data such as wet weight of sample or aliquot analyzed, weight of lipids extracted from the sample or aliquot, and total µg of compound detected should be available for each sample submitted for organochlorine analysis.

FINAL PROTOCOL FOR HANDLING AND DISSECTION OF BAT SAMPLES COLLECTED FROM THE VICINITY OF THE HUDSON RIVER, NEW YORK AND FROM REFERENCE AREAS IN NEW YORK STATE

HUDSON RIVER NATURAL RESOURCE DAMAGE ASSESSMENT

_____

Dissection Team Leader

_____

Quality Assurance Coordinator

November 13, 2002

B. DISSECTION METHODS

See also Appendix II for step-by-step dissection instructions.

Dissection tools and surfaces. Dissections will be carried out using standard stainless steel instruments for small mammals: dissection scissors, scalpels with disposable blades, forceps, and spatulas. Stainless steel dissecting instruments will be washed in a 1% solution of deionized water and Alconox detergent (lot A2 H2, Alconox Inc), rinsed in acetone, followed by a rinse in hexane, before each animal is dissected, and before each sample type (carcass, brain, stomach contents, carcass remainder) is prepared within individual animals. Alconox, acetone and hexane have been supplied in sealed containers through VWR Inc. and have been stored at USGS in Fort Collins. Both acetone (Burdick and Jackson, lot CD 935) and hexane (Burdick and Jackson, lot CF765) are capillary GC/GC-MS solvent grade suitable for trace analysis at ppb levels or less. During dissections solvents will be stored in 500 ml Teflon FEP wash bottles (Nalge Nunc. Inc. lot 465201)

All dissections will take place under a laboratory fume hood at the U.S. Geological Survey laboratory facility in Fort Collins. All contact of specimens with synthetic surfaces, plastics, soaps, or oils will be avoided. Laboratory bench surfaces will be wiped clean with acetone and hexane, and covered with aluminum foil obtained in sealed packages from commercial retail outlets. Aluminum foil in work areas will be changed and replaced with clean foil after each bat is dissected, and clean foil pieces will be used for dissections of individual bat subsamples. Dissectors will wear latex gloves, but will handle samples destined for PCB analysis only using stainless steel dissection tools washed in Alconox and rinsed in acetone and hexane. Latex gloves will be discarded and changed between bat samples.

Sequences of dissection. Technicians will dissect and process each bat separately, singly, and completely before another bat is processed. Each frozen bat will be thawed singly, dissections completed, samples placed in chemically clean glass sample containers, mass of samples determined and recorded, and samples returned to the freezer before another bat is processed by the same technician. Samples will be processed in two separate lots (reference area and Hudson River) to avoid potential cross-contamination, with reference area bats sampled first. Within each species, samples will be prepared beginning with individuals least likely to be contaminated (adult females will be dissected first, followed by young of the year, and then adult males).

Dissection procedures. Each bat will be thawed individually and separately at room temperature. Prior to dissection, technicians will verify sex and age class (adult or juvenile based on closure of phalangeal epiphyses—Anthony 1988) determinations made in the field in New York. Reproductive status (pregnant, lactating, non-reproductive, testes scrotal or non-scrotal—Racey 1988) will also be noted. Body mass will be recorded to the nearest 0.001 g on a Sartorius Model CP225D digital balance with internal calibration weights, and forearm lengths measured to the nearest 0.1 mm with dial calipers. This information will be recorded on data sheets (Appendix III) in permanent ink using field identification numbers originally assigned in New York. The Sartorius model CP225D balance has 220 g capacity and 0.1 mg precision.

The following subsamples will be collected, as described below and in Appendix II: carcass, brain, stomach contents, carcass remainders, and wipe blanks.

1. Carcasses. Carcasses will consist of the entire body minus the head, feet, wings, skin with pelage, and gastrointestinal tracts (total of these minus the stomach contents = carcass remainders, see item 4 below), following procedures established for contaminant analysis of bats by the Patuxent Wildlife Research Center (e.g. Clark et al. 1975, Clark and Lamont 1976, Clark and Prouty 1976, Clark and Stafford 1981, O'Shea et al. 2001). Dissections will be made using stainless steel dissection tools washed in the Alconox solution and rinsed in acetone followed by hexane. The head will be separated at the first cervical vertebrae. Major head musculature will be trimmed and placed with the carcass. This is the procedure followed by most of the published literature on contaminants in bats (see references in Clark and Shore 2001). Each carcass will be placed individually in chemically clean glass sample jars. Each jar will be tared on the Sartorius balance prior to addition of the sample. Mass of the sample will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (CA for carcasses), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

2. Brains. The dorsal and lateral skin of the head will be removed using stainless steel scissors or scalpels. The cranium will be opened by removal of the occipital region and portions of the parietal regions using stainless steel dissection scissors washed in Alconox and rinsed in acetone and hexane immediately prior to opening. The brain will be removed entire using stainless steel forceps or stainless steel spatulae, also washed in Alconox and rinsed in acetone and hexane immediately prior to use. The brain will be placed in a chemically clean glass sample jar. Each jar will be tared on the Sartorius balance prior to addition of the sample. The sample mass will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (BR for brains), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

3. Stomach contents. The stomach will be opened using stainless steel dissection scissors washed in Alconox solution and rinsed in acetone and hexane immediately prior to opening. Contents will be removed using stainless steel forceps or stainless steel spatulae, also washed in Alconox detergent and rinsed in acetone and hexane immediately prior to use. The stomach contents will be placed in chemically clean glass sample jars. The stomach and gastrointestinal tract tissues will be retained for inclusion with the other carcass remainders (see 4. below). Each jar will be tared on the Sartorius balance prior to addition of the sample. The sample mass will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (ST for stomach contents), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

4. Carcass remainders. Carcass remainders **(**head, feet, wings, skin with pelage and gastrointestinal tracts minus the stomach contents**)** will be placed in chemically clean glass jars using clean dissection tools as noted above for individual body component subsamples. Each jar will be tared on the Sartorius balance prior to addition of the sample. The sample mass will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (CR for carcass remainders), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

The following glass jars for sample storage have been provided through VWRI Inc, shipped directly to USGS:

TraceClean Precleaned 60 ml glass jars with Teflon lined lids (72 each), production no. 454070 W220-0160

TraceClean Precleaned 125 ml glass jars with Teflon lined lids (72 each), production no. 481036 W220-0125

5. Rinsate ("wipe") blanks. Rinsate blanks will be taken every 20 samples. Clean glass filter paper from a freshly opened package will be used to wipe a set of dissection tools on the areas that routinely contact the specimens during dissection. The paper is then placed in a chemically clean jar and labeled as a "wipe blank" with the date, time of collection, sample mass, and initials of the collector. The procedural wipes will be made using VWR glass microfiber filter 696, 5.5 cm, Lot B1031637 from a freshly opened package

Labeling and recording. Labeling and data recording procedures will follow protocols in Appendix I. Each specimen will have an individual identification number assigned in the field by the New York State Department of Environmental Conservation prior to shipment to USGS in Colorado. Data records to be entered by the dissection team in Colorado will include: identification number, sex, age class (adult or juvenile), reproductive status, intact mass of entire bat, forearm length, date of dissection, and mass of each sample (carcass, brain, stomach contents, carcass remainders, wipe blanks). Each jar will be labeled with field identification number, and contents description (carcass = CA, brain = BR, stomach contents = ST, carcass remainder = CR, or wipe blank), date, sample mass, and initials of dissector.

Sample storage. Samples will be stored as received in Colorado in a –20 C locking upright freezer. Thomas J. O'Shea will be the guardian of the key to the freezer. Prepared samples will also be stored at –20 C. Prepared samples will be held for no longer than two weeks after dissection prior to shipping to the analytical laboratory (Appendix I).

Chain of custody. Strict chain-of-custody protocols will be followed by the dissection team. Chain-of-custody (COC) procedures for sampling and shipment are attached to this document as Appendix I. Each subsample will be recorded on the COC form described in Appendix I.

Tissue disposition. Carcass remainders (heads, feet, legs, wings, skin with pelage, and the gastrointestinal tract minus the stomach contents) will be weighed, refrozen in chemically clean glass jars and shipped to the National Oceanic and Atmospheric Administration's archival facility at Sand Point, Washington. The shipment will be made to the custodian of the NOAA archive, who will be notified by e-mail prior to shipment:

Prior to freezing the head will be placed under a dissecting microscope and dentition examined to obtain an index of relative age. This will be important because of well-known age differences in accumulation of organochlorine contaminants by mammals. However, assignment of actual chronological age in bats is not feasible because of an absence of calibration against known-age specimens. Canine tooth wear (width of the upper canine teeth as measured by an ocular micrometer on a dissection microscope), has been used by others as a likely measure of relative age of adult big brown and little brown bats and will be measured from the remaining head of the specimen (Christian 1956, Clark and Lamont 1976). All surfaces where the head will rest while being measured will be covered with aluminum foil. The aluminum foil will be discarded after each specimen is measured. Dissecting tools will be washed in Alconox and rinsed in acetone and hexane as noted above after each specimen is returned to the glass jar (prior to dissection of a new specimen).

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APPENDIX I.

PROCEDURES FOR BAT SAMPLE SHIPMENT

Chain-of-Custody (COC)

1. Because the sub-samples (resulting from dissection) are considered "new" samples, new COC forms will need to be prepared. The COC form which will be used is attached. The "source" COC forms from NYSDEC should be maintained with the new COC forms.
2. On the new COC forms, list each sub-sample on a separate line, identifying with original sample field ID, sub-sample ID and description.
3. Record on the new COC, as appropriate:
    - Volume or quantity
    - Comments – apparent preservation problems or custody concerns
    - Missing samples or those transferred to another facility
4. Crosscheck all sample identifiers from container to COC before packaging the samples.
5. Sign and date the “Relinquishing Signature” block on the new COC forms.
6. Make a copy for your records and use your copy to fax a set of the new COC forms to the Hudson River Quality Assurance Coordinator.
7. Place both sets of COC forms ("source" and new) in a ziplock bag and tape it to the inside lid of the appropriate cooler.

Packaging/Shipping

1. Individually wrap each jar in bubble wrap and tape wrapping closed. It is important that each jar is well padded—as the jars will move around as the dry ice sublimates.
2. Place dry ice [< 5 kg. (~11 lbs.)/package] in the bottom of the cooler so that the contents will remain at temperature for a minimum of 48 hours (~5 kg (10 - 11 lbs.) is usually adequate for a standard size cooler).

    Note: When using dry ice as a coolant, the drain plug on the cooler must be taped open for ventilation of carbon dioxide (CO_2) gas that occurs when the dry ice vaporizes. Indicate that dry ice has been used as a coolant on the shipping documents, however, when less than 5 kg (11 lbs.) is used as a coolant, dry ice is not considered a “Dangerous Good”.
3. Place a cushioning layer of bubble wrap on top of dry ice, then place the wrapped jars on top of the dry ice. Fill any remaining space in the cooler with bubble wrap or other packing material so that the cooler is full, and jars will not move during transport. Paper material (newspaper, recycled paper, etc.) or vermiculite is not considered acceptable filler material.

4. Tape the COC form(s) to the lid of the cooler (see above).

5. Seal the lid shut. Wrap duct/shipping tape around either end of the cooler (three times) to ensure a tight seal.

6. Place a minimum of two COC seals on the front and two seals on the back of the cooler in such a manner that if the container were opened, the seals would have to be broken.

7. Sign and date the COC seals, which are placed on the outside of the cooler. The same person who signed the COC record should sign the seals.

8. Place clear shipping tape over the COC seals (to avoid smearing or wearing of ink).

9. Adhere the appropriate address label on the top, outside surface of the cooler with clear shipping tape.

10. Fill out appropriate shipping documents:

    - Coolers are to be sent by Federal Express Priority One-Day service or a comparable, traceable service.
    - The cooler/package should be sent to the analytical laboratory.
    - Notify the analytical laboratory by email prior to shipment and copy the Hudson River Quality Assurance Coordinator on the email.

- Do not send coolers on Friday

CHAIN OF CUSTODY RECORD
Hudson River NRDA

Collector:	
Contact Name:	Phone:
Shipping Date:	

Field Sample ID:	Sample ID:	Date Collected:	Description:	Preservation:	Comments:

Total number of items: _____

Relinquished by:	Accepted by:
Signature	Signature
Print Name	Print Name
Date Time	Date Time
Relinquished by:	Accepted by:
Signature	Signature
Print Name	Print Name
Date Time	Date Time
Sample receipt information:	

APPENDIX II. Step-by-step instructions for Hudson River bat dissections. Reference area bats will be dissected first. These will be followed by dissections of Hudson River bats in the following order: adult females, young of the year, adult males. In addition to the COC forms, two data forms will be completed as each specimen is processed (Appendix III). One is a specimen data sheet with information on characteristics of each intact specimen, The second is a biological sample data sheet with information on mass of each subsample.

Obtain the first rinsate blank prior to dissections.

1. Remove bat from freezer in original jar. Leave in jar at room temperature until thawed.

2. Obtain four chemically clean glass jars (2—60 ml and 2--125 ml). Determine mass of each jar without lid or label. Put blank label on each jar. Label jars with field identification number copied from the jar that the bat comes in from NY, and the following initials for identification of jar contents: BR for brain (60 ml jar), CA for carcass (125 ml jar), ST for stomach contents (60 ml jar) and CR for carcass remainders (125 ml jar).

3. Remove thawed bat from original jar using forceps previously washed in 1% Alconox and rinsed in acetone and hexane. Place on clean, unused sheet of aluminum foil. Examine bat to verify species, relative age, sex, and reproductive condition on the specimen data sheet (Appendix III). Handle only with forceps. Measure the length of the right forearm to the nearest 0.1 mm using dial calipers with contact points previously washed in Alconox, rinsed in acetone and hexane. Tare a smaller clean piece of aluminum foil on the Sartorius balance, and place the intact undissected carcass on the foil on the balance. Determine the mass of the bat to the nearest 0.001 g on the Sartorius balance. Record species, sex, age class, reproductive condition, forearm length, and body mass on the specimen data sheet (Appendix III).

4. Make a small incision through the skin in the abdomen with dissecting scissors previously washed in Alconox and rinsed in acetone and hexane. Be careful not to perforate the abdominal wall. Continue to slit the skin to the level of the neck. Using forceps previously washed in Alconox and rinsed in acetone and hexane, pull the skin and pelage away from the legs to the level of the foot. Cut the leg bones with scissors at the ankle, and pull the tail free from the uropatagium with forceps. Proceed cranially and work the skin free from the back and shoulders. Cut each humerus with scissors at the level of the shoulder. Continue to work the skin cranially over the head, cutting the external ear canal, eyelids, and lips free from the skull with a scalpel previously washed in Alconox and rinsed in acetone and hexane. Use the scalpel to cut the entire skin free from the bat at the nose. Place the skin with pelage, wings, and feet onto a small clean sheet of aluminum foil for the sample destined for the jar marked CR for carcass remainder. Remove any major visible deposits of fat adhering to the skin with the forceps and place those on a separate small clean sheet of aluminum foil for the sample destined for the jar marked CA.

5. Sever the head from the body at the first cranial vertebrae with scissors. Using a scissors washed in Alconox and rinsed in acetone and hexane, snip the occipital area of the skull starting at the foramen magnum to the lateral area of the parietal bones. Snip the rear of the skull across the top at the parietals. Using a stainless steel spatula or forceps washed in Alconox and rinsed in acetone and hexane, remove the brain and place it on a small clean sheet of aluminum foil. Tare the glass jar

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Name (printed): _____ Name (printed): _____

Signature: _____ Signature: _____

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HUDSON RIVER NATURAL RESOURCE DAMAGE ASSESSMENT

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Dissection Team Leader

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Quality Assurance Coordinator

November 13, 2002

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See also Appendix II for step-by-step dissection instructions.

Dissection tools and surfaces. Dissections will be carried out using standard stainless steel instruments for small mammals: dissection scissors, scalpels with disposable blades, forceps, and spatulas. Stainless steel dissecting instruments will be washed in a 1% solution of deionized water and Alconox detergent (lot A2 H2, Alconox Inc), rinsed in acetone, followed by a rinse in hexane, before each animal is dissected, and before each sample type (carcass, brain, stomach contents, carcass remainder) is prepared within individual animals. Alconox, acetone and hexane have been supplied in sealed containers through VWR Inc. and have been stored at USGS in Fort Collins. Both acetone (Burdick and Jackson, lot CD 935) and hexane (Burdick and Jackson, lot CF765) are capillary GC/GC-MS solvent grade suitable for trace analysis at ppb levels or less. During dissections solvents will be stored in 500 ml Teflon FEP wash bottles (Nalge Nunc. Inc. lot 465201)

All dissections will take place under a laboratory fume hood at the U.S. Geological Survey laboratory facility in Fort Collins. All contact of specimens with synthetic surfaces, plastics, soaps, or oils will be avoided. Laboratory bench surfaces will be wiped clean with acetone and hexane, and covered with aluminum foil obtained in sealed packages from commercial retail outlets. Aluminum foil in work areas will be changed and replaced with clean foil after each bat is dissected, and clean foil pieces will be used for dissections of individual bat subsamples. Dissectors will wear latex gloves, but will handle samples destined for PCB analysis only using stainless steel dissection tools washed in Alconox and rinsed in acetone and hexane. Latex gloves will be discarded and changed between bat samples.

Sequences of dissection. Technicians will dissect and process each bat separately, singly, and completely before another bat is processed. Each frozen bat will be thawed singly, dissections completed, samples placed in chemically clean glass sample containers, mass of samples determined and recorded, and samples returned to the freezer before another bat is processed by the same technician. Samples will be processed in two separate lots (reference area and Hudson River) to avoid potential cross-contamination, with reference area bats sampled first. Within each species, samples will be prepared beginning with individuals least likely to be contaminated (adult females will be dissected first, followed by young of the year, and then adult males).

Dissection procedures. Each bat will be thawed individually and separately at room temperature. Prior to dissection, technicians will verify sex and age class (adult or juvenile based on closure of phalangeal epiphyses—Anthony 1988) determinations made in the field in New York. Reproductive status (pregnant, lactating, non-reproductive, testes scrotal or non-scrotal—Racey 1988) will also be noted. Body mass will be recorded to the nearest 0.001 g on a Sartorius Model CP225D digital balance with internal calibration weights, and forearm lengths measured to the nearest 0.1 mm with dial calipers. This information will be recorded on data sheets (Appendix III) in permanent ink using field identification numbers originally assigned in New York. The Sartorius model CP225D balance has 220 g capacity and 0.1 mg precision.

The following subsamples will be collected, as described below and in Appendix II: carcass, brain, stomach contents, carcass remainders, and wipe blanks.

1. Carcasses. Carcasses will consist of the entire body minus the head, feet, wings, skin with pelage, and gastrointestinal tracts (total of these minus the stomach contents = carcass remainders, see item 4 below), following procedures established for contaminant analysis of bats by the Patuxent Wildlife Research Center (e.g. Clark et al. 1975, Clark and Lamont 1976, Clark and Prouty 1976, Clark and Stafford 1981, O'Shea et al. 2001). Dissections will be made using stainless steel dissection tools washed in the Alconox solution and rinsed in acetone followed by hexane. The head will be separated at the first cervical vertebrae. Major head musculature will be trimmed and placed with the carcass. This is the procedure followed by most of the published literature on contaminants in bats (see references in Clark and Shore 2001). Each carcass will be placed individually in chemically clean glass sample jars. Each jar will be tared on the Sartorius balance prior to addition of the sample. Mass of the sample will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (CA for carcasses), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

2. Brains. The dorsal and lateral skin of the head will be removed using stainless steel scissors or scalpels. The cranium will be opened by removal of the occipital region and portions of the parietal regions using stainless steel dissection scissors washed in Alconox and rinsed in acetone and hexane immediately prior to opening. The brain will be removed entire using stainless steel forceps or stainless steel spatulae, also washed in Alconox and rinsed in acetone and hexane immediately prior to use. The brain will be placed in a chemically clean glass sample jar. Each jar will be tared on the Sartorius balance prior to addition of the sample. The sample mass will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (BR for brains), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

3. Stomach contents. The stomach will be opened using stainless steel dissection scissors washed in Alconox solution and rinsed in acetone and hexane immediately prior to opening. Contents will be removed using stainless steel forceps or stainless steel spatulae, also washed in Alconox detergent and rinsed in acetone and hexane immediately prior to use. The stomach contents will be placed in chemically clean glass sample jars. The stomach and gastrointestinal tract tissues will be retained for inclusion with the other carcass remainders (see 4. below). Each jar will be tared on the Sartorius balance prior to addition of the sample. The sample mass will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (ST for stomach contents), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

4. Carcass remainders. Carcass remainders **(**head, feet, wings, skin with pelage and gastrointestinal tracts minus the stomach contents**)** will be placed in chemically clean glass jars using clean dissection tools as noted above for individual body component subsamples. Each jar will be tared on the Sartorius balance prior to addition of the sample. The sample mass will be determined to the nearest 0.001 g. Each jar will be labeled with the field identification number (from the New York State Department of Environmental Conservation), subsample identification (CR for carcass remainders), date of dissection (mm/dd/yy), sample mass, and initials of dissector. The jar and contents will then be immediately returned to the freezer.

The following glass jars for sample storage have been provided through VWRI Inc, shipped directly to USGS:

TraceClean Precleaned 60 ml glass jars with Teflon lined lids (72 each), production no. 454070 W220-0160

TraceClean Precleaned 125 ml glass jars with Teflon lined lids (72 each), production no. 481036 W220-0125

5. Rinsate ("wipe") blanks. Rinsate blanks will be taken every 20 samples. Clean glass filter paper from a freshly opened package will be used to wipe a set of dissection tools on the areas that routinely contact the specimens during dissection. The paper is then placed in a chemically clean jar and labeled as a "wipe blank" with the date, time of collection, sample mass, and initials of the collector. The procedural wipes will be made using VWR glass microfiber filter 696, 5.5 cm, Lot B1031637 from a freshly opened package

Labeling and recording. Labeling and data recording procedures will follow protocols in Appendix I. Each specimen will have an individual identification number assigned in the field by the New York State Department of Environmental Conservation prior to shipment to USGS in Colorado. Data records to be entered by the dissection team in Colorado will include: identification number, sex, age class (adult or juvenile), reproductive status, intact mass of entire bat, forearm length, date of dissection, and mass of each sample (carcass, brain, stomach contents, carcass remainders, wipe blanks). Each jar will be labeled with field identification number, and contents description (carcass = CA, brain = BR, stomach contents = ST, carcass remainder = CR, or wipe blank), date, sample mass, and initials of dissector.

Sample storage. Samples will be stored as received in Colorado in a –20 C locking upright freezer. Thomas J. O'Shea will be the guardian of the key to the freezer. Prepared samples will also be stored at –20 C. Prepared samples will be held for no longer than two weeks after dissection prior to shipping to the analytical laboratory (Appendix I).

Chain of custody. Strict chain-of-custody protocols will be followed by the dissection team. Chain-of-custody (COC) procedures for sampling and shipment are attached to this document as Appendix I. Each subsample will be recorded on the COC form described in Appendix I.

Tissue disposition. Carcass remainders (heads, feet, legs, wings, skin with pelage, and the gastrointestinal tract minus the stomach contents) will be weighed, refrozen in chemically clean glass jars and shipped to the National Oceanic and Atmospheric Administration's archival facility at Sand Point, Washington. The shipment will be made to the custodian of the NOAA archive, who will be notified by e-mail prior to shipment:

Prior to freezing the head will be placed under a dissecting microscope and dentition examined to obtain an index of relative age. This will be important because of well-known age differences in accumulation of organochlorine contaminants by mammals. However, assignment of actual chronological age in bats is not feasible because of an absence of calibration against known-age specimens. Canine tooth wear (width of the upper canine teeth as measured by an ocular micrometer on a dissection microscope), has been used by others as a likely measure of relative age of adult big brown and little brown bats and will be measured from the remaining head of the specimen (Christian 1956, Clark and Lamont 1976). All surfaces where the head will rest while being measured will be covered with aluminum foil. The aluminum foil will be discarded after each specimen is measured. Dissecting tools will be washed in Alconox and rinsed in acetone and hexane as noted above after each specimen is returned to the glass jar (prior to dissection of a new specimen).

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APPENDIX I.

PROCEDURES FOR BAT SAMPLE SHIPMENT

Chain-of-Custody (COC)

1. Because the sub-samples (resulting from dissection) are considered "new" samples, new COC forms will need to be prepared. The COC form which will be used is attached. The "source" COC forms from NYSDEC should be maintained with the new COC forms.
2. On the new COC forms, list each sub-sample on a separate line, identifying with original sample field ID, sub-sample ID and description.
3. Record on the new COC, as appropriate:
    - Volume or quantity
    - Comments – apparent preservation problems or custody concerns
    - Missing samples or those transferred to another facility
4. Crosscheck all sample identifiers from container to COC before packaging the samples.
5. Sign and date the “Relinquishing Signature” block on the new COC forms.
6. Make a copy for your records and use your copy to fax a set of the new COC forms to the Hudson River Quality Assurance Coordinator.
7. Place both sets of COC forms ("source" and new) in a ziplock bag and tape it to the inside lid of the appropriate cooler.

Packaging/Shipping

1. Individually wrap each jar in bubble wrap and tape wrapping closed. It is important that each jar is well padded—as the jars will move around as the dry ice sublimates.
2. Place dry ice [< 5 kg. (~11 lbs.)/package] in the bottom of the cooler so that the contents will remain at temperature for a minimum of 48 hours (~5 kg (10 - 11 lbs.) is usually adequate for a standard size cooler).

    Note: When using dry ice as a coolant, the drain plug on the cooler must be taped open for ventilation of carbon dioxide (CO_2) gas that occurs when the dry ice vaporizes. Indicate that dry ice has been used as a coolant on the shipping documents, however, when less than 5 kg (11 lbs.) is used as a coolant, dry ice is not considered a “Dangerous Good”.
3. Place a cushioning layer of bubble wrap on top of dry ice, then place the wrapped jars on top of the dry ice. Fill any remaining space in the cooler with bubble wrap or other packing material so that the cooler is full, and jars will not move during transport. Paper material (newspaper, recycled paper, etc.) or vermiculite is not considered acceptable filler material.

4. Tape the COC form(s) to the lid of the cooler (see above).

5. Seal the lid shut. Wrap duct/shipping tape around either end of the cooler (three times) to ensure a tight seal.

6. Place a minimum of two COC seals on the front and two seals on the back of the cooler in such a manner that if the container were opened, the seals would have to be broken.

7. Sign and date the COC seals, which are placed on the outside of the cooler. The same person who signed the COC record should sign the seals.

8. Place clear shipping tape over the COC seals (to avoid smearing or wearing of ink).

9. Adhere the appropriate address label on the top, outside surface of the cooler with clear shipping tape.

10. Fill out appropriate shipping documents:

    - Coolers are to be sent by Federal Express Priority One-Day service or a comparable, traceable service.
    - The cooler/package should be sent to the analytical laboratory.
    - Notify the analytical laboratory by email prior to shipment and copy the Hudson River Quality Assurance Coordinator on the email.

- Do not send coolers on Friday

CHAIN OF CUSTODY RECORD
Hudson River NRDA

Collector:	
Contact Name:	Phone:
Shipping Date:	

Field Sample ID:	Sample ID:	Date Collected:	Description:	Preservation:	Comments:

Total number of items: _____

Relinquished by:	Accepted by:
Signature	Signature
Print Name	Print Name
Date Time	Date Time
Relinquished by:	Accepted by:
Signature	Signature
Print Name	Print Name
Date Time	Date Time
Sample receipt information:	

APPENDIX II. Step-by-step instructions for Hudson River bat dissections. Reference area bats will be dissected first. These will be followed by dissections of Hudson River bats in the following order: adult females, young of the year, adult males. In addition to the COC forms, two data forms will be completed as each specimen is processed (Appendix III). One is a specimen data sheet with information on characteristics of each intact specimen, The second is a biological sample data sheet with information on mass of each subsample.

Obtain the first rinsate blank prior to dissections.

1. Remove bat from freezer in original jar. Leave in jar at room temperature until thawed.

2. Obtain four chemically clean glass jars (2—60 ml and 2--125 ml). Determine mass of each jar without lid or label. Put blank label on each jar. Label jars with field identification number copied from the jar that the bat comes in from NY, and the following initials for identification of jar contents: BR for brain (60 ml jar), CA for carcass (125 ml jar), ST for stomach contents (60 ml jar) and CR for carcass remainders (125 ml jar).

3. Remove thawed bat from original jar using forceps previously washed in 1% Alconox and rinsed in acetone and hexane. Place on clean, unused sheet of aluminum foil. Examine bat to verify species, relative age, sex, and reproductive condition on the specimen data sheet (Appendix III). Handle only with forceps. Measure the length of the right forearm to the nearest 0.1 mm using dial calipers with contact points previously washed in Alconox, rinsed in acetone and hexane. Tare a smaller clean piece of aluminum foil on the Sartorius balance, and place the intact undissected carcass on the foil on the balance. Determine the mass of the bat to the nearest 0.001 g on the Sartorius balance. Record species, sex, age class, reproductive condition, forearm length, and body mass on the specimen data sheet (Appendix III).

4. Make a small incision through the skin in the abdomen with dissecting scissors previously washed in Alconox and rinsed in acetone and hexane. Be careful not to perforate the abdominal wall. Continue to slit the skin to the level of the neck. Using forceps previously washed in Alconox and rinsed in acetone and hexane, pull the skin and pelage away from the legs to the level of the foot. Cut the leg bones with scissors at the ankle, and pull the tail free from the uropatagium with forceps. Proceed cranially and work the skin free from the back and shoulders. Cut each humerus with scissors at the level of the shoulder. Continue to work the skin cranially over the head, cutting the external ear canal, eyelids, and lips free from the skull with a scalpel previously washed in Alconox and rinsed in acetone and hexane. Use the scalpel to cut the entire skin free from the bat at the nose. Place the skin with pelage, wings, and feet onto a small clean sheet of aluminum foil for the sample destined for the jar marked CR for carcass remainder. Remove any major visible deposits of fat adhering to the skin with the forceps and place those on a separate small clean sheet of aluminum foil for the sample destined for the jar marked CA.

5. Sever the head from the body at the first cranial vertebrae with scissors. Using a scissors washed in Alconox and rinsed in acetone and hexane, snip the occipital area of the skull starting at the foramen magnum to the lateral area of the parietal bones. Snip the rear of the skull across the top at the parietals. Using a stainless steel spatula or forceps washed in Alconox and rinsed in acetone and hexane, remove the brain and place it on a small clean sheet of aluminum foil. Tare the glass jar

labeled BR for brain on the Sartorius balance. Place the brain in the jar and record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the contents on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

6. Return to the specimen. Remove the bulk of the major head musculature (masseter and temporalis) with scissors freshly washed in Alconox and rinsed in acetone and hexane, and place on the sheet of foil with the material destined for the jar marked CA. Place the head aside on clean aluminum foil for placement under a dissecting scope and measurement of tip width of both upper canine teeth using an ocular micrometer.

7. Return to the skinned carcass. Using scissors freshly washed in Alconox and rinsed in acetone and hexane, perforate the abdominal wall. Cut in a cranial direction through the ribs and retract the rib cage if necessary. Remove the gastrointestinal tract by handling with clean forceps, cutting at the esophagus above the stomach, at the colon above the rectum, and trimming any adhering mesenteries. Place the gastrointestinal tract on a clean piece of foil. Place the remaining carcass on the small piece of aluminum foil destined for the jar marked CA. Tare the jar marked CA, add the carcass material from the sheet of foil, and record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the contents on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

8. Tare the clean glass jar marked ST. Open the stomach with dissecting scissors, and use a freshly cleaned spatula to remove any visible contents and place contents in the tared clean glass jar marked ST. Record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the contents on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

9. Place the gastrointestinal tract empty of stomach contents on the sheet of foil destined for the jar marked CR. After the tooth width is measured, place the head on this sheet of foil. Tare the clean glass jar marked CR. Place the carcass remainders in the jar and record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the carcass remainder on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

10. Remove all foil from dissection area under hood and discard in biohazard bag. Change latex gloves. Place clean foil on bench surface under hood. Cut all clean sheets of foil necessary for next carcass and place in dissecting area. Rinse all dissecting tools in tap water, wash in Alconox, rinse thoroughly in deionized water, followed by acetone, followed by a hexane rinse. Use the Teflon wash bottles for the acetone and hexane rinses. Place clean tools in dissecting area for use with the next bat.

11. Repeat steps 1-10 with the next bat. At two points in the process – before beginning the first bat, and after the 20th bat is dissected– obtain a rinsate blank of the dissection tools using the glass filter paper and the protocol described in the "Rinsate ("wipe") blanks" section of this document. Obtain these after the tools have been cleaned, washed and rinsed using the standard protocols described in this document. Only wipe those parts of the tools that contact the bat samples during dissection.

labeled BR for brain on the Sartorius balance. Place the brain in the jar and record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the contents on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

6. Return to the specimen. Remove the bulk of the major head musculature (masseter and temporalis) with scissors freshly washed in Alconox and rinsed in acetone and hexane, and place on the sheet of foil with the material destined for the jar marked CA. Place the head aside on clean aluminum foil for placement under a dissecting scope and measurement of tip width of both upper canine teeth using an ocular micrometer.

7. Return to the skinned carcass. Using scissors freshly washed in Alconox and rinsed in acetone and hexane, perforate the abdominal wall. Cut in a cranial direction through the ribs and retract the rib cage if necessary. Remove the gastrointestinal tract by handling with clean forceps, cutting at the esophagus above the stomach, at the colon above the rectum, and trimming any adhering mesenteries. Place the gastrointestinal tract on a clean piece of foil. Place the remaining carcass on the small piece of aluminum foil destined for the jar marked CA. Tare the jar marked CA, add the carcass material from the sheet of foil, and record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the contents on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

8. Tare the clean glass jar marked ST. Open the stomach with dissecting scissors, and use a freshly cleaned spatula to remove any visible contents and place contents in the tared clean glass jar marked ST. Record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the contents on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

9. Place the gastrointestinal tract empty of stomach contents on the sheet of foil destined for the jar marked CR. After the tooth width is measured, place the head on this sheet of foil. Tare the clean glass jar marked CR. Place the carcass remainders in the jar and record the mass on the biological sample data sheet. Put the lid on the jar, record the mass of the carcass remainder on the label using an indelible "Sharpie" pen, cover the label with clear packing tape, and place the specimen in the freezer.

10. Remove all foil from dissection area under hood and discard in biohazard bag. Change latex gloves. Place clean foil on bench surface under hood. Cut all clean sheets of foil necessary for next carcass and place in dissecting area. Rinse all dissecting tools in tap water, wash in Alconox, rinse thoroughly in deionized water, followed by acetone, followed by a hexane rinse. Use the Teflon wash bottles for the acetone and hexane rinses. Place clean tools in dissecting area for use with the next bat.

11. Repeat steps 1-10 with the next bat. At two points in the process – before beginning the first bat, and after the 20th bat is dissected– obtain a rinsate blank of the dissection tools using the glass filter paper and the protocol described in the "Rinsate ("wipe") blanks" section of this document. Obtain these after the tools have been cleaned, washed and rinsed using the standard protocols described in this document. Only wipe those parts of the tools that contact the bat samples during dissection.

APPENDIX III. SPECIMEN AND BIOLOGICAL SAMPLE DATA FORMS

SPECIMEN DATA FORM—HUDSON RIVER BAT STUDY

Specimen Field ID	Species	Sex	Age	Forearm Length (mm)	Body Mass (g)	Canine Width (mm)	Reprod. Status	Comments

Abbreviations: EPFU = *Eptesicus fuscus*, MYLU = *Myotis lucifugus*; M = male, F = female; Ad = Adult, YOY = young-of-the-year; SC = Scrotal, NR = Non-Reproductive; P = pregnant; L = lactating; PL = post-lactating.

Page ____ of _____

BIOLOGICAL SAMPLE DATA FORM—HUDSON RIVER BAT STUDY

Sample ID	Sample Type	Jar Mass	Sample Mass	Dissector	Date

Sample type = brain, carcass, carcass remainder, stomach contents. Mass in grams to nearest 0.001 g. Jar mass is jar without lid or labels. Date format is mm/dd/yy.

Page ____ of _____

Appendix B

Bat Collection Data Sheet - 2002 Collection

Hudson River Bat Sampling Project Data Sheet Number _____

<u>Bat Collection Data Sheet</u>

COLLECTION:

Site Number:_____

Species (Bb or Lb) and specimen #:_____

Name of Field Crew Leader (print):
_____

Name of Field Data Recorder (print): _____

Date of Collection: _____ Time of Collection: _____

GPS location: _____

Note: If GPS location cannot be provided at time of collection, other sufficient identifying information for location shall be provided in "NOTES" section below to allow GPS coordinates to be subsequently obtained, if possible.

Site Name / Description:
_____
_____
_____

NOTES:
_____
_____
_____
_____
_____
_____

Initials/Date of Field Data Recorder: _____

Initials/Date of Field Crew Leader: _____

Initials/Date of Review of Data Sheet by Field Investigation Lead: _____

Appendix C

Chain-of-Custody Record

CHAIN OF CUSTODY RECORD

Project Name: Hudson River NRDA - Collection of Little Brown and Big Brown Bats

Collectors: Printed Name and Signature

Site Number (Bat Sample #)	Date Collected	Location	Remarks

Special Instructions/Comments:

	Signature	Print Name	Company/Title	Date	Time
Relinquished by:					
Received by:					
Relinquished by:					
Received by:					
Relinquished by:					
Received by:					
Relinquished by:					
Received by:					
Relinquished by:					

Appendix D

Data Quality Assessment Report - Bat Brain Tissue

DATA QUALITY ASSESSMENT REPORT

HUDSON RIVER NATURAL RESOURCE DAMAGE ASSESSMENT

FLOODPLAIN STUDY
Bat Brain Tissue

Prepared for:

State of New York
Department of Environmental Conservation

U.S. Department of Commerce
National Oceanic and Atmospheric Administration

U.S. Department of Interior
Fish and Wildlife Service

April 14, 2004

Version 1.0

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DATA QUALITY ASSESSMENT
Hudson River Natural Resource Damage Assessment
Bat Brain Tissue

1.0 INTRODUCTION

This report documents the results of a quality assurance review of data from bat brain tissue samples collected in support of the Hudson River Natural Resource Damage Assessment. The samples were analyzed for PCB congeners, PCB homologue groups, and total PCBs. Percent lipids and percent moisture were not analyzed due to limited sample size.

The bats were collected as part of a field test of the Anabat system. The Anabat system is a detector that provides a quantitative identification of free flying bats. During the field testing period (July 15 through July 26, 2001), a total of 75 bats were collected.

Thirty-one bats were selected for analysis for PCBs: 19 bats from the Hudson River study site and 12 bats from reference areas. The sample of bats from the Hudson River study site consists of 14 little brown bats (*Myotis lucifugus* – 2 adult males, 1 young-of-the-year male, and 11 adult females) and 5 big brown bats (*Eptesicus fuscus* – all adult females). The reference sample of bats consists of 12 adult female little brown bats from White Birch Pond and the Black River. The bats were frozen and stored at the New York State Department of Environmental Conservation Hale Creek laboratory until November, 2002.

The bats were transported under chain-of-custody to the US Geological Survey (USGS) Fort Collins Science Center. Carcass dissections were carried out by technicians from Colorado State University (CSU) under the supervision of Thomas J. O'Shea of the U.S. Geological Survey (USGS). The bat brains and stomach contents were removed, and were shipped (along with the carcass) to the analytical chemistry laboratory. The carcass remains were shipped to the NOAA archive at Sandpoint, Washington.

A total of 31 bat brains were submitted for analysis. These samples were analyzed in two laboratory analytical batches, 0307080 and 0307081. The brain tissues were prepped, extracted, and analyzed using laboratory Standard Operating Procedures (SOPs) that were submitted and reviewed by the Hudson River Quality Assurance Coordinator prior to sample receipt.

2.0 DATA VALIDATION PROCEDURES

Data validation was based on the quality assurance/quality control (QA/QC) criteria documented in the *Analytical Quality Assurance Plan for the Hudson River Natural Resource Damage Assessment*, Version 1.0, July 9, 2002, and USEPA *National Functional Guidelines for Organic Data Review*, 1999, and the following laboratory SOPs:

- SOP # HR NRDA Project Tissue Prep: Tissue Preparation and Homogenization, Revision #1.0, 9/25/02
- SOP # OP-016: Microscale Solvent Extraction (MSE), Revision #1.0, 1/29/04
- SOP # O-010: Determination of PCB Homologues and Individual Congeners by GC/MS - SIM, Revision # 2.2, 10/24/02
- Additional cleanup, sample handling, storage, custody SOPs as necessary.

Sample results and related QC data were received in both an electronic and hard copy format. Electronic data were verified against the hard copy data package. One data package received a full validation, and one package received a summary validation.

The following QC elements were reviewed for data packages undergoing summary validation:

- Analytical holding times
- Chain of custody and sample handling
- GC/MS tune verification (from summary forms)
- Method blank contamination (from summary forms)
- Initial and continuing calibration (from summary forms)
- Rinsate blank contamination (from sample result summaries)
- Analytical accuracy: surrogates, matrix spike samples, laboratory control samples, and standard reference material results (from summary forms)
- Analytical precision: laboratory duplicate samples (from summary forms)
- Internal standard areas (from summary forms)
- Reported detection limits (from sample result summaries)

Full validation included review of all the items listed above for summary validation, plus the following QC elements:

- Compound identification (from raw data)
- Compound quantitation, transcription and calculation checks performed at a frequency of 10% from raw data. If an error was noted, 100% of the calculations and transcriptions for that data set were verified.

This report summarizes the results of data validation relative to the analytical data quality objectives (ADQO) for precision, accuracy, and completeness. The report also provides a quantitative and qualitative assessment of the data and identifies potential sources of error, uncertainty, and bias that may affect the overall usability.

Laboratory QC samples were used to assess the effectiveness of homogenization procedures and to evaluate laboratory-derived contamination, laboratory performance, and sample matrix effects. Quality control samples included: method blanks, laboratory control samples (LCS), matrix spike (MS) samples, laboratory duplicate samples, and standard reference material (SRM) analyses. Surrogates were added to each sample analyzed for PCB congeners to further assess the effects of sample matrix on accuracy.

Data were qualified when associated QC sample results were outside the QC limits. The following definitions provide brief explanations of the qualifiers assigned to results in the data validation process:

J **Estimated**: The associated numerical value is an estimated quantity. The analyte was detected, but the reported value may not be accurate or precise. The "J" qualification indicates the data fell outside the QC limits, but the exceedance was not sufficient to cause rejection of the data.

3.0 DATA QUALITY ASSESSMENT

The data package submitted by the laboratory was reviewed to determine whether the ADQO specified in Tables 6.1a - 6.1c in the *Analytical Quality Assurance Plan* were met. Each quality control element is discussed briefly below. More details are available in the individual data validation reports presented in **Attachment A**.

3.1 Holding Times and Sample Preservation

The primary analytes of concern for this study are persistent compounds, which have been found to remain stable in tissue after several years of storage. Due to this, no maximum holding time criterion was established. The bat brain tissues were extracted approximately 13 months after collection. All extracts were analyzed within 30 days from sample extraction. Samples were kept frozen by the laboratory at the required temperature of -20°C ±2°.

3.2 Instrument Calibration

3.2.1 Initial Calibration (ICAL)

The ADQO specification for the initial calibration is that a minimum of a five-point calibration would be performed for all analytes, and that the percent relative standard deviation (%RSD) values for all analytes are less than 20%. All submitted ICAL data met the specified ADQO.

3.2.2 Continuing Calibration (CCAL)

The ADQO specified for the continuing (or daily) calibrations is that a CCAL must be analyzed at the beginning and end of each analytical sequence (or every 12 hours, whichever is more frequent), and that all percent difference (%D) values must be less than 20%. Up to three %D values can be greater than 20%, provided that all %D values are less than 30%.

All CCAL met the ADQO requirements. One %D value was greater than 20% (but less than 30%) in one CCAL. This outlier was associated only with one sample. The PCB31/28 result was estimated (J) in Sample White Birch 4 based on the CCAL %D outlier.

3.3 GC/MS Tune

GC/MS instrument tuning verifications were performed at the proper frequency, prior to each analytical sequence. All GC/MS tunes met the acceptance criteria specified in the laboratory standard operating procedures.

3.4 Blank Analyses

The method blank was acceptable, in that no target analytes were detected in the method blank.

3.5 Accuracy

Accuracy is evaluated by comparison of an analytical concentration to a known (true) value. Accuracy was monitored through the use of surrogate compounds in each sample, and SRM, MS, and LCS (blank spike) analyses. Each QC element is discussed below. Overall, accuracy was acceptable.

3.5.1 Surrogate Compounds

Two surrogate compounds, ^{13}C-BZ#19 and ^{13}C-BZ#202, were added to each sample prior to extraction.

The ADQO specified for surrogate compounds is that all percent recovery (%R) values would be within the 50% - 125% acceptance window. The recovery value from the late eluting surrogate (^{13}C-BZ#202) is used for the quantitation of the reported target analyte concentrations.

All of the surrogate %R values were within the 50% - 125% control limits.

3.5.2 Standard Reference Material Analyses

An SRM was extracted and analyzed with the analytical batch. The SRM selected for this study was 1974b, Organics in Mussel Tissue. This SRM has certified values for 27 PCB congeners. One SRM analysis was submitted with each laboratory batch, so two SRM analyses (with 27 certified values each, for a total of 54 data points) were performed.

The ADQO for the SRM is that the reported value must be within ±20% of the 95% confidence interval of the true value, for analytes with values greater than five times the method detection limit (MDL). This ADQO was used by the laboratory to evaluate the reported results. However, during data validation, no data were qualified unless the reported value was greater than ±25% of the 95% confidence interval.

Overall, the SRM accuracy results were acceptable. A total of five SRM results (9.3% of the total) were greater than the upper control limit (125% of the 95% confidence interval for an individual congener). The outliers were for each of the following congeners: PCB31/28, PCB87, PCB118, and PCB138. The positive results associated with these SRM outliers were estimated (J) to indicate a potential high bias. Forty-eight results (2.7% of the bat brain results) were estimated based on these SRM outliers.

Tables 1A and **1B** summarize the SRM results for this study.

3.5.3 Laboratory Control Samples

The laboratory performed LCS analyses at the required frequency of one for every 15 samples or analytical batch, whichever was more frequent. The ADQO for the LCS analyses is that all %R values must be within the acceptance limits of 75% to 125%; however, if only one analyte %R value is outside the control limits, the laboratory is not required to re-extract the associated samples.

Two LCS analyses were submitted with the bat brain tissue samples. Each LCS included 47 target analytes, for a total of 94 data points (%R values). Five LCS %R values were greater than the 125% upper control limit, indicating a potential high bias. Associated positive results were estimated (J); reporting limits were judged as unaffected and no action was taken. Thirty-one data points (1.7% of the bat brain results) were estimated based on the LCS %R outliers.

3.5.4 Matrix Spike Samples

Matrix spike analyses were not performed due to limited sample volume.

3.5.5 Internal Standards

Internal standards were added to each field and QC sample prior to injection onto the analytical instrument. The ADQO for internal standards is that the area of the internal standards in each analysis must be within ±50% of the area of the internal standard in the associated CCAL.

All internal standard areas met the ADQO.

3.6 Precision

Precision is evaluated through replicate analyses of a sample. For this study, laboratory duplicate samples could not be analyzed due to the limited sample volume. An SRM was analyzed with each batch. Overall, precision was judged as acceptable.

3.6.1 Standard Reference Material Analyses

Section 3.5.2 describes the frequency and criteria for the SRM analyses performed with each analytical batch. The results for the SRM analyses associated with the bat brain tissues are summarized in **Tables 1A** and **1B**.

SRM accuracy outliers are discussed in **Section 3.5.2**. For congeners with elevated recovery values, the precision %D results (compared to the certified values) were also elevated. For the other congeners, the average %D from the certified values for the bat brain SRM results ranged from 0.81% to 19.8% for each of the congeners with values greater than five times the MDL. All %D values are less than 20%, indicating good overall precision between the analytical batches.

3.7 Reporting Limits and Sample Results

Method detection limits (MDLs) were determined using low level spikes in tissue from laboratory mice following procedures outlined in the *US Code of Federal Regulations* (40 CFR Part 136, Appendix B).

The detection limits for target congeners were generally in the range of 0.02 µg/Kg to 0.22 µg/Kg, with a co-elution of PCB31/28 at 1.38 g/Kg. There were 14 target congeners with MDL values greater than the 0.1 µg/Kg target MDL. The PCB31/28 co-elution MDL value was elevated due to interferences which could not be resolved using the selected method.

Chromatography and mass spectral identification were reviewed for a minimum of 10% of the reported congeners. No instances of potential interference were noted. All reported positive results met the identification criteria and chromatographic peak shapes were acceptable. All reported results were judged to be accurate unless qualified for some other reason.

3.8 Completeness

Out of 1,798 results reported by the laboratory (31 samples each with 47 congeners, ten homologue groups, and total PCBs), a total of 66 (3.7%) data points were qualified as estimated (J). No data were rejected. The completeness level attained for the analysis of the field samples is 100%.

3.9 Summary of Data Usability

A total of 66 out of 1,798 results were estimated because of laboratory accuracy outliers. For all data, the overall quality of the data is acceptable and the results, as qualified, are considered usable.

TABLE 1A
Summary of Standard Reference Material Results

STANDARD REFERENCE MATERIAL 1974b - Organic in Mussel Tissue (Mytilus edulis)

Concentrations are ng/g, wet weight

PCB Congener	SDG 0307080 ng/g	SDG 0307081 ng/g	True Value ng/g	Uncertainty (+/-)	+/- 25% Limits ng/g From	To
PCB18	1.21 U	1.00 U	0.84	0.13	0.50	1.18
PCB28/31	21.8	10.5	6.31	0.48	4.25	8.37
PCB44	4.48	4.13	3.85	0.20	2.69	5.01
PCB49	4.45	4.63	5.66	0.23	4.02	7.31
PCB52	6.56	6.78	6.26	0.37	4.33	8.20
PCB66	6.98	6.60	6.37	0.37	4.41	8.33
PCB70	6.94	7.36	6.01	0.22	4.29	7.73
PCB74	4.32	3.52	3.55	0.23	2.43	4.67
PCB87	5.9	5.26	4.33	0.36	2.89	5.77
PCB95	6.53	7.10	6.04	0.36	4.17	7.91
PCB99	5.93	5.78	5.92	0.27	4.17	7.67
PCB101	11.8	13.3	10.70	1.10	6.93	14.5
PCB105	4.39	4.15	4.00	0.18	2.82	5.18
PCB110	9.63	10.2	10.0	0.70	6.80	13.2
PCB118	13.6	12.5	10.3	0.40	7.33	13.3
PCB128	1.77	2.15	1.79	0.12	1.22	2.36
PCB138	13.5	12.8	9.20	1.40	5.50	12.9
PCB146	1.86	1.89	1.92	0.16	1.28	2.56
PCB149	6.94	7.41	7.01	0.28	4.98	9.04
PCB151	2.02	1.97	1.86	0.16	1.24	2.49
PCB153	12.7	12.1	12.3	0.80	8.43	16.2
PCB156	0.694	0.815	0.72	0.08	0.46	0.98
PCB158	0.979	0.999	1.00	0.096	0.65	1.34
PCB170*	1.30 U	1.08 U	0.27	0.034	0.17	0.37
PCB180	1.26	1.26	1.17	0.10	0.78	1.56
PCB183	1.39	1.31	1.25	0.03	0.91	1.59
PCB187	2.87	2.94	2.94	0.15	2.06	3.83

SDG = Sample Delivery Group, also called analytical batch

*The "True Value" for PCB170 is less than five times the MDL. The acceptance criteria do not apply.

Shaded values are outside the acceptance criteria

TABLE 1B
Summary of Standard Reference Material Results: Statistical Evaluation

STANDARD REFERENCE MATERIAL 1974b - Organic in Mussel Tissue (Mytilus edulis)
Summary of Analytical Performance

PCB Congene	True Value ng/g	Uncertainty	+/- 25% Limits ng/g From	+/- 25% Limits ng/g To	Average Result ng/g	Minimum Result ng/g	Maximum result ng/g	Number of Analyses	Number of Outliers	Average vs. True %D**
PCB18	0.84	0.13	0.50	1.18	NA	1.00 U	1.21 U	2	0	NA
PCB28/31	6.31	0.48	4.25	8.37	16.15	10.50	21.80	2	2	-155.9
PCB44	3.85	0.2	2.69	5.01	4.31	4.13	4.48	2	0	-11.82
PCB49	5.66	0.23	4.02	7.31	4.54	4.45	4.63	2	0	19.8
PCB52	6.26	0.37	4.33	8.20	6.67	6.56	6.78	2	0	-6.55
PCB66	6.37	0.37	4.41	8.33	6.79	6.60	6.98	2	0	-6.59
PCB70	6.01	0.22	4.29	7.73	7.15	6.94	7.36	2	0	-18.97
PCB74	3.55	0.23	2.43	4.67	3.92	3.52	4.32	2	0	-10.4
PCB87	4.33	0.36	2.89	5.77	5.58	5.26	5.90	2	1	-28.9
PCB95	6.04	0.36	4.17	7.91	6.82	6.53	7.10	2	0	-12.83
PCB99	5.92	0.27	4.17	7.67	5.86	5.78	5.93	2	0	1.10
PCB101	10.70	1.10	6.93	14.48	12.55	11.80	13.30	2	0	-17.29
PCB105	4.00	0.18	2.82	5.18	4.27	4.15	4.39	2	0	-6.75
PCB110	10.0	0.70	6.80	13.20	9.92	9.63	10.20	2	0	0.85
PCB118	10.3	0.4	7.33	13.28	13.05	12.50	13.60	2	1	-26.70
PCB128	1.79	0.12	1.22	2.36	1.96	1.77	2.15	2	0	-9.50
PCB138	9.20	1.40	5.50	12.90	13.15	12.80	13.50	2	1	-42.9
PCB146	1.92	0.16	1.28	2.56	1.88	1.86	1.89	2	0	2.34
PCB149	7.01	0.28	4.98	9.04	7.18	6.94	7.41	2	0	-2.4
PCB151	1.86	0.16	1.24	2.49	2.00	1.97	2.02	2	0	-7.26
PCB153	12.3	0.80	8.43	16.18	12.40	12.10	12.70	2	0	-0.81
PCB156	0.72	0.08	0.46	0.98	0.75	0.69	0.82	2	0	-5.08
PCB158	1.00	0.096	0.65	1.34	0.99	0.98	1.00	2	0	1.0
PCB170*	0.27	0.034	0.17	0.37	NA	1.08 U	1.30 U	2	0	NA
PCB180	1.17	0.1	0.78	1.56	1.26	1.26	1.26	2	0	-7.69
PCB183	1.25	0.03	0.91	1.59	1.35	1.31	1.39	2	0	-8.00
PCB187	2.94	0.15	2.06	3.83	2.91	2.87	2.94	2	0	1.19

*The "True Value" for PCB170 is less than five times the MDL. The acceptance criteria do not apply.

**%D = [(True Value - Average Result) / True Value] X 100

ATTACHMENT A

Data Validation Reports by Sample Data Group (SDG)

DATA VALIDATION REPORT - FULL REVIEW
Hudson River
Polychlorinated Biphenyl Congeners
SDG: 0307080

This report documents the review of analytical data from the analysis of bat brain samples and the associated laboratory quality control samples. The following table is a list of samples reviewed.

Field ID	Laboratory ID	Common Name
Sarato 16	0307080-01	Bat brain tissue
Black R 16	0307080-02	Bat brain tissue
Sarato 7	0307080-03	Bat brain tissue
Sarato 15	0307080-04	Bat brain tissue
Sarato 5	0307080-05	Bat brain tissue
Sarato 13	0307080-06	Bat brain tissue
Sarato 9	0307080-07	Bat brain tissue
W Birch 6	0307080-08	Bat brain tissue
Sarato 12	0307080-09	Bat brain tissue
Sarato 22	0307080-10	Bat brain tissue
Sarato 4	0307080-11	Bat brain tissue
Sarato 21	0307080-12	Bat brain tissue
Sarato 20	0307080-13	Bat brain tissue
Sarato 19	0307080-14	Bat brain tissue
Sarato 1	0307080-15	Bat brain tissue

I. DATA PACKAGE COMPLETENESS

The laboratory followed adequate corrective action processes, and all anomalies were discussed in the case narrative. The laboratory submitted all required deliverables.

Due to limited sample size, percent lipids and percent moisture analyses were not performed.

II. TECHNICAL DATA VALIDATION

The quality control (QC) requirements that were reviewed are listed below.

	GC/MS Instrument Performance Check	2	Standard Reference Material (SRM)
	Initial Calibration (ICAL)	1	Laboratory Duplicate
	Continuing Calibration (CCAL)		Internal Standards
	Blanks		Compound Identification
	Surrogate Compounds		Calculation Verification
1	Matrix Spike (MS)	1	Reporting Limits and Sample Results
2	Laboratory Control Samples (LCS)		EDD Transcription Check

[1] ***Quality control results are discussed below, but no data were qualified.***
[2] ***Quality control outliers that impact the reported data were noted. Data qualifiers were issued as discussed below.***

Matrix Spike

No matrix spike analysis was performed due to limited sample size.

Laboratory Control Sample (LCS)

The percent recovery (%R) values for PCB95, PCB118, and PCB167 were greater than the upper control limit of 125%. As the outliers indicate a potential high bias, reporting limits were judged to be unaffected. PCB95 was not detected in any sample, and therefore no qualifiers were assigned to this analyte. Positive values for PCB118 and PCB167 were estimated (J-10).

Standard Reference Material (SRM)

The reported concentrations for PCB31/28, PCB87, PCB118, and PCB138 were greater than the upper control limit of the acceptance window [±20% of the 95% confidence interval]. Since the recoveries of these compounds were acceptable in the LCS analyses, no action was taken unless the reported concentrations were also outside a wider control limit of ±25% of the 95% confidence interval.

The reported values for these PCB were also greater than the +25% upper control limit. Positive values for these congeners were estimated (J-12) in all samples. As the SRM results indicate a potential high bias, no action was taken if the congeners were not detected.

Laboratory Duplicate

No laboratory duplicate analysis was performed due to limited sample size.

Reporting Limits and Sample Results

PCB congener response factors are generated for each target congener during the calibration process. The relative area of a peak is divided by the appropriate response factor to calculate the concentration of the congener. For the homologue groups (monochlorobiphenyl, dichlorobiphenyl, etc.), a representative response factor is used. For example, the response factor for PCB29 is used as the representative response factor for all trichlorobiphenyls.

In several samples, the reported trichlorobiphenyl and/or heptachlorobiphenyl homologue group total values are less than the sum of the individual congeners. This occurs because the representative response factor selected for homologue calculation is sufficiently different from the target congener response factors generated during the calibration. For example, the response factor for the PCB31/28 co-elution (detected in most samples) is lower than the trichlorobiphenyl representative response factor. Since the areas are divided by the response factors, this results in a lower concentration for the total trichlorobiphenyls, even if PCB31/28 is the only detected congener in the chlorination level.

Unless all 209 congeners are calibrated, any reported total for a chlorination level will have some inherent variability. The differences noted for the Total Homologue values are all less than 3.0%. This is within the variability of the method, thus no action was taken.

For some low concentration homologue group totals, the homologue value was changed from a non-detect (U-flag) value to a positive result. The positive result is the sum of all detected congeners in the level of chlorination. This revision was necessary because the automated routine that the instrument uses to calculate the value for a homologue total does not always differentiate between values near the detection limit and the detection limit, as these low concentration results are near the noise level for the homologue. After the corrections, all homologue group values are now either equal to or greater than the sum of the individual congeners because of the addition of non-target congeners for that chlorination level.

III Overall Assessment

As was determined by this evaluation, the laboratory followed the specified analytical methods. Accuracy was acceptable, as demonstrated by the surrogate, LCS, and SRM %R values, with the exceptions noted above. Precision was not evaluated.

Data were estimated due to LCS and SRM %R outliers.

All data, as qualified, are acceptable for use.

DATA VALIDATION REPORT - SUMMARY REVIEW
Hudson River
Polychlorinated Biphenyl Congeners
SDG: 0307081

This report documents the review of analytical data from the analysis of bat brain samples and the associated laboratory quality control samples. The following table is a list of samples reviewed.

Field ID	Laboratory ID	Common Name
Sarato 10	0307081-01	Bat brain tissue
Sarato 28	0307081-02	Bat brain tissue
W Birch 13	0307081-03	Bat brain tissue
Black R 7	0307081-04	Bat brain tissue
Black R 15	0307081-05	Bat brain tissue
Sarato 14	0307081-06	Bat brain tissue
Sarato 17	0307081-07	Bat brain tissue
W Birch 2	0307081-08	Bat brain tissue
W Birch 12	0307081-09	Bat brain tissue
W Birch 9	0307081-10	Bat brain tissue
W Birch 10	0307081-11	Bat brain tissue
Black R 5	0307081-12	Bat brain tissue
W Birch 4	0307081-13	Bat brain tissue
Sarato 18	0307081-14	Bat brain tissue
Black R 6	0307081-15	Bat brain tissue
Sarato 23	0307081-16	Bat brain tissue

I. DATA PACKAGE COMPLETENESS

The laboratory followed adequate corrective action processes, and all anomalies were discussed in the case narrative. The laboratory submitted all required deliverables.

Due to limited sample size, percent lipids and percent moisture analyses were not performed.

II. TECHNICAL DATA VALIDATION

The quality control (QC) requirements that were reviewed are listed below.

	GC/MS Instrument Performance Check	2	Standard Reference Material (SRM)
	Initial Calibration (ICAL)	1	Laboratory Duplicate
2	Continuing Calibration (CCAL)		Internal Standards
	Blanks		Compound Identification
	Surrogate Compounds		Calculation Verification
1	Matrix Spike (MS)	2	Reporting Limits and Sample Results
2	Laboratory Control Samples (LCS)		EDD Transcription Check

[1] ***Quality control results are discussed below, but no data were qualified.***
[2] ***Quality control outliers that impact the reported data were noted. Data qualifiers were issued as discussed below.***

Continuing Calibration (CCAL)

The percent difference (%D) value for PCB31/28 (at 22.4%) was greater than the control limit of ±20% in the end-of-sequence CCAL analyzed January 23, 2004 at 14:02. The value for PCB31/28 was estimated (J-5B) in the associated sample.

Matrix Spike

No matrix spike analysis was performed due to limited sample size.

Laboratory Control Sample (LCS)

The percent recovery (%R) values for PCB70 and PCB95 were greater than the upper control limit of 125%. As the outliers indicate a potential high bias, reporting limits were judged to be unaffected. Positive values for these analytes were estimated (J-10).

Standard Reference Material (SRM)

The reported concentrations for PCB31/28 and PCB138 were greater than the upper control limits of the acceptance window [±20% of the 95% confidence interval]. Since the recoveries of these compounds were acceptable in the LCS analyses, no action was taken unless the reported concentrations were also outside a wider control limit of ±25% of the 95% confidence interval.

The reported value for PCB31/28 was also greater than 25% of the 95% confidence interval. Positive values for PCB31/28 were estimated (J-12) in all samples. The reported value for PCB138 was within 25% of the 95% confidence interval, and no action was taken for this outlier.

Laboratory Duplicate

No laboratory duplicate analysis was performed due to limited sample size.

Reporting Limits and Sample Results

In Samples Sarato 10, Sarato 17, Sarato 18, and Sarato 23 the values reported for the trichlorobiphenyl and heptachlorobiphenyl homologue groups are less than the sum of the individual congeners reported from these chlorination levels.

In Samples Sarato 28 and W Birch 4 the values reported for the trichlorobiphenyl homologue group are less than the sum of the individual congeners reported from this chlorination level.

PCB congener response factors are generated for each target congener during the calibration process. The relative area of a peak is divided by the appropriate response factor to calculate the concentration of the congener. For the homologue groups (monochlorobiphenyl, dichlorobiphenyl, etc.), a representative response factor is used. For example, the response factor for PCB29 is used as the representative response factor for all trichlorobiphenyls.

In the cases above, where the reported trichlorobiphenyl or heptachlorobiphenyl homologue group total values are less than the sum of the individual congeners, the representative response factor selected for homologue calculation is sufficiently different from the target congener response factors generated during the calibration. For example, the response factor for the PCB31/28 co-elution (detected in most samples) is lower than the trichlorobiphenyl representative response factor. Since the areas are divided by the response factors, this results in a lower concentration for the total trichlorobiphenyls, even if PCB31/28 is the only detected congener in the chlorination level.

Unless all 209 congeners are calibrated, any reported total for a chlorination level will have some inherent variability. For example, for the trichlorobiphenyl and heptachlorobiphenyl results, the greatest difference between the calculated (by summing the congeners) and reported result is 2.6% (for values with concentrations greater than the reporting limit). The differences noted for the Total Homologue values are all less than 2.7%. This is within the variability of the method, thus no action was taken.

III Overall Assessment

As was determined by this evaluation, the laboratory followed the specified analytical methods. Accuracy was acceptable, as demonstrated by the surrogate, LCS, and SRM %R values, with the exceptions noted above. Precision was not evaluated.

Data were estimated due to a calibration %D outlier, and LCS and SRM %R outliers.

All data, as qualified, are acceptable for use.

Appendix E

Bat Brain Tissue Data Sheets

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl2-BZ#8	1.91 U	µg/Kg	1.91	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl3-BZ#18	4.24 U	µg/Kg	4.24	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl3-BZ#31/#28	30.6 U	µg/Kg	30.6	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#44	2.91 U	µg/Kg	2.91	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#45	3.89 U	µg/Kg	3.89	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#47	36.3	µg/Kg	4.93	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#49	2.42 U	µg/Kg	2.42	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#52	2.75 U	µg/Kg	2.75	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#56	1.44 U	µg/Kg	1.44	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#66	1.93 U	µg/Kg	1.93	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#70	1.69 U	µg/Kg	1.69	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#74	10.8 J	µg/Kg	2.98	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#77	2.66 U	µg/Kg	2.66	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl4-BZ#81	2.64 U	µg/Kg	2.64	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#87	5.55 J	µg/Kg	1.18	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#95	.910 U	µg/Kg	.910	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#99	21.5	µg/Kg	.622	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#101	1.78 U	µg/Kg	1.78	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#105	5.66	µg/Kg	.866	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#110	1.69 U	µg/Kg	1.69	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#114	.777 U	µg/Kg	.777	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#118	21.6 J	µg/Kg	1.82	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#123	1.53 U	µg/Kg	1.53	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl5-BZ#126	1.36 U	µg/Kg	1.36	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#128	1.22 U	µg/Kg	1.22	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#138	16.1 J	µg/Kg	1.75	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#146	1.11 U	µg/Kg	1.11	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#149	1.42 U	µg/Kg	1.42	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#151	2.35 U	µg/Kg	2.35	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#153	23.8	µg/Kg	2.02	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#156	2.66 J	µg/Kg	1.44	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#157	1.13 U	µg/Kg	1.13	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#158	.955 U	µg/Kg	.955	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#167	1.22 U	µg/Kg	1.22	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl6-BZ#169	3.93 U	µg/Kg	3.93	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl7-BZ#170	6.00 J	µg/Kg	4.58	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl7-BZ#174	.933 U	µg/Kg	.933	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl7-BZ#177	1.31 U	µg/Kg	1.31	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl7-BZ#180	6.00 J	µg/Kg	1.95	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl7-BZ#183	.888 U	µg/Kg	.888	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl7-BZ#189	1.75 U	µg/Kg	1.75	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl7-BZ#187	.977 U	µg/Kg	.977	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl8-BZ#194	2.04 U	µg/Kg	2.04	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl8-BZ#195	2.02 U	µg/Kg	2.02	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl8-BZ#201	1.78 U	µg/Kg	1.78	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl9-BZ#206	2.38 U	µg/Kg	2.38	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Cl10-BZ#209	1.11 U	µg/Kg	1.11	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Monochlorobiphenyls	65.3 U	µg/Kg	65.3	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Dichlorobiphenyls	1.91 U	µg/Kg	1.91	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Trichlorobiphenyls	30.6 U	µg/Kg	30.6	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Tetrachlorobiphenyls	62.7	µg/Kg	4.93	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Pentachlorobiphenyls	117.	µg/Kg	1.82	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Hexachlorobiphenyls	55.0	µg/Kg	3.93	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Heptachlorobiphenyls	15.9	µg/Kg	4.58	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Octachlorobiphenyls	2.02 U	µg/Kg	2.02	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Nonachlorobiphenyls	2.38 U	µg/Kg	2.38	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Decachlorobiphenyl	1.11 U	µg/Kg	1.11	µg/Kg
7/25/2001	Sarato 16	613520	4761876	Big Brown Bat	Female	0307080-01	Total Homologues	252.	µg/Kg	65.3	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl2-BZ#8	1.83 U	µg/Kg	1.83	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl3-BZ#18	4.06 U	µg/Kg	4.06	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl3-BZ#31/#28	29.3 U	µg/Kg	29.3	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#44	2.79 U	µg/Kg	2.79	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#45	3.72 U	µg/Kg	3.72	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#47	4.72 U	µg/Kg	4.72	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#49	2.32 U	µg/Kg	2.32	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#52	2.64 U	µg/Kg	2.64	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#56	1.38 U	µg/Kg	1.38	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#66	1.85 U	µg/Kg	1.85	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#70	1.62 U	µg/Kg	1.62	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#74	2.85 U	µg/Kg	2.85	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#77	2.55 U	µg/Kg	2.55	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl4-BZ#81	2.53 U	µg/Kg	2.53	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#87	1.13 U	µg/Kg	1.13	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#95	.873 U	µg/Kg	.873	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#99	.596 U	µg/Kg	.596	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#101	1.70 U	µg/Kg	1.70	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#105	.830 U	µg/Kg	.830	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#110	1.62 U	µg/Kg	1.62	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#114	.745 U	µg/Kg	.745	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#118	1.74 U	µg/Kg	1.74	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#123	1.47 U	µg/Kg	1.47	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl5-BZ#126	1.30 U	µg/Kg	1.30	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#128	1.17 U	µg/Kg	1.17	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#138	1.68 U	µg/Kg	1.68	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#146	1.06 U	µg/Kg	1.06	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#149	1.36 U	µg/Kg	1.36	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#151	2.26 U	µg/Kg	2.26	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#153	1.94 U	µg/Kg	1.94	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#156	1.38 U	µg/Kg	1.38	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#157	1.08 U	µg/Kg	1.08	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#158	.915 U	µg/Kg	.915	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#167	1.17 U	µg/Kg	1.17	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl6-BZ#169	3.77 U	µg/Kg	3.77	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl7-BZ#170	4.38 U	µg/Kg	4.38	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl7-BZ#174	.894 U	µg/Kg	.894	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl7-BZ#177	1.25 U	µg/Kg	1.25	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl7-BZ#180	1.87 U	µg/Kg	1.87	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl7-BZ#183	.851 U	µg/Kg	.851	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl7-BZ#189	1.68 U	µg/Kg	1.68	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl7-BZ#187	.936 U	µg/Kg	.936	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl8-BZ#194	1.96 U	µg/Kg	1.96	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl8-BZ#195	1.94 U	µg/Kg	1.94	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl8-BZ#201	1.70 U	µg/Kg	1.70	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl9-BZ#206	2.28 U	µg/Kg	2.28	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Cl10-BZ#209	1.06 U	µg/Kg	1.06	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Monochlorobiphenyls	62.6 U	µg/Kg	62.6	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Dichlorobiphenyls	1.83 U	µg/Kg	1.83	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Trichlorobiphenyls	29.3 U	µg/Kg	29.3	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Tetrachlorobiphenyls	4.72 U	µg/Kg	4.72	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Pentachlorobiphenyls	1.74 U	µg/Kg	1.74	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Hexachlorobiphenyls	3.77 U	µg/Kg	3.77	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Heptachlorobiphenyls	4.38 U	µg/Kg	4.38	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Octachlorobiphenyls	1.94 U	µg/Kg	1.94	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Nonachlorobiphenyls	2.28 U	µg/Kg	2.28	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Decachlorobiphenyl	1.06 U	µg/Kg	1.06	µg/Kg
7/21/2001	Black R 16	470549	4836810	Little Brown Bat	Female	0307080-02	Total Homologues	62.6 U	µg/Kg	62.6	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl2-BZ#8	3.22 U	µg/Kg	3.22	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl3-BZ#18	7.14 U	µg/Kg	7.14	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl3-BZ#31/#28	58.7 J	µg/Kg	51.5	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#44	4.90 U	µg/Kg	4.90	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#45	6.54 U	µg/Kg	6.54	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#47	75.0	µg/Kg	8.30	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#49	4.08 U	µg/Kg	4.08	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#52	13.5 J	µg/Kg	4.64	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#56	10.1 J	µg/Kg	2.43	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#66	58.9	µg/Kg	3.25	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#70	5.61 J	µg/Kg	2.84	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#74	41.9	µg/Kg	5.01	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#77	4.49 U	µg/Kg	4.49	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl4-BZ#81	4.45 U	µg/Kg	4.45	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#87	12.2 J	µg/Kg	1.98	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#95	1.53 U	µg/Kg	1.53	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#99	75.5	µg/Kg	1.05	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#101	5.24 J	µg/Kg	2.99	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#105	22.8	µg/Kg	1.46	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#110	2.84 U	µg/Kg	2.84	µg/Kg

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J/UJ=Estimated result or detection limit

[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#114	1.31 U	µg/Kg	1.31	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#118	111. J	µg/Kg	3.07	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#123	2.58 U	µg/Kg	2.58	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl5-BZ#126	2.28 U	µg/Kg	2.28	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#128	4.30 J	µg/Kg	2.06	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#138	59.5 J	µg/Kg	2.95	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#146	3.93 J	µg/Kg	1.87	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#149	2.39 U	µg/Kg	2.39	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#151	3.96 U	µg/Kg	3.96	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#153	121.	µg/Kg	3.40	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#156	9.91 J	µg/Kg	2.43	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#157	1.91 U	µg/Kg	1.91	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#158	1.61 U	µg/Kg	1.61	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#167	5.24 J	µg/Kg	2.06	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl6-BZ#169	6.62 U	µg/Kg	6.62	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl7-BZ#170	26.4 J	µg/Kg	7.70	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl7-BZ#174	1.57 U	µg/Kg	1.57	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl7-BZ#177	2.21 U	µg/Kg	2.21	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl7-BZ#180	38.1	µg/Kg	3.29	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl7-BZ#183	5.98 J	µg/Kg	1.50	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl7-BZ#189	2.95 U	µg/Kg	2.95	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl7-BZ#187	5.42 J	µg/Kg	1.64	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl8-BZ#194	8.98 J	µg/Kg	3.44	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl8-BZ#195	3.40 U	µg/Kg	3.40	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl8-BZ#201	4.67 J	µg/Kg	2.99	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl9-BZ#206	4.00 U	µg/Kg	4.00	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Cl10-BZ#209	1.87 U	µg/Kg	1.87	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Monochlorobiphenyls	110. U	µg/Kg	110.	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Dichlorobiphenyls	3.22 U	µg/Kg	3.22	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Trichlorobiphenyls	58.7 J	µg/Kg	51.5	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Tetrachlorobiphenyls	267.	µg/Kg	8.30	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Pentachlorobiphenyls	411.	µg/Kg	3.07	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Hexachlorobiphenyls	241.	µg/Kg	6.62	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Heptachlorobiphenyls	75.5	µg/Kg	7.70	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Octachlorobiphenyls	45.3	µg/Kg	3.44	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Nonachlorobiphenyls	4.00 U	µg/Kg	4.00	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Decachlorobiphenyl	1.87 U	µg/Kg	1.87	µg/Kg
7/23/2001	Sarato 7	613520	4761876	Little Brown Bat	Female	0307080-03	Total Homologues	1080.	µg/Kg	110.	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl2-BZ#8	2.99 U	µg/Kg	2.99	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl3-BZ#18	6.65 U	µg/Kg	6.65	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl3-BZ#31/#28	71.5 J	µg/Kg	47.9	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#44	4.56 U	µg/Kg	4.56	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#45	6.09 U	µg/Kg	6.09	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#47	67.9	µg/Kg	7.73	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#49	4.70 J	µg/Kg	3.79	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#52	38.8	µg/Kg	4.32	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#56	7.31 J	µg/Kg	2.26	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#66	28.5	µg/Kg	3.03	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#70	5.05 J	µg/Kg	2.64	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#74	20.4 J	µg/Kg	4.66	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#77	4.18 U	µg/Kg	4.18	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl4-BZ#81	4.14 U	µg/Kg	4.14	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#87	5.22 J	µg/Kg	1.84	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#95	1.43 U	µg/Kg	1.43	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#99	13.4	µg/Kg	.975	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#101	3.65 J	µg/Kg	2.78	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#105	4.70 J	µg/Kg	1.36	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#110	2.64 U	µg/Kg	2.64	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#114	1.22 U	µg/Kg	1.22	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#118	17.6 J	µg/Kg	2.85	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#123	2.40 U	µg/Kg	2.40	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl5-BZ#126	2.12 U	µg/Kg	2.12	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#128	1.91 U	µg/Kg	1.91	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#138	13.4 J	µg/Kg	2.75	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#146	3.31 J	µg/Kg	1.74	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#149	2.23 U	µg/Kg	2.23	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#151	3.69 U	µg/Kg	3.69	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#153	11.7 J	µg/Kg	3.17	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#156	2.26 U	µg/Kg	2.26	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#157	1.78 U	µg/Kg	1.78	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#158	1.50 U	µg/Kg	1.50	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#167	1.91 U	µg/Kg	1.91	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl6-BZ#169	6.16 U	µg/Kg	6.16	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl7-BZ#170	7.17 U	µg/Kg	7.17	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl7-BZ#174	1.46 U	µg/Kg	1.46	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl7-BZ#177	2.05 U	µg/Kg	2.05	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl7-BZ#180	4.87 J	µg/Kg	3.06	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl7-BZ#183	1.74 J	µg/Kg	1.39	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl7-BZ#189	2.75 U	µg/Kg	2.75	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl7-BZ#187	4.52 J	µg/Kg	1.53	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl8-BZ#194	3.20 U	µg/Kg	3.20	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl8-BZ#195	3.17 U	µg/Kg	3.17	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl8-BZ#201	2.78 U	µg/Kg	2.78	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl9-BZ#206	3.72 U	µg/Kg	3.72	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Cl10-BZ#209	1.74 U	µg/Kg	1.74	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Monochlorobiphenyls	102. U	µg/Kg	102.	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Dichlorobiphenyls	2.99 U	µg/Kg	2.99	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Trichlorobiphenyls	72.0	µg/Kg	47.9	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Tetrachlorobiphenyls	224.	µg/Kg	7.73	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Pentachlorobiphenyls	141.	µg/Kg	2.85	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Hexachlorobiphenyls	36.9	µg/Kg	6.16	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Heptachlorobiphenyls	20.5	µg/Kg	7.17	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Octachlorobiphenyls	16.7	µg/Kg	3.20	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Nonachlorobiphenyls	3.72 U	µg/Kg	3.72	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Decachlorobiphenyl	1.74 U	µg/Kg	1.74	µg/Kg
7/24/2001	Sarato 15	613520	4761876	Little Brown Bat	Female	0307080-04	Total Homologues	511.	µg/Kg	102.	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl2-BZ#8	1.37 U	µg/Kg	1.37	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl3-BZ#18	3.04 U	µg/Kg	3.04	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl3-BZ#31/#28	21.9 U	µg/Kg	21.9	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#44	2.09 U	µg/Kg	2.09	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#45	2.79 U	µg/Kg	2.79	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#47	10.9 J	µg/Kg	3.54	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#49	1.74 U	µg/Kg	1.74	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#52	1.98 U	µg/Kg	1.98	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#56	1.04 U	µg/Kg	1.04	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#66	1.39 U	µg/Kg	1.39	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#70	1.21 U	µg/Kg	1.21	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#74	8.45 J	µg/Kg	2.13	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#77	1.91 U	µg/Kg	1.91	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl4-BZ#81	1.90 U	µg/Kg	1.90	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#87	.845 U	µg/Kg	.845	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#95	.653 U	µg/Kg	.653	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#99	14.1	µg/Kg	.446	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#101	1.27 U	µg/Kg	1.27	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#105	6.14	µg/Kg	.622	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#110	1.21 U	µg/Kg	1.21	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#114	.558 U	µg/Kg	.558	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#118	20.0 J	µg/Kg	1.31	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#123	1.10 U	µg/Kg	1.10	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl5-BZ#126	.972 U	µg/Kg	.972	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#128	.876 U	µg/Kg	.876	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#138	11.2 J	µg/Kg	1.26	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#146	.797 U	µg/Kg	.797	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#149	1.02 U	µg/Kg	1.02	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#151	1.69 U	µg/Kg	1.69	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#153	17.7	µg/Kg	1.45	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#156	1.91 J	µg/Kg	1.04	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#157	.813 U	µg/Kg	.813	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#158	.685 U	µg/Kg	.685	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#167	.876 U	µg/Kg	.876	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl6-BZ#169	2.82 U	µg/Kg	2.82	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl7-BZ#170	4.06 J	µg/Kg	3.28	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl7-BZ#174	.669 U	µg/Kg	.669	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl7-BZ#177	.940 U	µg/Kg	.940	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl7-BZ#180	6.06 J	µg/Kg	1.40	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl7-BZ#183	1.75 J	µg/Kg	.637	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl7-BZ#189	1.26 U	µg/Kg	1.26	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl7-BZ#187	.701 U	µg/Kg	.701	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl8-BZ#194	1.47 U	µg/Kg	1.47	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl8-BZ#195	1.45 U	µg/Kg	1.45	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl8-BZ#201	1.27 U	µg/Kg	1.27	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl9-BZ#206	1.70 U	µg/Kg	1.70	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Cl10-BZ#209	.797 U	µg/Kg	.797	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Monochlorobiphenyls	46.8 U	µg/Kg	46.8	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Dichlorobiphenyls	1.37 U	µg/Kg	1.37	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Trichlorobiphenyls	21.9 U	µg/Kg	21.9	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Tetrachlorobiphenyls	25.9	µg/Kg	3.54	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Pentachlorobiphenyls	95.4	µg/Kg	1.31	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Hexachlorobiphenyls	52.0	µg/Kg	2.82	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Heptachlorobiphenyls	10.8	µg/Kg	3.28	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Octachlorobiphenyls	1.47 U	µg/Kg	1.47	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Nonachlorobiphenyls	1.70 U	µg/Kg	1.70	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Decachlorobiphenyl	.797 U	µg/Kg	.797	µg/Kg
7/23/2001	Sarato 5	613520	4761876	Big Brown Bat	Female	0307080-05	Total Homologues	184.	µg/Kg	46.8	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl2-BZ#8	5.70 U	µg/Kg	5.70	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl3-BZ#18	12.7 U	µg/Kg	12.7	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl3-BZ#31/#28	91.3 U	µg/Kg	91.3	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#44	8.69 U	µg/Kg	8.69	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#45	11.6 U	µg/Kg	11.6	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#47	57.0 J	µg/Kg	14.7	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#49	7.23 U	µg/Kg	7.23	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#52	18.6 J	µg/Kg	8.22	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#56	8.96 J	µg/Kg	4.31	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#66	37.2	µg/Kg	5.77	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#70	5.04 U	µg/Kg	5.04	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#74	27.9 J	µg/Kg	8.89	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#77	7.96 U	µg/Kg	7.96	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl4-BZ#81	7.89 U	µg/Kg	7.89	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#87	3.52 U	µg/Kg	3.52	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#95	2.72 U	µg/Kg	2.72	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#99	17.2	µg/Kg	1.86	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#101	8.62 J	µg/Kg	5.31	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#105	8.62 J	µg/Kg	2.59	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#110	5.04 U	µg/Kg	5.04	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#114	2.32 U	µg/Kg	2.32	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#118	32.2 J	µg/Kg	5.44	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#123	4.58 U	µg/Kg	4.58	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl5-BZ#126	4.05 U	µg/Kg	4.05	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#128	3.65 U	µg/Kg	3.65	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#138	14.9 J	µg/Kg	5.24	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#146	3.32 U	µg/Kg	3.32	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#149	4.24 U	µg/Kg	4.24	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#151	7.03 U	µg/Kg	7.03	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#153	14.9 J	µg/Kg	6.04	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#156	4.31 U	µg/Kg	4.31	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#157	3.38 U	µg/Kg	3.38	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#158	2.85 U	µg/Kg	2.85	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#167	3.65 U	µg/Kg	3.65	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl6-BZ#169	11.7 U	µg/Kg	11.7	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl7-BZ#170	13.7 U	µg/Kg	13.7	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl7-BZ#174	2.79 U	µg/Kg	2.79	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl7-BZ#177	3.91 U	µg/Kg	3.91	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl7-BZ#180	5.97 J	µg/Kg	5.84	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl7-BZ#183	2.65 U	µg/Kg	2.65	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl7-BZ#189	5.24 U	µg/Kg	5.24	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl7-BZ#187	2.92 U	µg/Kg	2.92	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl8-BZ#194	6.10 U	µg/Kg	6.10	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl8-BZ#195	6.04 U	µg/Kg	6.04	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl8-BZ#201	5.31 U	µg/Kg	5.31	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl9-BZ#206	7.10 U	µg/Kg	7.10	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Cl10-BZ#209	3.32 U	µg/Kg	3.32	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Monochlorobiphenyls	195. U	µg/Kg	195.	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Dichlorobiphenyls	5.70 U	µg/Kg	5.70	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Trichlorobiphenyls	91.3 U	µg/Kg	91.3	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Tetrachlorobiphenyls	186.	µg/Kg	14.7	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Pentachlorobiphenyls	222.	µg/Kg	5.44	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Hexachlorobiphenyls	40.8	µg/Kg	11.7	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Heptachlorobiphenyls	13.7 U	µg/Kg	13.7	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Octachlorobiphenyls	6.10 U	µg/Kg	6.10	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Nonachlorobiphenyls	7.10 U	µg/Kg	7.10	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Decachlorobiphenyl	3.32 U	µg/Kg	3.32	µg/Kg
7/24/2001	Sarato 13	613520	4761876	Little Brown Bat	Female	0307080-06	Total Homologues	503.	µg/Kg	195.	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl2-BZ#8	2.95 U	µg/Kg	2.95	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl3-BZ#18	6.55 U	µg/Kg	6.55	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl3-BZ#31/#28	47.2 U	µg/Kg	47.2	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#44	4.49 U	µg/Kg	4.49	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#45	6.00 U	µg/Kg	6.00	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#47	8.05 J	µg/Kg	7.61	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#49	3.74 U	µg/Kg	3.74	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#52	4.25 U	µg/Kg	4.25	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#56	2.23 U	µg/Kg	2.23	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#66	2.98 U	µg/Kg	2.98	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#70	2.74 J	µg/Kg	2.60	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#74	4.63 J	µg/Kg	4.59	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#77	4.11 U	µg/Kg	4.11	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl4-BZ#81	4.08 U	µg/Kg	4.08	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#87	1.82 U	µg/Kg	1.82	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#95	1.40 U	µg/Kg	1.40	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#99	5.31	µg/Kg	.960	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#101	2.74 U	µg/Kg	2.74	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#105	1.34 U	µg/Kg	1.34	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#110	2.60 U	µg/Kg	2.60	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#114	1.20 U	µg/Kg	1.20	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#118	6.00 J	µg/Kg	2.81	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#123	2.37 U	µg/Kg	2.37	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl5-BZ#126	2.09 U	µg/Kg	2.09	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#128	1.88 U	µg/Kg	1.88	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#138	4.97 J	µg/Kg	2.71	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#146	1.71 U	µg/Kg	1.71	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#149	2.19 U	µg/Kg	2.19	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#151	3.63 U	µg/Kg	3.63	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#153	3.12 U	µg/Kg	3.12	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#156	2.23 U	µg/Kg	2.23	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#157	1.75 U	µg/Kg	1.75	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#158	1.47 U	µg/Kg	1.47	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#167	1.88 U	µg/Kg	1.88	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl6-BZ#169	6.07 U	µg/Kg	6.07	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl7-BZ#170	7.06 U	µg/Kg	7.06	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl7-BZ#174	1.44 U	µg/Kg	1.44	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl7-BZ#177	2.02 U	µg/Kg	2.02	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl7-BZ#180	3.02 U	µg/Kg	3.02	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl7-BZ#183	1.37 U	µg/Kg	1.37	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl7-BZ#189	2.71 U	µg/Kg	2.71	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl7-BZ#187	1.51 U	µg/Kg	1.51	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl8-BZ#194	3.15 U	µg/Kg	3.15	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl8-BZ#195	3.12 U	µg/Kg	3.12	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl8-BZ#201	2.74 U	µg/Kg	2.74	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl9-BZ#206	3.67 U	µg/Kg	3.67	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Cl10-BZ#209	1.71 U	µg/Kg	1.71	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Monochlorobiphenyls	101. U	µg/Kg	101.	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Dichlorobiphenyls	2.95 U	µg/Kg	2.95	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Trichlorobiphenyls	47.2 U	µg/Kg	47.2	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Tetrachlorobiphenyls	15.4	µg/Kg	7.61	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Pentachlorobiphenyls	11.3	µg/Kg	2.81	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Hexachlorobiphenyls	4.97 J	µg/Kg	6.07	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Heptachlorobiphenyls	7.06 U	µg/Kg	7.06	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Octachlorobiphenyls	3.15 U	µg/Kg	3.15	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Nonachlorobiphenyls	3.67 U	µg/Kg	3.67	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Decachlorobiphenyl	1.71 U	µg/Kg	1.71	µg/Kg
7/23/2001	Sarato 9	613520	4761876	Big Brown Bat	Female	0307080-07	Total Homologues	31.7 J	µg/Kg	101.	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl2-BZ#8	15.2 U	µg/Kg	15.2	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl3-BZ#18	33.9 U	µg/Kg	33.9	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl3-BZ#31/#28	244. U	µg/Kg	244.	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#44	23.2 U	µg/Kg	23.2	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#45	31.0 U	µg/Kg	31.0	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#47	39.4 U	µg/Kg	39.4	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#49	19.3 U	µg/Kg	19.3	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#52	22.0 U	µg/Kg	22.0	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#56	11.5 U	µg/Kg	11.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#66	15.4 U	µg/Kg	15.4	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#70	13.5 U	µg/Kg	13.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#74	23.8 U	µg/Kg	23.8	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#77	21.3 U	µg/Kg	21.3	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl4-BZ#81	21.1 U	µg/Kg	21.1	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#87	9.40 U	µg/Kg	9.40	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#95	7.27 U	µg/Kg	7.27	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#99	4.97 U	µg/Kg	4.97	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#101	14.2 U	µg/Kg	14.2	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#105	6.92 U	µg/Kg	6.92	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#110	13.5 U	µg/Kg	13.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#114	6.21 U	µg/Kg	6.21	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#118	14.5 U	µg/Kg	14.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#123	12.2 U	µg/Kg	12.2	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl5-BZ#126	10.8 U	µg/Kg	10.8	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#128	9.75 U	µg/Kg	9.75	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#138	14.0 U	µg/Kg	14.0	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#146	8.87 U	µg/Kg	8.87	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#149	11.4 U	µg/Kg	11.4	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#151	18.8 U	µg/Kg	18.8	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#153	16.1 U	µg/Kg	16.1	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#156	11.5 U	µg/Kg	11.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#157	9.04 U	µg/Kg	9.04	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#158	7.63 U	µg/Kg	7.63	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#167	9.75 U	µg/Kg	9.75	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl6-BZ#169	31.4 U	µg/Kg	31.4	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl7-BZ#170	36.5 U	µg/Kg	36.5	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl7-BZ#174	7.45 U	µg/Kg	7.45	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl7-BZ#177	10.5 U	µg/Kg	10.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl7-BZ#180	15.6 U	µg/Kg	15.6	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl7-BZ#183	7.09 U	µg/Kg	7.09	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl7-BZ#189	14.0 U	µg/Kg	14.0	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl7-BZ#187	7.80 U	µg/Kg	7.80	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl8-BZ#194	16.3 U	µg/Kg	16.3	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl8-BZ#195	16.1 U	µg/Kg	16.1	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl8-BZ#201	14.2 U	µg/Kg	14.2	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl9-BZ#206	19.0 U	µg/Kg	19.0	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Cl10-BZ#209	8.87 U	µg/Kg	8.87	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Monochlorobiphenyls	521. U	µg/Kg	521.	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Dichlorobiphenyls	15.2 U	µg/Kg	15.2	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Trichlorobiphenyls	244. U	µg/Kg	244.	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Tetrachlorobiphenyls	39.4 U	µg/Kg	39.4	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Pentachlorobiphenyls	14.5 U	µg/Kg	14.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Hexachlorobiphenyls	31.4 U	µg/Kg	31.4	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Heptachlorobiphenyls	36.5 U	µg/Kg	36.5	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Octachlorobiphenyls	16.3 U	µg/Kg	16.3	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Nonachlorobiphenyls	19.0 U	µg/Kg	19.0	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Decachlorobiphenyl	8.87 U	µg/Kg	8.87	µg/Kg
7/15/2001	W Birch 6	569295	4712892	Little Brown Bat	Female	0307080-08	Total Homologues	521. U	µg/Kg	521.	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl2-BZ#8	3.17 U	µg/Kg	3.17	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl3-BZ#18	7.04 U	µg/Kg	7.04	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl3-BZ#31/#28	50.8 U	µg/Kg	50.8	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#44	4.83 U	µg/Kg	4.83	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#45	6.45 U	µg/Kg	6.45	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#47	364.	µg/Kg	8.18	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#49	4.02 U	µg/Kg	4.02	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#52	13.1 J	µg/Kg	4.57	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#56	21.2	µg/Kg	2.40	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#66	106.	µg/Kg	3.21	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#70	2.80 U	µg/Kg	2.80	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#74	81.8	µg/Kg	4.94	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#77	4.42 U	µg/Kg	4.42	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl4-BZ#81	4.39 U	µg/Kg	4.39	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#87	44.0 J	µg/Kg	1.95	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#95	1.51 U	µg/Kg	1.51	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#99	128.	µg/Kg	1.03	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#101	18.2	µg/Kg	2.95	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#105	49.7	µg/Kg	1.44	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#110	2.80 U	µg/Kg	2.80	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#114	4.61 J	µg/Kg	1.29	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#118	148. J	µg/Kg	3.02	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#123	2.54 U	µg/Kg	2.54	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl5-BZ#126	2.25 U	µg/Kg	2.25	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#128	11.6	µg/Kg	2.03	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#138	123. J	µg/Kg	2.91	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#146	18.4	µg/Kg	1.84	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#149	2.36 U	µg/Kg	2.36	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#151	3.91 U	µg/Kg	3.91	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#153	121.	µg/Kg	3.35	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#156	11.8 J	µg/Kg	2.40	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#157	1.88 U	µg/Kg	1.88	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#158	3.50 J	µg/Kg	1.58	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#167	4.24 J	µg/Kg	2.03	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl6-BZ#169	6.52 U	µg/Kg	6.52	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl7-BZ#170	21.0 J	µg/Kg	7.59	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl7-BZ#174	1.55 U	µg/Kg	1.55	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl7-BZ#177	6.63 J	µg/Kg	2.17	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl7-BZ#180	34.5	µg/Kg	3.24	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl7-BZ#183	6.45 J	µg/Kg	1.47	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl7-BZ#189	2.91 U	µg/Kg	2.91	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl7-BZ#187	25.1	µg/Kg	1.62	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl8-BZ#194	11.6 J	µg/Kg	3.39	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl8-BZ#195	3.35 U	µg/Kg	3.35	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl8-BZ#201	11.2 J	µg/Kg	2.95	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl9-BZ#206	3.94 U	µg/Kg	3.94	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Cl10-BZ#209	1.84 U	µg/Kg	1.84	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Monochlorobiphenyls	108. U	µg/Kg	108.	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Dichlorobiphenyls	3.17 U	µg/Kg	3.17	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Trichlorobiphenyls	43.3 J	µg/Kg	50.8	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Tetrachlorobiphenyls	693.	µg/Kg	8.18	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Pentachlorobiphenyls	639.	µg/Kg	3.02	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Hexachlorobiphenyls	361.	µg/Kg	6.52	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Heptachlorobiphenyls	94.0	µg/Kg	7.59	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Octachlorobiphenyls	48.1	µg/Kg	3.39	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Nonachlorobiphenyls	3.94 U	µg/Kg	3.94	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Decachlorobiphenyl	1.84 U	µg/Kg	1.84	µg/Kg
7/24/2001	Sarato 12	613520	4761876	Little Brown Bat	Male	0307080-09	Total Homologues	1880.	µg/Kg	108.	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl2-BZ#8	2.95 U	µg/Kg	2.95	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl3-BZ#18	6.56 U	µg/Kg	6.56	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl3-BZ#31/#28	60.1 J	µg/Kg	47.3	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#44	4.50 U	µg/Kg	4.50	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#45	6.01 U	µg/Kg	6.01	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#47	106.	µg/Kg	7.62	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#49	4.63 J	µg/Kg	3.74	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#52	17.2 J	µg/Kg	4.26	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#56	20.1	µg/Kg	2.23	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#66	89.8	µg/Kg	2.99	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#70	2.61 U	µg/Kg	2.61	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#74	63.3	µg/Kg	4.60	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#77	4.12 U	µg/Kg	4.12	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl4-BZ#81	4.08 U	µg/Kg	4.08	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#87	24.4 J	µg/Kg	1.82	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#95	1.41 U	µg/Kg	1.41	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#99	119.	µg/Kg	.961	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#101	8.24 J	µg/Kg	2.75	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#105	40.8	µg/Kg	1.34	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#110	3.95 J	µg/Kg	2.61	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#114	1.20 U	µg/Kg	1.20	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#118	151. J	µg/Kg	2.81	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#123	2.37 U	µg/Kg	2.37	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl5-BZ#126	2.09 U	µg/Kg	2.09	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#128	6.69 J	µg/Kg	1.89	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#138	102. J	µg/Kg	2.71	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#146	8.07 J	µg/Kg	1.72	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#149	2.20 U	µg/Kg	2.20	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#151	3.64 U	µg/Kg	3.64	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#153	190.	µg/Kg	3.12	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#156	17.7	µg/Kg	2.23	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#157	1.89 J	µg/Kg	1.75	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#158	3.43 J	µg/Kg	1.48	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#167	8.41 J	µg/Kg	1.89	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl6-BZ#169	6.08 U	µg/Kg	6.08	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl7-BZ#170	40.7	µg/Kg	7.07	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl7-BZ#174	1.44 U	µg/Kg	1.44	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl7-BZ#177	2.02 U	µg/Kg	2.02	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl7-BZ#180	59.6	µg/Kg	3.02	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl7-BZ#183	10.1	µg/Kg	1.37	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl7-BZ#189	2.71 U	µg/Kg	2.71	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl7-BZ#187	8.92	µg/Kg	1.51	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl8-BZ#194	18.5	µg/Kg	3.16	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl8-BZ#195	3.78 J	µg/Kg	3.12	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl8-BZ#201	7.55 J	µg/Kg	2.75	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl9-BZ#206	14.9 J	µg/Kg	3.67	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Cl10-BZ#209	1.72 U	µg/Kg	1.72	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Monochlorobiphenyls	101. U	µg/Kg	101.	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Dichlorobiphenyls	2.95 U	µg/Kg	2.95	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Trichlorobiphenyls	42.7 J	µg/Kg	47.3	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Tetrachlorobiphenyls	386.	µg/Kg	7.62	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Pentachlorobiphenyls	566.	µg/Kg	2.81	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Hexachlorobiphenyls	384.	µg/Kg	6.08	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Heptachlorobiphenyls	106.	µg/Kg	7.07	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Octachlorobiphenyls	53.5	µg/Kg	3.16	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Nonachlorobiphenyls	14.9 J	µg/Kg	3.67	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Decachlorobiphenyl	1.72 U	µg/Kg	1.72	µg/Kg
7/25/2001	Sarato 22	613520	4761876	Little Brown Bat	Female	0307080-10	Total Homologues	1550.	µg/Kg	101.	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl2-BZ#8	3.37 U	µg/Kg	3.37	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl3-BZ#18	7.48 U	µg/Kg	7.48	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl3-BZ#31/#28	128. J	µg/Kg	53.9	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#44	5.13 U	µg/Kg	5.13	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#45	6.85 U	µg/Kg	6.85	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#47	158.	µg/Kg	8.69	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#49	4.27 U	µg/Kg	4.27	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#52	17.8 J	µg/Kg	4.86	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#56	26.8	µg/Kg	2.54	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#66	134.	µg/Kg	3.41	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#70	2.98 U	µg/Kg	2.98	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#74	95.4	µg/Kg	5.25	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#77	4.70 U	µg/Kg	4.70	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl4-BZ#81	4.66 U	µg/Kg	4.66	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#87	20.6 J	µg/Kg	2.08	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#95	1.60 U	µg/Kg	1.60	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#99	65.0	µg/Kg	1.10	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#101	10.8 J	µg/Kg	3.13	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#105	26.4	µg/Kg	1.53	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#110	2.98 U	µg/Kg	2.98	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#114	5.48 J	µg/Kg	1.37	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#118	92.2 J	µg/Kg	3.21	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#123	2.70 U	µg/Kg	2.70	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl5-BZ#126	2.39 U	µg/Kg	2.39	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#128	5.48 J	µg/Kg	2.15	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#138	44.6 J	µg/Kg	3.09	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#146	5.48 J	µg/Kg	1.96	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#149	2.51 U	µg/Kg	2.51	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#151	4.15 U	µg/Kg	4.15	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#153	49.5	µg/Kg	3.56	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#156	4.70 J	µg/Kg	2.54	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#157	2.00 U	µg/Kg	2.00	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#158	2.15 J	µg/Kg	1.68	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#167	2.54 J	µg/Kg	2.15	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl6-BZ#169	6.93 U	µg/Kg	6.93	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl7-BZ#170	11.0 J	µg/Kg	8.07	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl7-BZ#174	1.64 U	µg/Kg	1.64	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl7-BZ#177	2.54 J	µg/Kg	2.31	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl7-BZ#180	14.3 J	µg/Kg	3.45	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl7-BZ#183	3.33 J	µg/Kg	1.57	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl7-BZ#189	3.09 U	µg/Kg	3.09	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl7-BZ#187	6.66 J	µg/Kg	1.72	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl8-BZ#194	9.79 J	µg/Kg	3.60	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl8-BZ#195	3.56 U	µg/Kg	3.56	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl8-BZ#201	4.11 J	µg/Kg	3.13	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl9-BZ#206	13.1 J	µg/Kg	4.19	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Cl10-BZ#209	9.59 J	µg/Kg	1.96	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Monochlorobiphenyls	115. U	µg/Kg	115.	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Dichlorobiphenyls	3.37 U	µg/Kg	3.37	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Trichlorobiphenyls	93.4	µg/Kg	53.9	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Tetrachlorobiphenyls	547.	µg/Kg	8.69	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Pentachlorobiphenyls	400.	µg/Kg	3.21	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Hexachlorobiphenyls	136.	µg/Kg	6.93	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Heptachlorobiphenyls	36.6	µg/Kg	8.07	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Octachlorobiphenyls	26.4	µg/Kg	3.60	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Nonachlorobiphenyls	13.1 J	µg/Kg	4.19	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Decachlorobiphenyl	9.59 J	µg/Kg	1.96	µg/Kg
7/23/2001	Sarato 4	613520	4761876	Little Brown Bat	Female	0307080-11	Total Homologues	1260.	µg/Kg	115.	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl2-BZ#8	3.61 U	µg/Kg	3.61	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl3-BZ#18	8.02 U	µg/Kg	8.02	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl3-BZ#31/#28	58.1 J	µg/Kg	57.8	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#44	5.50 U	µg/Kg	5.50	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#45	7.35 U	µg/Kg	7.35	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#47	237.	µg/Kg	9.32	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#49	4.58 U	µg/Kg	4.58	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#52	15.3 J	µg/Kg	5.21	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#56	19.9	µg/Kg	2.73	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#66	123.	µg/Kg	3.65	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#70	3.19 U	µg/Kg	3.19	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#74	81.4	µg/Kg	5.63	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#77	5.04 U	µg/Kg	5.04	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl4-BZ#81	5.00 U	µg/Kg	5.00	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#87	30.9 J	µg/Kg	2.22	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#95	1.72 U	µg/Kg	1.72	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#99	115.	µg/Kg	1.18	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#101	14.5 J	µg/Kg	3.36	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#105	45.1	µg/Kg	1.64	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#110	3.19 U	µg/Kg	3.19	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#114	4.41 J	µg/Kg	1.47	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#118	141. J	µg/Kg	3.44	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#123	2.90 U	µg/Kg	2.90	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl5-BZ#126	2.56 U	µg/Kg	2.56	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#128	18.3	µg/Kg	2.31	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#138	113. J	µg/Kg	3.32	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#146	16.0	µg/Kg	2.10	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#149	2.69 U	µg/Kg	2.69	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#151	4.45 U	µg/Kg	4.45	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#153	120.	µg/Kg	3.82	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#156	10.7 J	µg/Kg	2.73	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#157	2.14 U	µg/Kg	2.14	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#158	2.94 J	µg/Kg	1.80	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#167	5.67 J	µg/Kg	2.31	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl6-BZ#169	7.43 U	µg/Kg	7.43	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl7-BZ#170	22.7 J	µg/Kg	8.65	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl7-BZ#174	1.76 U	µg/Kg	1.76	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl7-BZ#177	5.25 J	µg/Kg	2.48	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl7-BZ#180	40.1	µg/Kg	3.69	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl7-BZ#183	8.19 J	µg/Kg	1.68	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl7-BZ#189	3.32 U	µg/Kg	3.32	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl7-BZ#187	24.1	µg/Kg	1.85	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl8-BZ#194	15.1 J	µg/Kg	3.86	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl8-BZ#195	3.82 U	µg/Kg	3.82	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl8-BZ#201	21.4	µg/Kg	3.36	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl9-BZ#206	14.5 J	µg/Kg	4.49	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Cl10-BZ#209	2.10 U	µg/Kg	2.10	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Monochlorobiphenyls	123. U	µg/Kg	123.	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Dichlorobiphenyls	3.61 U	µg/Kg	3.61	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Trichlorobiphenyls	50.8 J	µg/Kg	57.8	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Tetrachlorobiphenyls	600.	µg/Kg	9.32	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Pentachlorobiphenyls	628.	µg/Kg	3.44	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Hexachlorobiphenyls	362.	µg/Kg	7.43	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Heptachlorobiphenyls	95.5	µg/Kg	8.65	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Octachlorobiphenyls	68.2	µg/Kg	3.86	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Nonachlorobiphenyls	14.5 J	µg/Kg	4.49	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Decachlorobiphenyl	2.10 U	µg/Kg	2.10	µg/Kg
7/25/2001	Sarato 21	613520	4761876	Little Brown Bat	Female	0307080-12	Total Homologues	1820.	µg/Kg	123.	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl2-BZ#8	4.74 U	µg/Kg	4.74	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl3-BZ#18	10.5 U	µg/Kg	10.5	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl3-BZ#31/#28	76.0 U	µg/Kg	76.0	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#44	7.23 U	µg/Kg	7.23	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#45	9.65 U	µg/Kg	9.65	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#47	130.	µg/Kg	12.3	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#49	6.01 U	µg/Kg	6.01	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#52	19.6 J	µg/Kg	6.84	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#56	14.6 J	µg/Kg	3.58	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#66	79.2	µg/Kg	4.80	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#70	6.90 J	µg/Kg	4.19	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#74	53.8	µg/Kg	7.39	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#77	6.62 U	µg/Kg	6.62	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl4-BZ#81	6.56 U	µg/Kg	6.56	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#87	15.2 J	µg/Kg	2.92	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#95	2.26 U	µg/Kg	2.26	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#99	47.4	µg/Kg	1.54	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#101	8.82 J	µg/Kg	4.41	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#105	19.6	µg/Kg	2.15	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#110	4.19 U	µg/Kg	4.19	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#114	1.93 U	µg/Kg	1.93	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#118	54.9 J	µg/Kg	4.52	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#123	3.81 U	µg/Kg	3.81	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl5-BZ#126	3.36 U	µg/Kg	3.36	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#128	4.69 J	µg/Kg	3.03	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#138	38.6 J	µg/Kg	4.36	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#146	6.07 J	µg/Kg	2.76	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#149	3.53 U	µg/Kg	3.53	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#151	5.85 U	µg/Kg	5.85	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#153	38.3	µg/Kg	5.02	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#156	3.58 U	µg/Kg	3.58	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#157	2.81 U	µg/Kg	2.81	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#158	2.37 U	µg/Kg	2.37	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#167	3.03 U	µg/Kg	3.03	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl6-BZ#169	9.76 U	µg/Kg	9.76	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl7-BZ#170	11.4 U	µg/Kg	11.4	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl7-BZ#174	2.32 U	µg/Kg	2.32	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl7-BZ#177	3.25 U	µg/Kg	3.25	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl7-BZ#180	14.6 J	µg/Kg	4.85	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl7-BZ#183	3.31 J	µg/Kg	2.21	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl7-BZ#189	4.36 U	µg/Kg	4.36	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl7-BZ#187	7.45 J	µg/Kg	2.43	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl8-BZ#194	5.52 J	µg/Kg	5.08	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl8-BZ#195	5.02 U	µg/Kg	5.02	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl8-BZ#201	4.41 U	µg/Kg	4.41	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl9-BZ#206	5.90 U	µg/Kg	5.90	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Cl10-BZ#209	2.76 U	µg/Kg	2.76	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Monochlorobiphenyls	162. U	µg/Kg	162.	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Dichlorobiphenyls	4.74 U	µg/Kg	4.74	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Trichlorobiphenyls	76.0 J	µg/Kg	76.0	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Tetrachlorobiphenyls	394.	µg/Kg	12.3	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Pentachlorobiphenyls	334.	µg/Kg	4.52	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Hexachlorobiphenyls	109.	µg/Kg	9.76	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Heptachlorobiphenyls	43.8	µg/Kg	11.4	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Octachlorobiphenyls	5.52	µg/Kg	5.08	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Nonachlorobiphenyls	5.90 U	µg/Kg	5.90	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Decachlorobiphenyl	2.76 U	µg/Kg	2.76	µg/Kg
7/25/2001	Sarato 20	613520	4761876	Little Brown Bat	Female	0307080-13	Total Homologues	942.	µg/Kg	162.	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl2-BZ#8	2.86 U	µg/Kg	2.86	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl3-BZ#18	6.36 U	µg/Kg	6.36	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl3-BZ#31/#28	45.8 U	µg/Kg	45.8	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#44	4.36 U	µg/Kg	4.36	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#45	5.82 U	µg/Kg	5.82	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#47	25.8 J	µg/Kg	7.39	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#49	3.63 U	µg/Kg	3.63	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#52	6.82 J	µg/Kg	4.13	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#56	4.49 J	µg/Kg	2.16	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#66	22.3	µg/Kg	2.90	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#70	2.53 U	µg/Kg	2.53	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#74	14.2 J	µg/Kg	4.46	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#77	3.99 U	µg/Kg	3.99	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl4-BZ#81	3.96 U	µg/Kg	3.96	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#87	3.33 J	µg/Kg	1.76	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#95	1.36 U	µg/Kg	1.36	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#99	11.2	µg/Kg	.932	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#101	4.99 J	µg/Kg	2.66	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#105	6.49 J	µg/Kg	1.30	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#110	2.83 J	µg/Kg	2.53	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#114	1.16 U	µg/Kg	1.16	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#118	18.0 J	µg/Kg	2.73	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#123	2.30 U	µg/Kg	2.30	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl5-BZ#126	2.03 U	µg/Kg	2.03	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#128	1.83 U	µg/Kg	1.83	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#138	7.16 J	µg/Kg	2.63	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#146	1.66 U	µg/Kg	1.66	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#149	2.13 U	µg/Kg	2.13	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#151	3.53 U	µg/Kg	3.53	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#153	8.15 J	µg/Kg	3.03	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#156	2.16 U	µg/Kg	2.16	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#157	1.70 U	µg/Kg	1.70	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#158	1.43 U	µg/Kg	1.43	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#167	1.83 U	µg/Kg	1.83	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl6-BZ#169	5.89 U	µg/Kg	5.89	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl7-BZ#170	6.86 U	µg/Kg	6.86	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl7-BZ#174	1.40 U	µg/Kg	1.40	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl7-BZ#177	1.96 U	µg/Kg	1.96	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl7-BZ#180	2.93 U	µg/Kg	2.93	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl7-BZ#183	1.33 U	µg/Kg	1.33	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl7-BZ#189	2.63 U	µg/Kg	2.63	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl7-BZ#187	1.46 U	µg/Kg	1.46	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl8-BZ#194	3.06 U	µg/Kg	3.06	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl8-BZ#195	3.03 U	µg/Kg	3.03	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl8-BZ#201	2.66 U	µg/Kg	2.66	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl9-BZ#206	3.56 U	µg/Kg	3.56	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Cl10-BZ#209	1.66 U	µg/Kg	1.66	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Monochlorobiphenyls	97.9 U	µg/Kg	97.9	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Dichlorobiphenyls	2.86 U	µg/Kg	2.86	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Trichlorobiphenyls	19.0 J	µg/Kg	45.8	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Tetrachlorobiphenyls	107.	µg/Kg	7.39	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Pentachlorobiphenyls	122.	µg/Kg	2.73	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Hexachlorobiphenyls	26.0	µg/Kg	5.89	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Heptachlorobiphenyls	6.86 U	µg/Kg	6.86	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Octachlorobiphenyls	3.06 U	µg/Kg	3.06	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Nonachlorobiphenyls	3.56 U	µg/Kg	3.56	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Decachlorobiphenyl	1.66 U	µg/Kg	1.66	µg/Kg
7/25/2001	Sarato 19	613520	4761876	Little Brown Bat	Female	0307080-14	Total Homologues	274.	µg/Kg	97.9	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl2-BZ#8	3.66 U	µg/Kg	3.66	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl3-BZ#18	8.12 U	µg/Kg	8.12	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl3-BZ#31/#28	65.1 J	µg/Kg	58.6	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#44	5.57 U	µg/Kg	5.57	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#45	7.44 U	µg/Kg	7.44	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#47	248.	µg/Kg	9.44	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#49	4.64 U	µg/Kg	4.64	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#52	9.57 J	µg/Kg	5.27	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#56	37.6	µg/Kg	2.76	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#66	206.	µg/Kg	3.70	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#70	3.23 U	µg/Kg	3.23	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#74	153.	µg/Kg	5.70	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#77	5.10 U	µg/Kg	5.10	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl4-BZ#81	5.06 U	µg/Kg	5.06	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#87	43.2 J	µg/Kg	2.25	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#95	1.74 U	µg/Kg	1.74	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#99	174.	µg/Kg	1.19	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#101	11.9 J	µg/Kg	3.40	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#105	63.4	µg/Kg	1.66	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#110	3.23 U	µg/Kg	3.23	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#114	10.4	µg/Kg	1.49	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#118	250. J	µg/Kg	3.49	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#123	2.93 U	µg/Kg	2.93	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl5-BZ#126	2.59 U	µg/Kg	2.59	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#128	13.4	µg/Kg	2.34	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#138	137. J	µg/Kg	3.36	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#146	11.9	µg/Kg	2.13	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#149	2.72 U	µg/Kg	2.72	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#151	4.51 U	µg/Kg	4.51	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#153	214.	µg/Kg	3.87	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#156	14.7	µg/Kg	2.76	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#157	2.55 J	µg/Kg	2.17	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#158	1.83 U	µg/Kg	1.83	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#167	7.23 J	µg/Kg	2.34	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl6-BZ#169	7.53 U	µg/Kg	7.53	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl7-BZ#170	37.0 J	µg/Kg	8.76	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl7-BZ#174	1.79 U	µg/Kg	1.79	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl7-BZ#177	3.83 J	µg/Kg	2.51	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl7-BZ#180	54.0	µg/Kg	3.74	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl7-BZ#183	8.51	µg/Kg	1.70	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl7-BZ#189	3.36 U	µg/Kg	3.36	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl7-BZ#187	9.78	µg/Kg	1.87	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl8-BZ#194	16.6 J	µg/Kg	3.91	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl8-BZ#195	4.68 J	µg/Kg	3.87	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl8-BZ#201	6.38 J	µg/Kg	3.40	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl9-BZ#206	4.55 U	µg/Kg	4.55	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Cl10-BZ#209	2.13 U	µg/Kg	2.13	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Monochlorobiphenyls	125. U	µg/Kg	125.	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Dichlorobiphenyls	3.66 U	µg/Kg	3.66	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Trichlorobiphenyls	64.6	µg/Kg	58.6	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Tetrachlorobiphenyls	778.	µg/Kg	9.44	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Pentachlorobiphenyls	903.	µg/Kg	3.49	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Hexachlorobiphenyls	471.	µg/Kg	7.53	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Heptachlorobiphenyls	113.	µg/Kg	8.76	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Octachlorobiphenyls	56.6	µg/Kg	3.91	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Nonachlorobiphenyls	4.55 U	µg/Kg	4.55	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Decachlorobiphenyl	2.13 U	µg/Kg	2.13	µg/Kg
7/23/2001	Sarato 1	613520	4761876	Little Brown Bat	Male	0307080-15	Total Homologues	2390.	µg/Kg	125.	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl2-BZ#8	2.70 U	µg/Kg	2.70	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl3-BZ#18	6.01 U	µg/Kg	6.01	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl3-BZ#31/#28	76.1 J	µg/Kg	43.3	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#44	4.12 U	µg/Kg	4.12	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#45	5.50 U	µg/Kg	5.50	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#47	85.2	µg/Kg	6.98	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#49	5.03 J	µg/Kg	3.43	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#52	13.7 J	µg/Kg	3.90	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#56	18.7	µg/Kg	2.04	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#66	92.9	µg/Kg	2.74	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#70	7.39 J	µg/Kg	2.39	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#74	65.1	µg/Kg	4.21	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#77	3.77 U	µg/Kg	3.77	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl4-BZ#81	3.74 U	µg/Kg	3.74	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#87	13.4	µg/Kg	1.67	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#95	2.20 J	µg/Kg	1.29	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#99	106.	µg/Kg	.881	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#101	6.13 J	µg/Kg	2.52	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#105	26.6	µg/Kg	1.23	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#110	3.46 J	µg/Kg	2.39	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#114	8.65	µg/Kg	1.10	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#118	153.	µg/Kg	2.58	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#123	2.17 U	µg/Kg	2.17	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl5-BZ#126	1.92 U	µg/Kg	1.92	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#128	4.40 J	µg/Kg	1.73	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#138	76.1	µg/Kg	2.48	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#146	5.19 J	µg/Kg	1.57	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#149	2.01 U	µg/Kg	2.01	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#151	3.33 U	µg/Kg	3.33	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#153	256.	µg/Kg	2.86	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#156	23.1	µg/Kg	2.04	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#157	3.30 J	µg/Kg	1.60	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#158	2.20 J	µg/Kg	1.35	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#167	11.0	µg/Kg	1.73	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl6-BZ#169	5.57 U	µg/Kg	5.57	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl7-BZ#170	67.4	µg/Kg	6.48	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl7-BZ#174	1.32 U	µg/Kg	1.32	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl7-BZ#177	1.86 U	µg/Kg	1.86	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl7-BZ#180	107.	µg/Kg	2.77	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl7-BZ#183	15.4	µg/Kg	1.26	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl7-BZ#189	2.48 U	µg/Kg	2.48	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl7-BZ#187	7.23	µg/Kg	1.38	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl8-BZ#194	26.7	µg/Kg	2.89	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl8-BZ#195	7.08 J	µg/Kg	2.86	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl8-BZ#201	7.70 J	µg/Kg	2.52	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl9-BZ#206	10.7 J	µg/Kg	3.36	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Cl10-BZ#209	3.93 J	µg/Kg	1.57	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Monochlorobiphenyls	92.4 U	µg/Kg	92.4	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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J/UJ=Estimated result or detection limit

[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Dichlorobiphenyls	2.70 U	µg/Kg	2.70	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Trichlorobiphenyls	66.8	µg/Kg	43.3	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Tetrachlorobiphenyls	439.	µg/Kg	6.98	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Pentachlorobiphenyls	539.	µg/Kg	2.58	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Hexachlorobiphenyls	474.	µg/Kg	5.57	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Heptachlorobiphenyls	169.	µg/Kg	6.48	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Octachlorobiphenyls	58.0	µg/Kg	2.89	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Nonachlorobiphenyls	10.7 J	µg/Kg	3.36	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Decachlorobiphenyl	3.93 J	µg/Kg	1.57	µg/Kg
7/23/2001	Sarato 10	613520	4761876	Little Brown Bat	Male	0307081-01	Total Homologues	1760.	µg/Kg	92.4	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl2-BZ#8	1.21 U	µg/Kg	1.21	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl3-BZ#18	2.69 U	µg/Kg	2.69	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl3-BZ#31/#28	34.7 J	µg/Kg	19.4	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#44	1.84 U	µg/Kg	1.84	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#45	4.01 J	µg/Kg	2.46	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#47	16.0	µg/Kg	3.12	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#49	2.25 J	µg/Kg	1.53	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#52	4.57 J	µg/Kg	1.74	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#56	2.60 J	µg/Kg	.915	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#66	6.19	µg/Kg	1.22	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#70	3.94 J	µg/Kg	1.07	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#74	9.64	µg/Kg	1.89	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#77	1.69 U	µg/Kg	1.69	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl4-BZ#81	1.67 U	µg/Kg	1.67	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#87	3.52 J	µg/Kg	.746	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#95	.915 J	µg/Kg	.577	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#99	8.09	µg/Kg	.394	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#101	2.04 J	µg/Kg	1.13	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#105	3.94	µg/Kg	.549	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#110	1.07 U	µg/Kg	1.07	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#114	.493 U	µg/Kg	.493	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#118	14.2	µg/Kg	1.15	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#123	.971 U	µg/Kg	.971	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl5-BZ#126	.858 U	µg/Kg	.858	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#128	.844 J	µg/Kg	.774	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#138	10.1	µg/Kg	1.11	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#146	.704 U	µg/Kg	.704	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#149	.901 U	µg/Kg	.901	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#151	1.49 U	µg/Kg	1.49	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#153	10.8	µg/Kg	1.28	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#156	.985 J	µg/Kg	.915	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#157	.718 U	µg/Kg	.718	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#158	.704 J	µg/Kg	.605	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#167	.774 U	µg/Kg	.774	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl6-BZ#169	2.49 U	µg/Kg	2.49	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl7-BZ#170	2.90 U	µg/Kg	2.90	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl7-BZ#174	.591 U	µg/Kg	.591	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl7-BZ#177	.830 U	µg/Kg	.830	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl7-BZ#180	4.36 J	µg/Kg	1.24	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl7-BZ#183	.844 J	µg/Kg	.563	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl7-BZ#189	1.11 U	µg/Kg	1.11	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl7-BZ#187	1.13 J	µg/Kg	.619	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl8-BZ#194	2.74 J	µg/Kg	1.30	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl8-BZ#195	1.28 U	µg/Kg	1.28	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl8-BZ#201	1.13 U	µg/Kg	1.13	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl9-BZ#206	3.31 J	µg/Kg	1.51	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Cl10-BZ#209	.704 U	µg/Kg	.704	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Monochlorobiphenyls	41.4 U	µg/Kg	41.4	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Dichlorobiphenyls	1.21 U	µg/Kg	1.21	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Trichlorobiphenyls	14.4 J	µg/Kg	19.4	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Tetrachlorobiphenyls	88.6	µg/Kg	3.12	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Pentachlorobiphenyls	84.8	µg/Kg	1.15	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Hexachlorobiphenyls	40.9	µg/Kg	2.49	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Heptachlorobiphenyls	10.2	µg/Kg	2.90	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Octachlorobiphenyls	8.16	µg/Kg	1.30	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Nonachlorobiphenyls	3.31 J	µg/Kg	1.51	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Decachlorobiphenyl	.704 U	µg/Kg	.704	µg/Kg
7/26/2001	Sarato 28	613520	4761876	Big Brown Bat	Female	0307081-02	Total Homologues	250.	µg/Kg	41.4	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl2-BZ#8	5.67 U	µg/Kg	5.67	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl3-BZ#18	12.6 U	µg/Kg	12.6	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl3-BZ#31/#28	90.9 U	µg/Kg	90.9	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#44	8.64 U	µg/Kg	8.64	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#45	11.6 U	µg/Kg	11.6	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#47	14.6 U	µg/Kg	14.6	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#49	7.19 U	µg/Kg	7.19	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#52	8.18 U	µg/Kg	8.18	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#56	4.29 U	µg/Kg	4.29	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#66	5.74 U	µg/Kg	5.74	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#70	5.01 U	µg/Kg	5.01	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#74	8.84 U	µg/Kg	8.84	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#77	7.92 U	µg/Kg	7.92	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl4-BZ#81	7.85 U	µg/Kg	7.85	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#87	3.50 U	µg/Kg	3.50	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#95	2.70 U	µg/Kg	2.70	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#99	1.85 U	µg/Kg	1.85	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#101	5.28 U	µg/Kg	5.28	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#105	2.57 U	µg/Kg	2.57	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#110	5.01 U	µg/Kg	5.01	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#114	2.31 U	µg/Kg	2.31	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#118	7.26 J	µg/Kg	5.41	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#123	4.55 U	µg/Kg	4.55	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl5-BZ#126	4.03 U	µg/Kg	4.03	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#128	3.63 U	µg/Kg	3.63	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#138	9.57 J	µg/Kg	5.21	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#146	3.30 U	µg/Kg	3.30	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#149	4.22 U	µg/Kg	4.22	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#151	6.99 U	µg/Kg	6.99	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#153	10.6 J	µg/Kg	6.00	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#156	4.29 U	µg/Kg	4.29	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#157	3.36 U	µg/Kg	3.36	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#158	2.84 U	µg/Kg	2.84	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#167	3.63 U	µg/Kg	3.63	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl6-BZ#169	11.7 U	µg/Kg	11.7	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl7-BZ#170	13.6 U	µg/Kg	13.6	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl7-BZ#174	2.77 U	µg/Kg	2.77	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl7-BZ#177	3.89 U	µg/Kg	3.89	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl7-BZ#180	5.81 U	µg/Kg	5.81	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl7-BZ#183	2.64 U	µg/Kg	2.64	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl7-BZ#189	5.21 U	µg/Kg	5.21	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl7-BZ#187	2.90 U	µg/Kg	2.90	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl8-BZ#194	6.07 U	µg/Kg	6.07	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl8-BZ#195	6.00 U	µg/Kg	6.00	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl8-BZ#201	5.28 U	µg/Kg	5.28	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl9-BZ#206	7.06 U	µg/Kg	7.06	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Cl10-BZ#209	3.30 U	µg/Kg	3.30	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Monochlorobiphenyls	194. U	µg/Kg	194.	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Dichlorobiphenyls	5.67 U	µg/Kg	5.67	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Trichlorobiphenyls	31.3 J	µg/Kg	90.9	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Tetrachlorobiphenyls	88.7	µg/Kg	14.6	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Pentachlorobiphenyls	76.9	µg/Kg	5.41	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Hexachlorobiphenyls	36.6	µg/Kg	11.7	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Heptachlorobiphenyls	10.2 J	µg/Kg	13.6	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Octachlorobiphenyls	10.6 J	µg/Kg	6.07	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Nonachlorobiphenyls	7.06 U	µg/Kg	7.06	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Decachlorobiphenyl	3.30 U	µg/Kg	3.30	µg/Kg
7/16/2001	W Birch 13	569295	4712892	Little Brown Bat	Female	0307081-03	Total Homologues	254.	µg/Kg	194.	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl2-BZ#8	3.62 U	µg/Kg	3.62	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl3-BZ#18	8.03 U	µg/Kg	8.03	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl3-BZ#31/#28	57.9 U	µg/Kg	57.9	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#44	5.51 U	µg/Kg	5.51	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#45	7.36 U	µg/Kg	7.36	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#47	9.33 U	µg/Kg	9.33	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#49	4.58 U	µg/Kg	4.58	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#52	5.21 U	µg/Kg	5.21	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#56	2.73 U	µg/Kg	2.73	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#66	3.66 U	µg/Kg	3.66	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#70	3.20 U	µg/Kg	3.20	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#74	5.63 U	µg/Kg	5.63	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#77	5.04 U	µg/Kg	5.04	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl4-BZ#81	5.00 U	µg/Kg	5.00	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#87	2.23 U	µg/Kg	2.23	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#95	1.72 U	µg/Kg	1.72	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#99	1.18 U	µg/Kg	1.18	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#101	3.36 U	µg/Kg	3.36	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#105	1.64 U	µg/Kg	1.64	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#110	3.20 U	µg/Kg	3.20	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#114	1.47 U	µg/Kg	1.47	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#118	3.45 U	µg/Kg	3.45	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#123	2.90 U	µg/Kg	2.90	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl5-BZ#126	2.56 U	µg/Kg	2.56	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#128	2.31 U	µg/Kg	2.31	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#138	3.99 J	µg/Kg	3.32	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#146	2.10 U	µg/Kg	2.10	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#149	2.69 U	µg/Kg	2.69	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#151	4.46 U	µg/Kg	4.46	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#153	5.04 J	µg/Kg	3.83	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#156	2.73 U	µg/Kg	2.73	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#157	2.14 U	µg/Kg	2.14	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#158	1.81 U	µg/Kg	1.81	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#167	2.31 U	µg/Kg	2.31	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl6-BZ#169	7.44 U	µg/Kg	7.44	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl7-BZ#170	8.66 U	µg/Kg	8.66	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl7-BZ#174	1.77 U	µg/Kg	1.77	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl7-BZ#177	2.48 U	µg/Kg	2.48	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl7-BZ#180	6.10 J	µg/Kg	3.70	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl7-BZ#183	1.68 U	µg/Kg	1.68	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl7-BZ#189	3.32 U	µg/Kg	3.32	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl7-BZ#187	1.85 U	µg/Kg	1.85	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl8-BZ#194	3.87 U	µg/Kg	3.87	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl8-BZ#195	3.83 U	µg/Kg	3.83	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl8-BZ#201	3.36 U	µg/Kg	3.36	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl9-BZ#206	4.50 U	µg/Kg	4.50	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Cl10-BZ#209	2.10 U	µg/Kg	2.10	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Monochlorobiphenyls	124. U	µg/Kg	124.	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Dichlorobiphenyls	3.62 U	µg/Kg	3.62	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Trichlorobiphenyls	23.1 J	µg/Kg	57.9	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Tetrachlorobiphenyls	9.33 U	µg/Kg	9.33	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Pentachlorobiphenyls	46.5	µg/Kg	3.45	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Hexachlorobiphenyls	17.7	µg/Kg	7.44	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Heptachlorobiphenyls	13.9	µg/Kg	8.66	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Octachlorobiphenyls	10.9	µg/Kg	3.87	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Nonachlorobiphenyls	4.50 U	µg/Kg	4.50	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Decachlorobiphenyl	2.10 U	µg/Kg	2.10	µg/Kg
7/20/2001	Black R 7	470549	4836810	Little Brown Bat	Female	0307081-04	Total Homologues	112.	µg/Kg	124.	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl2-BZ#8	4.43 U	µg/Kg	4.43	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl3-BZ#18	9.85 U	µg/Kg	9.85	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl3-BZ#31/#28	71.0 U	µg/Kg	71.0	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#44	6.75 U	µg/Kg	6.75	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#45	9.02 U	µg/Kg	9.02	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#47	11.4 U	µg/Kg	11.4	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#49	5.62 U	µg/Kg	5.62	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#52	6.39 U	µg/Kg	6.39	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#56	3.35 U	µg/Kg	3.35	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#66	4.49 U	µg/Kg	4.49	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#70	3.92 U	µg/Kg	3.92	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#74	6.91 U	µg/Kg	6.91	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#77	6.19 U	µg/Kg	6.19	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl4-BZ#81	6.13 U	µg/Kg	6.13	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#87	2.73 U	µg/Kg	2.73	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#95	2.11 U	µg/Kg	2.11	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#99	1.44 U	µg/Kg	1.44	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#101	4.12 U	µg/Kg	4.12	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#105	2.01 U	µg/Kg	2.01	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#110	3.92 U	µg/Kg	3.92	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#114	1.80 U	µg/Kg	1.80	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#118	7.22 J	µg/Kg	4.23	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#123	3.56 U	µg/Kg	3.56	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl5-BZ#126	3.14 U	µg/Kg	3.14	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#128	2.84 U	µg/Kg	2.84	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#138	10.3 J	µg/Kg	4.07	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#146	2.58 U	µg/Kg	2.58	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#149	3.30 U	µg/Kg	3.30	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#151	5.46 U	µg/Kg	5.46	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#153	21.6 J	µg/Kg	4.69	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#156	3.35 U	µg/Kg	3.35	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#157	8.25 J	µg/Kg	2.63	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#158	2.22 U	µg/Kg	2.22	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#167	2.84 U	µg/Kg	2.84	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl6-BZ#169	9.12 U	µg/Kg	9.12	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl7-BZ#170	10.6 U	µg/Kg	10.6	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl7-BZ#174	2.16 U	µg/Kg	2.16	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl7-BZ#177	3.04 U	µg/Kg	3.04	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl7-BZ#180	21.6 J	µg/Kg	4.54	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl7-BZ#183	2.84 J	µg/Kg	2.06	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl7-BZ#189	4.07 U	µg/Kg	4.07	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl7-BZ#187	3.09 J	µg/Kg	2.27	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl8-BZ#194	5.67 J	µg/Kg	4.74	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl8-BZ#195	4.69 U	µg/Kg	4.69	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl8-BZ#201	4.12 U	µg/Kg	4.12	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl9-BZ#206	5.52 U	µg/Kg	5.52	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Cl10-BZ#209	30.2	µg/Kg	2.58	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Monochlorobiphenyls	152. U	µg/Kg	152.	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Dichlorobiphenyls	4.43 U	µg/Kg	4.43	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Trichlorobiphenyls	15.5 J	µg/Kg	71.0	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Tetrachlorobiphenyls	80.9	µg/Kg	11.4	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Pentachlorobiphenyls	382.	µg/Kg	4.23	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Hexachlorobiphenyls	81.7	µg/Kg	9.12	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Heptachlorobiphenyls	35.6	µg/Kg	10.6	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Octachlorobiphenyls	36.9	µg/Kg	4.74	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Nonachlorobiphenyls	5.52 U	µg/Kg	5.52	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Decachlorobiphenyl	30.2	µg/Kg	2.58	µg/Kg
7/21/2001	Black R 15	470549	4836810	Little Brown Bat	Female	0307081-05	Total Homologues	662.	µg/Kg	152.	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl2-BZ#8	3.38 U	µg/Kg	3.38	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl3-BZ#18	7.50 U	µg/Kg	7.50	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl3-BZ#31/#28	54.1 U	µg/Kg	54.1	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#44	5.14 U	µg/Kg	5.14	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#45	6.87 U	µg/Kg	6.87	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#47	40.4 J	µg/Kg	8.72	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#49	9.23 J	µg/Kg	4.28	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#52	26.9	µg/Kg	4.87	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#56	12.0 J	µg/Kg	2.55	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#66	33.6	µg/Kg	3.42	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#70	12.4 J	µg/Kg	2.98	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#74	25.5 J	µg/Kg	5.26	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#77	4.71 U	µg/Kg	4.71	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl4-BZ#81	4.67 U	µg/Kg	4.67	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#87	8.84 J	µg/Kg	2.08	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#95	3.14 J	µg/Kg	1.61	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#99	17.3	µg/Kg	1.10	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#101	9.82 J	µg/Kg	3.14	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#105	7.46 J	µg/Kg	1.53	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#110	4.91 J	µg/Kg	2.98	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#114	1.38 U	µg/Kg	1.38	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#118	29.1	µg/Kg	3.22	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#123	2.71 U	µg/Kg	2.71	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl5-BZ#126	2.40 U	µg/Kg	2.40	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#128	3.34 J	µg/Kg	2.16	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#138	19.4	µg/Kg	3.10	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#146	3.14 J	µg/Kg	1.96	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#149	2.51 U	µg/Kg	2.51	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#151	4.16 U	µg/Kg	4.16	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#153	18.5	µg/Kg	3.57	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#156	2.55 U	µg/Kg	2.55	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#157	2.00 U	µg/Kg	2.00	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#158	1.69 U	µg/Kg	1.69	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#167	2.16 U	µg/Kg	2.16	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl6-BZ#169	6.95 U	µg/Kg	6.95	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl7-BZ#170	8.09 U	µg/Kg	8.09	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl7-BZ#174	1.65 U	µg/Kg	1.65	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl7-BZ#177	2.32 U	µg/Kg	2.32	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl7-BZ#180	8.05 J	µg/Kg	3.46	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl7-BZ#183	1.57 U	µg/Kg	1.57	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl7-BZ#189	3.10 U	µg/Kg	3.10	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl7-BZ#187	3.73 J	µg/Kg	1.73	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl8-BZ#194	5.30 J	µg/Kg	3.61	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl8-BZ#195	3.57 U	µg/Kg	3.57	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl8-BZ#201	3.14 J	µg/Kg	3.14	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl9-BZ#206	7.86 J	µg/Kg	4.20	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Cl10-BZ#209	5.30 J	µg/Kg	1.96	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Monochlorobiphenyls	116. U	µg/Kg	116.	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Dichlorobiphenyls	3.38 U	µg/Kg	3.38	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Trichlorobiphenyls	44.4 J	µg/Kg	54.1	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Tetrachlorobiphenyls	254.	µg/Kg	8.72	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Pentachlorobiphenyls	215.	µg/Kg	3.22	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Hexachlorobiphenyls	66.8	µg/Kg	6.95	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Heptachlorobiphenyls	20.2	µg/Kg	8.09	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Octachlorobiphenyls	18.5	µg/Kg	3.61	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Nonachlorobiphenyls	19.6 J	µg/Kg	4.20	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Decachlorobiphenyl	5.30 J	µg/Kg	1.96	µg/Kg
7/24/2001	Sarato 14	613520	4761876	Little Brown Bat	Female	0307081-06	Total Homologues	644.	µg/Kg	116.	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl2-BZ#8	2.26 U	µg/Kg	2.26	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl3-BZ#18	5.02 U	µg/Kg	5.02	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl3-BZ#31/#28	45.2 J	µg/Kg	36.2	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#44	3.44 U	µg/Kg	3.44	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#45	4.73 J	µg/Kg	4.60	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#47	47.6	µg/Kg	5.83	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#49	3.55 J	µg/Kg	2.86	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#52	24.8	µg/Kg	3.26	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#56	10.4	µg/Kg	1.71	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#66	37.4	µg/Kg	2.29	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#70	9.20 J	µg/Kg	2.00	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#74	27.2	µg/Kg	3.52	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#77	3.15 U	µg/Kg	3.15	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl4-BZ#81	3.13 U	µg/Kg	3.13	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#87	8.54	µg/Kg	1.39	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#95	1.08 U	µg/Kg	1.08	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#99	20.8	µg/Kg	.736	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#101	8.93 J	µg/Kg	2.10	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#105	8.41	µg/Kg	1.02	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#110	3.02 J	µg/Kg	2.00	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#114	1.58 J	µg/Kg	.919	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#118	33.8	µg/Kg	2.15	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#123	1.81 U	µg/Kg	1.81	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl5-BZ#126	1.60 U	µg/Kg	1.60	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#128	3.68 J	µg/Kg	1.44	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#138	24.2	µg/Kg	2.08	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#146	3.42 J	µg/Kg	1.31	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#149	1.97 J	µg/Kg	1.68	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#151	2.78 U	µg/Kg	2.78	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#153	24.4	µg/Kg	2.39	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#156	1.84 J	µg/Kg	1.71	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#157	1.34 U	µg/Kg	1.34	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#158	1.13 U	µg/Kg	1.13	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#167	1.44 U	µg/Kg	1.44	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl6-BZ#169	4.65 U	µg/Kg	4.65	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl7-BZ#170	6.17 J	µg/Kg	5.41	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl7-BZ#174	1.10 U	µg/Kg	1.10	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl7-BZ#177	1.55 U	µg/Kg	1.55	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl7-BZ#180	10.8 J	µg/Kg	2.31	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl7-BZ#183	2.10 J	µg/Kg	1.05	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl7-BZ#189	2.08 U	µg/Kg	2.08	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl7-BZ#187	4.20 J	µg/Kg	1.16	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl8-BZ#194	5.25 J	µg/Kg	2.42	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl8-BZ#195	2.39 U	µg/Kg	2.39	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl8-BZ#201	3.15 J	µg/Kg	2.10	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl9-BZ#206	6.83 J	µg/Kg	2.81	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Cl10-BZ#209	4.99 J	µg/Kg	1.31	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Monochlorobiphenyls	77.2 U	µg/Kg	77.2	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Dichlorobiphenyls	2.26 U	µg/Kg	2.26	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Trichlorobiphenyls	37.4 J	µg/Kg	36.2	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Tetrachlorobiphenyls	219.	µg/Kg	5.83	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Pentachlorobiphenyls	164.	µg/Kg	2.15	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Hexachlorobiphenyls	80.6	µg/Kg	4.65	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Heptachlorobiphenyls	23.0	µg/Kg	5.41	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Octachlorobiphenyls	13.9	µg/Kg	2.42	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Nonachlorobiphenyls	14.4	µg/Kg	2.81	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Decachlorobiphenyl	4.99 J	µg/Kg	1.31	µg/Kg
7/25/2001	Sarato 17	613520	4761876	Little Brown Bat	Female	0307081-07	Total Homologues	557.	µg/Kg	77.2	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl2-BZ#8	3.34 U	µg/Kg	3.34	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl3-BZ#18	7.43 U	µg/Kg	7.43	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl3-BZ#31/#28	53.6 U	µg/Kg	53.6	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#44	5.10 U	µg/Kg	5.10	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#45	6.81 U	µg/Kg	6.81	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#47	8.64 U	µg/Kg	8.64	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#49	4.24 U	µg/Kg	4.24	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#52	4.82 U	µg/Kg	4.82	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#56	4.08 J	µg/Kg	2.53	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#66	3.38 U	µg/Kg	3.38	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#70	2.96 U	µg/Kg	2.96	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#74	5.21 U	µg/Kg	5.21	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#77	4.67 U	µg/Kg	4.67	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl4-BZ#81	4.63 U	µg/Kg	4.63	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#87	2.06 U	µg/Kg	2.06	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#95	1.60 U	µg/Kg	1.60	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#99	1.09 U	µg/Kg	1.09	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#101	3.11 U	µg/Kg	3.11	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#105	1.52 U	µg/Kg	1.52	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#110	2.96 U	µg/Kg	2.96	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#114	1.36 U	µg/Kg	1.36	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#118	4.28 J	µg/Kg	3.19	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#123	2.68 U	µg/Kg	2.68	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl5-BZ#126	2.37 U	µg/Kg	2.37	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#128	2.14 U	µg/Kg	2.14	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#138	4.28 J	µg/Kg	3.07	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#146	1.94 U	µg/Kg	1.94	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#149	2.49 U	µg/Kg	2.49	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#151	4.12 U	µg/Kg	4.12	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#153	4.67 J	µg/Kg	3.54	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#156	2.53 U	µg/Kg	2.53	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#157	1.98 U	µg/Kg	1.98	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#158	1.67 U	µg/Kg	1.67	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#167	2.14 U	µg/Kg	2.14	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl6-BZ#169	6.88 U	µg/Kg	6.88	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl7-BZ#170	8.01 U	µg/Kg	8.01	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl7-BZ#174	1.63 U	µg/Kg	1.63	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl7-BZ#177	2.30 U	µg/Kg	2.30	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl7-BZ#180	3.42 U	µg/Kg	3.42	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl7-BZ#183	1.56 U	µg/Kg	1.56	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl7-BZ#189	3.07 U	µg/Kg	3.07	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl7-BZ#187	1.71 U	µg/Kg	1.71	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl8-BZ#194	3.58 U	µg/Kg	3.58	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl8-BZ#195	3.54 U	µg/Kg	3.54	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl8-BZ#201	3.11 U	µg/Kg	3.11	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl9-BZ#206	4.16 U	µg/Kg	4.16	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Cl10-BZ#209	4.67 J	µg/Kg	1.94	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Monochlorobiphenyls	114. U	µg/Kg	114.	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Dichlorobiphenyls	3.34 U	µg/Kg	3.34	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Trichlorobiphenyls	53.6 U	µg/Kg	53.6	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Tetrachlorobiphenyls	44.3	µg/Kg	8.64	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Pentachlorobiphenyls	69.4	µg/Kg	3.19	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Hexachlorobiphenyls	24.9	µg/Kg	6.88	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Heptachlorobiphenyls	10.1	µg/Kg	8.01	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Octachlorobiphenyls	6.03 J	µg/Kg	3.58	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Nonachlorobiphenyls	4.16 U	µg/Kg	4.16	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Decachlorobiphenyl	4.67 J	µg/Kg	1.94	µg/Kg
7/15/2001	W Birch 2	569295	4712892	Little Brown Bat	Female	0307081-08	Total Homologues	167.	µg/Kg	114.	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl2-BZ#8	2.65 U	µg/Kg	2.65	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl3-BZ#18	5.88 U	µg/Kg	5.88	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl3-BZ#31/#28	42.4 U	µg/Kg	42.4	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#44	4.03 U	µg/Kg	4.03	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#45	5.38 U	µg/Kg	5.38	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#47	6.83 U	µg/Kg	6.83	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#49	3.35 U	µg/Kg	3.35	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#52	3.82 U	µg/Kg	3.82	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#56	2.00 U	µg/Kg	2.00	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#66	2.68 U	µg/Kg	2.68	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#70	2.34 U	µg/Kg	2.34	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#74	4.12 U	µg/Kg	4.12	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#77	3.69 U	µg/Kg	3.69	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl4-BZ#81	3.66 U	µg/Kg	3.66	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#87	1.63 U	µg/Kg	1.63	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#95	1.26 U	µg/Kg	1.26	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#99	1.69 J	µg/Kg	.862	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#101	2.46 U	µg/Kg	2.46	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#105	1.69 J	µg/Kg	1.20	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#110	2.34 U	µg/Kg	2.34	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#114	1.08 U	µg/Kg	1.08	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#118	4.92 J	µg/Kg	2.52	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#123	2.12 U	µg/Kg	2.12	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl5-BZ#126	1.88 U	µg/Kg	1.88	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#128	1.69 U	µg/Kg	1.69	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#138	5.69 J	µg/Kg	2.43	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#146	1.54 U	µg/Kg	1.54	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#149	1.97 U	µg/Kg	1.97	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#151	3.26 U	µg/Kg	3.26	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#153	8.92 J	µg/Kg	2.80	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#156	2.00 U	µg/Kg	2.00	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#157	1.57 U	µg/Kg	1.57	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#158	1.32 U	µg/Kg	1.32	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#167	1.69 U	µg/Kg	1.69	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl6-BZ#169	5.45 U	µg/Kg	5.45	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl7-BZ#170	6.34 U	µg/Kg	6.34	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl7-BZ#174	1.29 U	µg/Kg	1.29	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl7-BZ#177	1.82 U	µg/Kg	1.82	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl7-BZ#180	5.08 J	µg/Kg	2.71	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl7-BZ#183	1.23 U	µg/Kg	1.23	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl7-BZ#189	2.43 U	µg/Kg	2.43	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl7-BZ#187	1.35 U	µg/Kg	1.35	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl8-BZ#194	2.83 U	µg/Kg	2.83	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl8-BZ#195	2.80 U	µg/Kg	2.80	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl8-BZ#201	2.46 U	µg/Kg	2.46	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl9-BZ#206	3.29 U	µg/Kg	3.29	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Cl10-BZ#209	1.54 U	µg/Kg	1.54	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Monochlorobiphenyls	90.5 U	µg/Kg	90.5	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Dichlorobiphenyls	2.65 U	µg/Kg	2.65	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Trichlorobiphenyls	13.5 J	µg/Kg	42.4	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Tetrachlorobiphenyls	43.7	µg/Kg	6.83	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Pentachlorobiphenyls	46.6	µg/Kg	2.52	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Hexachlorobiphenyls	22.8	µg/Kg	5.45	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Heptachlorobiphenyls	8.62	µg/Kg	6.34	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Octachlorobiphenyls	7.08	µg/Kg	2.83	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Nonachlorobiphenyls	3.29 U	µg/Kg	3.29	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Decachlorobiphenyl	1.54 U	µg/Kg	1.54	µg/Kg
7/16/2001	W Birch 12	569295	4712892	Little Brown Bat	Female	0307081-09	Total Homologues	142.	µg/Kg	90.5	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl2-BZ#8	2.39 U	µg/Kg	2.39	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl3-BZ#18	5.31 U	µg/Kg	5.31	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl3-BZ#31/#28	38.3 U	µg/Kg	38.3	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#44	3.64 U	µg/Kg	3.64	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#45	4.86 U	µg/Kg	4.86	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#47	6.17 U	µg/Kg	6.17	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#49	3.03 U	µg/Kg	3.03	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#52	3.44 U	µg/Kg	3.44	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#56	1.81 U	µg/Kg	1.81	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#66	2.42 U	µg/Kg	2.42	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#70	2.11 U	µg/Kg	2.11	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#74	3.72 U	µg/Kg	3.72	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#77	3.33 U	µg/Kg	3.33	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl4-BZ#81	3.31 U	µg/Kg	3.31	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#87	1.47 U	µg/Kg	1.47	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#95	1.14 U	µg/Kg	1.14	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#99	.778 U	µg/Kg	.778	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#101	2.22 U	µg/Kg	2.22	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#105	1.08 U	µg/Kg	1.08	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#110	2.11 U	µg/Kg	2.11	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#114	.973 U	µg/Kg	.973	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#118	2.50 J	µg/Kg	2.28	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#123	1.92 U	µg/Kg	1.92	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl5-BZ#126	1.70 U	µg/Kg	1.70	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#128	1.53 U	µg/Kg	1.53	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#138	2.50 J	µg/Kg	2.19	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#146	1.39 U	µg/Kg	1.39	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#149	1.78 U	µg/Kg	1.78	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#151	2.94 U	µg/Kg	2.94	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#153	2.53 U	µg/Kg	2.53	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#156	1.81 U	µg/Kg	1.81	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#157	1.42 U	µg/Kg	1.42	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#158	1.20 U	µg/Kg	1.20	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#167	1.53 U	µg/Kg	1.53	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl6-BZ#169	4.92 U	µg/Kg	4.92	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl7-BZ#170	5.72 U	µg/Kg	5.72	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl7-BZ#174	1.17 U	µg/Kg	1.17	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl7-BZ#177	1.64 U	µg/Kg	1.64	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl7-BZ#180	2.44 U	µg/Kg	2.44	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl7-BZ#183	1.11 U	µg/Kg	1.11	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl7-BZ#189	2.19 U	µg/Kg	2.19	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl7-BZ#187	1.22 U	µg/Kg	1.22	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl8-BZ#194	2.56 U	µg/Kg	2.56	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl8-BZ#195	2.53 U	µg/Kg	2.53	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl8-BZ#201	2.22 U	µg/Kg	2.22	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl9-BZ#206	2.97 U	µg/Kg	2.97	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Cl10-BZ#209	1.39 U	µg/Kg	1.39	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Monochlorobiphenyls	81.7 U	µg/Kg	81.7	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Dichlorobiphenyls	2.39 U	µg/Kg	2.39	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Trichlorobiphenyls	14.3 J	µg/Kg	38.3	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Tetrachlorobiphenyls	19.3	µg/Kg	6.17	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Pentachlorobiphenyls	22.6	µg/Kg	2.28	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Hexachlorobiphenyls	9.72 J	µg/Kg	4.92	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Heptachlorobiphenyls	5.42 J	µg/Kg	5.72	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Octachlorobiphenyls	3.61 J	µg/Kg	2.56	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Nonachlorobiphenyls	2.97 U	µg/Kg	2.97	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Decachlorobiphenyl	1.39 U	µg/Kg	1.39	µg/Kg
7/15/2001	W Birch 9	569295	4712892	Little Brown Bat	Female	0307081-10	Total Homologues	75.0 J	µg/Kg	81.7	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl2-BZ#8	2.16 U	µg/Kg	2.16	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl3-BZ#18	4.80 U	µg/Kg	4.80	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl3-BZ#31/#28	34.6 U	µg/Kg	34.6	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#44	3.29 U	µg/Kg	3.29	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#45	4.39 U	µg/Kg	4.39	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#47	5.57 U	µg/Kg	5.57	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#49	2.74 U	µg/Kg	2.74	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#52	3.11 U	µg/Kg	3.11	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#56	1.63 U	µg/Kg	1.63	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#66	2.18 U	µg/Kg	2.18	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#70	1.91 U	µg/Kg	1.91	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#74	3.36 U	µg/Kg	3.36	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

[2]U=Non-detected result at the detection limit
J/UJ=Estimated result or detection limit

[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]	DETECTION LIMIT[3]
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#77	3.01 U µg/Kg	3.01 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl4-BZ#81	2.99 U µg/Kg	2.99 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#87	1.33 U µg/Kg	1.33 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#95	1.03 U µg/Kg	1.03 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#99	.703 U µg/Kg	.703 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#101	2.01 U µg/Kg	2.01 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#105	.979 U µg/Kg	.979 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#110	1.91 U µg/Kg	1.91 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#114	.879 U µg/Kg	.879 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#118	4.52 J µg/Kg	2.06 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#123	1.73 U µg/Kg	1.73 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl5-BZ#126	1.53 U µg/Kg	1.53 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#128	1.38 U µg/Kg	1.38 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#138	5.27 J µg/Kg	1.98 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#146	1.25 U µg/Kg	1.25 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#149	1.61 U µg/Kg	1.61 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#151	2.66 U µg/Kg	2.66 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#153	9.66 J µg/Kg	2.28 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#156	1.63 U µg/Kg	1.63 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#157	1.28 U µg/Kg	1.28 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#158	1.08 U µg/Kg	1.08 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#167	1.38 U µg/Kg	1.38 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl6-BZ#169	4.44 U µg/Kg	4.44 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl7-BZ#170	5.17 U µg/Kg	5.17 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl7-BZ#174	1.05 U µg/Kg	1.05 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl7-BZ#177	1.48 U µg/Kg	1.48 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl7-BZ#180	6.90 J µg/Kg	2.21 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl7-BZ#183	1.00 U µg/Kg	1.00 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl7-BZ#189	1.98 U µg/Kg	1.98 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl7-BZ#187	1.38 J µg/Kg	1.10 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl8-BZ#194	2.31 U µg/Kg	2.31 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl8-BZ#195	2.28 U µg/Kg	2.28 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl8-BZ#201	2.01 U µg/Kg	2.01 µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl9-BZ#206	2.69 U µg/Kg	2.69 µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Cl10-BZ#209	1.25 U	µg/Kg	1.25	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Monochlorobiphenyls	73.8 U	µg/Kg	73.8	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Dichlorobiphenyls	2.16 U	µg/Kg	2.16	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Trichlorobiphenyls	17.8 J	µg/Kg	34.6	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Tetrachlorobiphenyls	14.7	µg/Kg	5.57	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Pentachlorobiphenyls	27.7	µg/Kg	2.06	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Hexachlorobiphenyls	23.0	µg/Kg	4.44	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Heptachlorobiphenyls	11.4	µg/Kg	5.17	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Octachlorobiphenyls	6.40	µg/Kg	2.31	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Nonachlorobiphenyls	2.69 U	µg/Kg	2.69	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Decachlorobiphenyl	1.25 U	µg/Kg	1.25	µg/Kg
7/16/2001	W Birch 10	569295	4712892	Little Brown Bat	Female	0307081-11	Total Homologues	102.	µg/Kg	73.8	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl2-BZ#8	2.09 U	µg/Kg	2.09	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl3-BZ#18	4.64 U	µg/Kg	4.64	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl3-BZ#31/#28	33.5 U	µg/Kg	33.5	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#44	3.18 U	µg/Kg	3.18	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#45	4.25 J	µg/Kg	4.25	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#47	5.39 U	µg/Kg	5.39	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#49	2.65 U	µg/Kg	2.65	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#52	3.01 U	µg/Kg	3.01	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#56	1.58 U	µg/Kg	1.58	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#66	2.11 U	µg/Kg	2.11	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#70	1.85 U	µg/Kg	1.85	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#74	3.26 U	µg/Kg	3.26	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#77	2.92 U	µg/Kg	2.92	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl4-BZ#81	2.89 U	µg/Kg	2.89	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#87	1.29 U	µg/Kg	1.29	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#95	4.74 J	µg/Kg	.996	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#99	.680 U	µg/Kg	.680	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#101	1.94 U	µg/Kg	1.94	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#105	.948 U	µg/Kg	.948	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#110	1.85 U	µg/Kg	1.85	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#114	.850 U	µg/Kg	.850	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#118	1.99 U	µg/Kg	1.99	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#123	1.68 U	µg/Kg	1.68	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl5-BZ#126	1.48 U	µg/Kg	1.48	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#128	1.34 U	µg/Kg	1.34	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#138	1.92 U	µg/Kg	1.92	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#146	1.22 U	µg/Kg	1.22	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#149	1.56 U	µg/Kg	1.56	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#151	2.58 U	µg/Kg	2.58	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#153	2.21 U	µg/Kg	2.21	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#156	1.58 U	µg/Kg	1.58	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#157	1.24 U	µg/Kg	1.24	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#158	1.04 U	µg/Kg	1.04	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#167	1.34 U	µg/Kg	1.34	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl6-BZ#169	4.30 U	µg/Kg	4.30	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl7-BZ#170	5.00 U	µg/Kg	5.00	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl7-BZ#174	1.02 U	µg/Kg	1.02	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl7-BZ#177	1.43 U	µg/Kg	1.43	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl7-BZ#180	2.14 U	µg/Kg	2.14	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl7-BZ#183	.972 U	µg/Kg	.972	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl7-BZ#189	1.92 U	µg/Kg	1.92	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl7-BZ#187	1.07 U	µg/Kg	1.07	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl8-BZ#194	2.24 U	µg/Kg	2.24	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl8-BZ#195	2.21 U	µg/Kg	2.21	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl8-BZ#201	1.94 U	µg/Kg	1.94	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl9-BZ#206	2.60 U	µg/Kg	2.60	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Cl10-BZ#209	1.22 U	µg/Kg	1.22	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Monochlorobiphenyls	71.4 U	µg/Kg	71.4	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Dichlorobiphenyls	2.09 U	µg/Kg	2.09	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Trichlorobiphenyls	7.90 J	µg/Kg	33.5	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Tetrachlorobiphenyls	35.2	µg/Kg	5.39	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Pentachlorobiphenyls	46.3	µg/Kg	1.99	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Hexachlorobiphenyls	10.1 J	µg/Kg	4.30	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Heptachlorobiphenyls	2.92 J	µg/Kg	5.00	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Octachlorobiphenyls	5.59	µg/Kg	2.24	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Nonachlorobiphenyls	2.60 U	µg/Kg	2.60	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number
[2]U=Non-detected result at the detection limit
J/UJ=Estimated result or detection limit
[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Decachlorobiphenyl	1.22 U	µg/Kg	1.22	µg/Kg
7/20/2001	Black R 5	470549	4836810	Little Brown Bat	Female	0307081-12	Total Homologues	108.	µg/Kg	71.4	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl2-BZ#8	2.72 U	µg/Kg	2.72	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl3-BZ#18	6.04 U	µg/Kg	6.04	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl3-BZ#31/#28	46.8 J	µg/Kg	43.5	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#44	4.14 U	µg/Kg	4.14	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#45	5.53 U	µg/Kg	5.53	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#47	7.02 U	µg/Kg	7.02	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#49	3.44 U	µg/Kg	3.44	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#52	3.92 U	µg/Kg	3.92	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#56	2.06 U	µg/Kg	2.06	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#66	2.75 U	µg/Kg	2.75	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#70	2.40 U	µg/Kg	2.40	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#74	4.24 U	µg/Kg	4.24	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#77	3.79 U	µg/Kg	3.79	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl4-BZ#81	3.76 U	µg/Kg	3.76	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#87	1.74 J	µg/Kg	1.68	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#95	1.30 U	µg/Kg	1.30	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#99	.885 U	µg/Kg	.885	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#101	2.53 U	µg/Kg	2.53	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#105	1.23 U	µg/Kg	1.23	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#110	2.40 U	µg/Kg	2.40	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#114	1.11 U	µg/Kg	1.11	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#118	2.59 U	µg/Kg	2.59	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#123	2.18 U	µg/Kg	2.18	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl5-BZ#126	1.93 U	µg/Kg	1.93	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#128	1.74 U	µg/Kg	1.74	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#138	2.50 U	µg/Kg	2.50	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#146	1.58 U	µg/Kg	1.58	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#149	2.02 U	µg/Kg	2.02	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#151	3.35 U	µg/Kg	3.35	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#153	2.88 U	µg/Kg	2.88	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#156	2.06 U	µg/Kg	2.06	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#157	1.61 U	µg/Kg	1.61	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#158	1.36 U	µg/Kg	1.36	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#167	1.74 U	µg/Kg	1.74	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl6-BZ#169	5.60 U	µg/Kg	5.60	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl7-BZ#170	6.51 U	µg/Kg	6.51	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl7-BZ#174	1.33 U	µg/Kg	1.33	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl7-BZ#177	1.86 U	µg/Kg	1.86	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl7-BZ#180	2.78 U	µg/Kg	2.78	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl7-BZ#183	1.26 U	µg/Kg	1.26	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl7-BZ#189	2.50 U	µg/Kg	2.50	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl7-BZ#187	1.39 U	µg/Kg	1.39	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl8-BZ#194	2.91 U	µg/Kg	2.91	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl8-BZ#195	2.88 U	µg/Kg	2.88	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl8-BZ#201	2.53 U	µg/Kg	2.53	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl9-BZ#206	3.38 U	µg/Kg	3.38	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Cl10-BZ#209	4.58 J	µg/Kg	1.58	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Monochlorobiphenyls	92.9 U	µg/Kg	92.9	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Dichlorobiphenyls	2.72 U	µg/Kg	2.72	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Trichlorobiphenyls	9.80 J	µg/Kg	43.5	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Tetrachlorobiphenyls	41.1	µg/Kg	7.02	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Pentachlorobiphenyls	72.7	µg/Kg	2.59	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Hexachlorobiphenyls	17.1	µg/Kg	5.60	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Heptachlorobiphenyls	8.38	µg/Kg	6.51	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Octachlorobiphenyls	8.38	µg/Kg	2.91	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Nonachlorobiphenyls	5.85 J	µg/Kg	3.38	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Decachlorobiphenyl	4.58 J	µg/Kg	1.58	µg/Kg
7/15/2001	W Birch 4	569295	4712892	Little Brown Bat	Female	0307081-13	Total Homologues	168.	µg/Kg	92.9	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl2-BZ#8	2.11 U	µg/Kg	2.11	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl3-BZ#18	4.69 U	µg/Kg	4.69	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl3-BZ#31/#28	67.5 J	µg/Kg	33.8	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#44	3.22 U	µg/Kg	3.22	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#45	4.30 U	µg/Kg	4.30	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#47	70.6	µg/Kg	5.45	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#49	2.68 U	µg/Kg	2.68	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#52	13.6 J	µg/Kg	3.04	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#56	11.0	µg/Kg	1.60	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#66	51.9	µg/Kg	2.13	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#70	4.05 J	µg/Kg	1.86	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#74	36.3	µg/Kg	3.29	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#77	2.94 U	µg/Kg	2.94	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl4-BZ#81	2.92 U	µg/Kg	2.92	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#87	9.08	µg/Kg	1.30	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#95	1.47 J	µg/Kg	1.01	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#99	24.4	µg/Kg	.687	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#101	5.28 J	µg/Kg	1.96	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#105	10.4	µg/Kg	.957	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#110	1.96 J	µg/Kg	1.86	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#114	1.72 J	µg/Kg	.859	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#118	36.9	µg/Kg	2.01	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#123	1.69 U	µg/Kg	1.69	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl5-BZ#126	1.50 U	µg/Kg	1.50	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#128	4.30 J	µg/Kg	1.35	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#138	22.3	µg/Kg	1.94	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#146	3.19 J	µg/Kg	1.23	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#149	1.57 U	µg/Kg	1.57	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#151	2.60 U	µg/Kg	2.60	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#153	22.5	µg/Kg	2.23	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#156	2.33 J	µg/Kg	1.60	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#157	1.25 U	µg/Kg	1.25	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#158	1.35 J	µg/Kg	1.06	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#167	1.60 J	µg/Kg	1.35	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl6-BZ#169	4.34 U	µg/Kg	4.34	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl7-BZ#170	5.89 J	µg/Kg	5.06	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl7-BZ#174	1.03 U	µg/Kg	1.03	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl7-BZ#177	1.72 J	µg/Kg	1.45	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl7-BZ#180	8.35 J	µg/Kg	2.16	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl7-BZ#183	2.21 J	µg/Kg	.982	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl7-BZ#189	1.94 U	µg/Kg	1.94	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl7-BZ#187	4.05 J	µg/Kg	1.08	µg/Kg

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl8-BZ#194	5.03 J	µg/Kg	2.26	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl8-BZ#195	2.23 U	µg/Kg	2.23	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl8-BZ#201	3.19 J	µg/Kg	1.96	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl9-BZ#206	6.50 J	µg/Kg	2.63	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Cl10-BZ#209	5.15 J	µg/Kg	1.23	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Monochlorobiphenyls	72.2 U	µg/Kg	72.2	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Dichlorobiphenyls	2.11 U	µg/Kg	2.11	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Trichlorobiphenyls	53.9	µg/Kg	33.8	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Tetrachlorobiphenyls	279.	µg/Kg	5.45	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Pentachlorobiphenyls	168.	µg/Kg	2.01	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Hexachlorobiphenyls	81.2	µg/Kg	4.34	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Heptachlorobiphenyls	21.2	µg/Kg	5.06	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Octachlorobiphenyls	18.0	µg/Kg	2.26	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Nonachlorobiphenyls	13.4	µg/Kg	2.63	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Decachlorobiphenyl	5.15 J	µg/Kg	1.23	µg/Kg
7/25/2001	Sarato 18	613520	4761876	Little Brown Bat	Female	0307081-14	Total Homologues	640.	µg/Kg	72.2	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl2-BZ#8	2.33 U	µg/Kg	2.33	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl3-BZ#18	5.17 U	µg/Kg	5.17	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl3-BZ#31/#28	37.3 U	µg/Kg	37.3	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#44	3.55 U	µg/Kg	3.55	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#45	6.23 J	µg/Kg	4.74	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#47	6.01 U	µg/Kg	6.01	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#49	2.95 U	µg/Kg	2.95	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#52	3.36 U	µg/Kg	3.36	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#56	1.76 U	µg/Kg	1.76	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#66	2.36 U	µg/Kg	2.36	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#70	2.06 U	µg/Kg	2.06	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#74	3.63 U	µg/Kg	3.63	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#77	3.25 U	µg/Kg	3.25	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl4-BZ#81	3.22 U	µg/Kg	3.22	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#87	1.44 U	µg/Kg	1.44	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#95	1.11 U	µg/Kg	1.11	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#99	.758 U	µg/Kg	.758	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#101	2.17 U	µg/Kg	2.17	µg/Kg

[1]BZ#=PCB congener Ballschmiter & Zell number

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[3]PCB results & detection limit reported on wet weight basis

SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#105	1.06 U	µg/Kg	1.06	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#110	2.06 U	µg/Kg	2.06	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#114	.947 U	µg/Kg	.947	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#118	2.57 J	µg/Kg	2.22	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#123	1.87 U	µg/Kg	1.87	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl5-BZ#126	1.65 U	µg/Kg	1.65	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#128	1.49 U	µg/Kg	1.49	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#138	2.57 J	µg/Kg	2.14	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#146	1.35 U	µg/Kg	1.35	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#149	1.73 U	µg/Kg	1.73	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#151	2.87 U	µg/Kg	2.87	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#153	2.98 J	µg/Kg	2.46	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#156	1.76 U	µg/Kg	1.76	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#157	1.38 U	µg/Kg	1.38	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#158	1.16 U	µg/Kg	1.16	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#167	1.49 U	µg/Kg	1.49	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl6-BZ#169	4.79 U	µg/Kg	4.79	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl7-BZ#170	5.58 U	µg/Kg	5.58	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl7-BZ#174	1.14 U	µg/Kg	1.14	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl7-BZ#177	1.60 U	µg/Kg	1.60	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl7-BZ#180	2.71 J	µg/Kg	2.38	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl7-BZ#183	1.08 U	µg/Kg	1.08	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl7-BZ#189	2.14 U	µg/Kg	2.14	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl7-BZ#187	1.19 U	µg/Kg	1.19	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl8-BZ#194	2.49 U	µg/Kg	2.49	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl8-BZ#195	2.46 U	µg/Kg	2.46	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl8-BZ#201	2.17 U	µg/Kg	2.17	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl9-BZ#206	2.90 U	µg/Kg	2.90	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Cl10-BZ#209	7.31	µg/Kg	1.35	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Monochlorobiphenyls	79.6 U	µg/Kg	79.6	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Dichlorobiphenyls	2.33 U	µg/Kg	2.33	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Trichlorobiphenyls	16.6 J	µg/Kg	37.3	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Tetrachlorobiphenyls	22.3	µg/Kg	6.01	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Pentachlorobiphenyls	24.6	µg/Kg	2.22	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Hexachlorobiphenyls	14.2	µg/Kg	4.79	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Heptachlorobiphenyls	8.26	µg/Kg	5.58	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Octachlorobiphenyls	7.44	µg/Kg	2.49	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Nonachlorobiphenyls	5.14 J	µg/Kg	2.90	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Decachlorobiphenyl	7.31	µg/Kg	1.35	µg/Kg
7/20/2001	Black R 6	470549	4836810	Little Brown Bat	Female	0307081-15	Total Homologues	106.	µg/Kg	79.6	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl2-BZ#8	1.42 U	µg/Kg	1.42	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl3-BZ#18	3.15 U	µg/Kg	3.15	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl3-BZ#31/#28	24.4 J	µg/Kg	22.7	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#44	2.16 U	µg/Kg	2.16	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#45	2.89 U	µg/Kg	2.89	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#47	39.8	µg/Kg	3.66	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#49	1.80 U	µg/Kg	1.80	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#52	2.04 U	µg/Kg	2.04	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#56	2.80 J	µg/Kg	1.07	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#66	5.69 J	µg/Kg	1.43	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#70	2.14 J	µg/Kg	1.25	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#74	15.6	µg/Kg	2.21	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#77	1.98 U	µg/Kg	1.98	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl4-BZ#81	1.96 U	µg/Kg	1.96	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#87	1.65 J	µg/Kg	.874	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#95	1.15 J	µg/Kg	.676	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#99	46.1	µg/Kg	.462	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#101	3.96 J	µg/Kg	1.32	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#105	11.5	µg/Kg	.643	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#110	2.06 J	µg/Kg	1.25	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#114	1.57 J	µg/Kg	.577	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#118	41.6	µg/Kg	1.35	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#123	1.14 U	µg/Kg	1.14	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl5-BZ#126	1.01 U	µg/Kg	1.01	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#128	2.64 J	µg/Kg	.907	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#138	57.7	µg/Kg	1.30	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#146	.989 J	µg/Kg	.825	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#149	2.47 J	µg/Kg	1.06	µg/Kg

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SAMPLING DATE	FIELD ID	EASTING (NAD 83 UTM18N)	NORTHING (NAD 83 UTM18N)	COMMON NAME	Sex	LAB ID	ANALYTE	VALUE, INTERPRETIVE QUALIFIER[2] AND UNITS[3]		DETECTION LIMIT[3]	
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#151	1.75 U	µg/Kg	1.75	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#153	90.5	µg/Kg	1.50	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#156	6.35	µg/Kg	1.07	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#157	1.81 J	µg/Kg	.841	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#158	2.31 J	µg/Kg	.709	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#167	.989 J	µg/Kg	.907	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl6-BZ#169	2.92 U	µg/Kg	2.92	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl7-BZ#170	22.6	µg/Kg	3.40	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl7-BZ#174	.692 U	µg/Kg	.692	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl7-BZ#177	.973 U	µg/Kg	.973	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl7-BZ#180	39.2	µg/Kg	1.45	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl7-BZ#183	7.67	µg/Kg	.660	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl7-BZ#189	1.90 J	µg/Kg	1.30	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl7-BZ#187	2.31 J	µg/Kg	.725	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl8-BZ#194	12.9	µg/Kg	1.52	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl8-BZ#195	3.30 J	µg/Kg	1.50	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl8-BZ#201	1.32 U	µg/Kg	1.32	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl9-BZ#206	8.24 J	µg/Kg	1.76	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Cl10-BZ#209	1.32 J	µg/Kg	.825	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Monochlorobiphenyls	48.5 U	µg/Kg	48.5	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Dichlorobiphenyls	1.42 U	µg/Kg	1.42	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Trichlorobiphenyls	16.6 J	µg/Kg	22.7	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Tetrachlorobiphenyls	108.	µg/Kg	3.66	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Pentachlorobiphenyls	208.	µg/Kg	1.35	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Hexachlorobiphenyls	211.	µg/Kg	2.92	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Heptachlorobiphenyls	58.2	µg/Kg	3.40	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Octachlorobiphenyls	24.2	µg/Kg	1.52	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Nonachlorobiphenyls	15.0	µg/Kg	1.76	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Decachlorobiphenyl	1.32 J	µg/Kg	.825	µg/Kg
7/25/2001	Sarato 23	613520	4761876	Big Brown Bat	Female	0307081-16	Total Homologues	642.	µg/Kg	48.5	µg/Kg

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U.S. DEPARTMENT OF THE INTERIOR
MARCH 3, 1849

NATIONAL OCEANIC AND ATMOSPHERIC ADMINISTRATION
NOAA
U.S. DEPARTMENT OF COMMERCE

DEPARTMENT OF ENVIRONMENTAL CONSERVATION
NEW YORK STATE